

Steno 2 study 20 year follow-up — end of 2014

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Chapter 1

Data

1.1 Clinical visits

This file has data on the clinical visits that the trial participants have attended.

First we read the data and strip the irrelevant attributes:

```
> library( Epi )
> library( foreign )
> library(readstata13)
> clear()
> mic <- read.dta13( "../data/micro2.dta", nonint.factors=TRUE )
```

We now construct a groomed base analysis data set for the clinical visits — mainly the microvascular outcomes:

```
> ( na <- names( attributes(mic) ) )
 [1] "row.names"      "names"          "datalabel"      "time.stamp"
 [5] "formats"        "types"          "val.labels"     "var.labels"
 [9] "version"        "label.table"    "expansion.fields" "str1"
[13] "byteorder"      "class"
> wh <- match( c("names","row.names","class","var.labels"), na )
> attributes( mic )[-wh] <- NULL
> dn <- grep( "date", names(mic) )
> dn <- dn[c(1,4,5,6,10,11,13,15)]
> # A few things that need to be corrected
> str( mic[,dn] )
'data.frame':      748 obs. of  8 variables:
 $ date      : Date, format: "1993-05-11" "1995-02-28" ...
 $ date_birth : Date, format: "1932-04-20" "1932-04-20" ...
 $ date_blind : Date, format: NA NA ...
 $ date_death : Date, format: NA NA ...
 $ date_end   : Date, format: "2014-10-12" "2014-10-12" ...
 $ date_esrd  : Date, format: NA NA ...
 $ date_photocoag: num  NA NA NA NA NA ...
 $ dateeye2014 : Date, format: "2014-10-21" "2014-10-21" ...
> levels( mic$eds_r)
 [1] "No retinopathy"          "Minimal non-proliferative" "Moderate nonproliferative"
 [4] "Pre-proliferative"      "Photocoagulation"         "Proliferative"
> levels( mic$eds_l)
 [1] "No retinopathy"          "Minimal non-proliferative" "Moderate nonproliferative"
 [4] "Pre-proliferative"      "Photocoagulation"         "Proliferative"
> # transform.data.frame purges attributes so save them and put them back
> mat <- attributes(mic)
> mic <- transform( mic, date_photocoag = as.Date(date_photocoag,origin="1960-01-01"),
+                   eds_r = Relevel( eds_r, c(1:4,6:5) ),
+                   eds_l = Relevel( eds_l, c(1:4,6:5) ) )
> mat -> attributes(mic)
> names( mic )[dn]
```

```
[1] "date"           "date_birth"    "date_blind"    "date_death"    "date_end"
[6] "date_esrd"     "date_photocoag" "dateeye2014"
```

```
> nnam <- c("doVis",
+          "doBth",
+          "doBlind",
+          "doDth",
+          "doEnd",
+          "doESRD",
+          "doPhC",
+          "doEye")
> cbind( names( mic )[dn], nnam )
```

```
          nnam
[1,] "date"           "doVis"
[2,] "date_birth"    "doBth"
[3,] "date_blind"    "doBlind"
[4,] "date_death"    "doDth"
[5,] "date_end"      "doEnd"
[6,] "date_esrd"     "doESRD"
[7,] "date_photocoag" "doPhC"
[8,] "dateeye2014"   "doEye"
```

```
> names( mic )[dn] <- nnam
> cbind( names(mic),
+       attributes(mic)$var.labels )
```

```
      [,1]           [,2]
[1,] "ID"           "Patient ID"
[2,] "SOPNR"        "SOPNR"
[3,] "_merge"       ""
[4,] "aer_level"    "u-AER class"
[5,] "allocation"   "Treatment allocation"
[6,] "autoprog"     "Progression in autonomic neuropathy"
[7,] "blind"        "blindness according to WHO criteria"
[8,] "doVis"        "Visit date"
[9,] "date_aerprog" ""
[10,] "date_autoprog" ""
[11,] "doBth"       "Date of birth"
[12,] "doBlind"     "Date of first eye-examination with blindness"
[13,] "doDth"       "Date of death"
[14,] "date_eds_l_prog" ""
[15,] "date_eds_prog" ""
[16,] "date_eds_r_prog" ""
[17,] "doEnd"       "Date of last observation"
[18,] "doESRD"      "Date of Renal Replacement Therapy initiation"
[19,] "date_periprog" ""
[20,] "doPhC"       ""
[21,] "date_progeds" ""
[22,] "doEye"       "Date of latest retinography after 2006"
[23,] "eds"         ""
[24,] "eds_l"       "EURO-DIAB score, left"
[25,] "eds_l_prog" ""
[26,] "eds_max"     ""
[27,] "eds_r"       "EURO-DIAB score, right"
[28,] "eds_r_prog" ""
[29,] "esrd"        "End Stage Renal Disease"
[30,] "if_aerprog"  ""
[31,] "if_autoprog" ""
[32,] "if_periprog" ""
[33,] "if_photocoag" ""
[34,] "laser"       "Photocoagulation"
[35,] "mean_aer"    "Geometric mean of u-AER"
[36,] "med_aer"     "Median u-AER"
[37,] "periprog"    "Progression in peripheral neuropathy"
[38,] "prog_eds_max" ""
[39,] "sex"         "Gender"
[40,] "year"        "Follow-up point"
```

```

> mic <- cal.yr( mic )
> str( mic )
'data.frame':      748 obs. of  40 variables:
 $ ID          : num  1 1 1 1 1 1 2 2 2 2 ...
 $ SOPNR       : num  1 1 1 1 1 1 2 2 2 2 ...
 $ _merge      : Factor w/ 5 levels "master only (1)",...: 3 3 3 3 3 3 3 3 3 3 ...
 $ aer_level   : Factor w/ 3 levels "Normoalbuminuria",...: 2 2 1 2 1 2 2 1 2 3 ...
 $ allocation  : Factor w/ 2 levels "Intensive","Conventional": 1 1 1 1 1 1 1 1 1 1 ...
 $ autoprog    : int  NA 0 0 0 0 0 NA 1 1 1 ...
 $ blind       : int  0 NA 0 0 0 0 0 NA 0 0 ...
 $ doVis       :Classes 'cal.yr', 'numeric' num [1:748] 1993 1995 1997 2001 2007 ...
 $ date_aerprog : num  NA NA NA NA NA ...
 $ date_autoprog : num  NA NA NA NA NA ...
 $ doBth       :Classes 'cal.yr', 'numeric' num [1:748] 1932 1932 1932 1932 1932 ...
 $ doBlind     :Classes 'cal.yr', 'numeric' num [1:748] NA NA NA NA NA NA NA NA NA ...
 $ doDth       :Classes 'cal.yr', 'numeric' num [1:748] NA NA NA NA NA NA NA NA NA ...
 $ date_eds_l_prog: num  NA NA NA NA NA ...
 $ date_eds_prog : num  NA NA NA NA NA ...
 $ date_eds_r_prog: num  NA NA NA NA NA ...
 $ doEnd       :Classes 'cal.yr', 'numeric' num [1:748] 2015 2015 2015 2015 2015 ...
 $ doESRD      :Classes 'cal.yr', 'numeric' num [1:748] NA NA NA NA NA NA NA NA NA ...
 $ date_periprog : num  NA NA NA NA NA ...
 $ doPhC       :Classes 'cal.yr', 'numeric' num [1:748] NA NA NA NA NA ...
 $ date_progeds : num  NA NA NA NA NA ...
 $ doEye       :Classes 'cal.yr', 'numeric' num [1:748] 2015 2015 2015 2015 2015 ...
 $ eds         : num  NA NA NA NA NA 1 NA NA NA NA ...
 $ eds_l       : Factor w/ 6 levels "No retinopathy",...: 1 NA 1 1 1 NA 2 NA 2 6 ...
 $ eds_l_prog  : int  NA NA 0 0 0 NA NA NA 0 1 ...
 $ eds_max     : num  0 NA 0 0 0 1 1 NA 1 4 ...
 $ eds_r       : Factor w/ 6 levels "No retinopathy",...: 1 NA 1 1 1 NA 2 NA 2 3 ...
 $ eds_r_prog  : int  NA NA 0 0 0 NA NA NA 0 1 ...
 $ esrd        : int  0 0 0 0 0 0 0 0 0 0 ...
 $ if_aerprog  : int  0 0 0 0 0 0 0 0 0 1 ...
 $ if_autoprog : int  0 0 0 0 0 0 0 1 1 1 ...
 $ if_periprog : int  0 0 0 0 0 0 0 0 0 1 ...
 $ if_photocoag : int  0 0 0 0 0 0 0 0 0 1 ...
 $ laser       : int  NA NA NA 0 0 NA NA NA NA 1 ...
 $ mean_aer    : num  76.9 34.9 21 92.8 19.5 ...
 $ med_aer     : num  76.5 34 23 105 18 83 69.5 17 86 328 ...
 $ periprog    : int  NA NA 0 0 0 0 NA NA 0 1 ...
 $ prog_eds_max : int  0 NA 0 0 0 1 0 NA 0 1 ...
 $ sex         : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 2 2 2 ...
 $ year        : int  1993 1995 1997 2001 2006 2014 1993 1995 1997 2001 ...
 - attr(*, "var.labels")= chr  "Patient ID" "SOPNR" "" "u-AER class" ...
> # How many have how many visits
> addmargins( t( apply(mic$ID,mic$allocation), 2, table ) )[, -1 ]
      1 2 3 4 5 6 Sum
Intensive  2 2 10 12 15 39 80
Conventional 1 3 13 27 15 21 80
Sum        3 5 23 39 30 60 160

```

We devise a variable `visit` that just numbers the visits from 1 to 6, and also plot the visit dates versus the visit numbers:

```

> mic$visit <- Relevel( factor(floor(mic$doVis)),
+                       list("1"=1:2,
+                             "2"=3,
+                             "3"=4,
+                             "4"=5,
+                             "5"=6:7,
+                             "6"=8:9) )
> mic$visit <- as.numeric( as.character( mic$visit ) )
> # Check that we actually got the numbering correct:
> print( with( mic, addmargins( table( floor(doVis*5)/5, visit, useNA="ifany" ),
+                                   margin=1 ) ),
+        zero.print="." )

```

	visit					
	1	2	3	4	5	6
1993	35
1993.2	43
1993.4	16
1993.6	19
1993.8	16
1994	21
1994.2	10
1995	.	50
1995.2	.	40
1995.4	.	8
1995.6	.	46
1995.8	.	12
1997	.	.	52	.	.	.
1997.2	.	.	33	.	.	.
1997.4	.	.	10	.	.	.
1997.6	.	.	45	.	.	.
1997.8	.	.	9	.	.	.
2001	.	.	.	33	.	.
2001.2	.	.	.	38	.	.
2001.4	.	.	.	1	.	.
2001.6	.	.	.	39	.	.
2001.8	.	.	.	17	.	.
2006.6	38	.
2006.8	47	.
2007	8	.
2014	2
2014.2	2
2014.4	9
2014.6	30
2014.8	17
2015	2
Sum	160	156	149	128	93	62

```
> with( mic, plot( doVis,
+                 visit+ID/300-88/300,
+                 pch=16, cex=0.5,
+                 xlab="Date of examination",
+                 ylab="Visit no" ) )
> abline( v=1990:2015, h=seq(-0.5,5.5,1), col=gray(0.8) )
```

Finally we restrict the visits dataset to the relevant variables:

```
> mic <- mic[,c("ID","allocation","sex",
+              "doBth","doVis","doESRD","doEye","doPhC","doBlind","doEnd","doDth",
+              "visit","aer_level","med_aer","eds_l","eds_r","autoprog","periprogram")]
> save( mic, file="../data/mic.Rda" )
```

For the sake of analysis of ESRD occurrence we make a small dataset with ID and ESRD dates for merging with the baseline data set:

```
> zz <- with( mic, tapply( doESRD, ID, mean, na.rm=TRUE ) )
> ESRD <- data.frame( ID = as.numeric(names(zz)),
+                    doESRD = as.numeric(zz) )
> str(ESRD)
'data.frame':      160 obs. of  2 variables:
 $ ID      : num  1 2 3 4 5 6 7 8 9 10 ...
 $ doESRD: num  NaN NaN NaN NaN 1998 ...
> save( ESRD, file="../data/ESRD.Rda" )
```



```
> ## Change name of date variables
> names(st2) <- gsub("date.", "do", gsub("_", ".", names(st2)))
> # Assign a date of DM diagnosis, randomly in the year given
> set.seed(78468)
> st2$dodm <- st2$dobaseline - ( st2$dmvar93 + runif(nrow(st2)) )
```

Now rename the date variables to something a bit more mnemonic:

```
> names(st2)[wh <- substr(names(st2), 1, 2) == "do"]
[1] "dobaseline" "dobirth" "dodeath" "doend" "doevent1" "doevent2"
[7] "doevent3" "doESRD" "dodm"
> newnam <- c("doBase", "doBth", "doDth", "doEnd", "doCVD1", "doCVD2", "doCVD3", "doESRD", "doDM")
> cbind( names(st2)[wh], newnam )

      newnam
[1,] "dobaseline" "doBase"
[2,] "dobirth"    "doBth"
[3,] "dodeath"    "doDth"
[4,] "doend"      "doEnd"
[5,] "doevent1"  "doCVD1"
[6,] "doevent2"  "doCVD2"
[7,] "doevent3"  "doCVD3"
[8,] "doESRD"    "doESRD"
[9,] "dodm"       "doDM"
> names(st2)[wh] <- newnam
> names(st2)[wh <- grep("cvd", names(st2))]
[1] "base.cvd" "cvd.death"
> newnam <- c("baseCVD", "deathCVD")
> cbind( names(st2)[wh], newnam )

      newnam
[1,] "base.cvd" "baseCVD"
[2,] "cvd.death" "deathCVD"
> names(st2)[wh] <- newnam
```

Finally we subset and reorder the variables — note that we for convenience have two versions of the allocation variable with short and long levels names.

```
> st2 <- st2[, c("ID", "allocation", "sex", "baseCVD", "deathCVD",
+              "doBth", "doDM", "doBase", "doCVD1", "doCVD2", "doCVD3", "doESRD", "doEnd", "doDth")]
```

1.2.1 Checking the date variables

We make pairwise plot of all date variables, in the logical order of the variables so that we can see that all points should be on the same side of the red identity lines in figure 1.2:

```
> names(st2)[dvar <- grep("do", names(st2))]
[1] "doBth" "doDM" "doBase" "doCVD1" "doCVD2" "doCVD3" "doESRD" "doEnd" "doDth"
> par( bty="o" )
> pairs( st2[, dvar], gap=0,
+       panel=function(x,y){points(x,y,pch=16,cex=0.8)
+       abline(0,1,col="red")} )
> par( bty="n" )
```

From the pairwise plots it is seen that the end-dates do not have a single value at the end where all persons alive are censored; the date in the variable doEnd is the date of last examination.

It seems as if some of the event dates for successive events are identical, but this is actually not the case; here is a list of all (CVD)event dates recorded after baseline, and of the differences (in years) between successive CVD event dates:

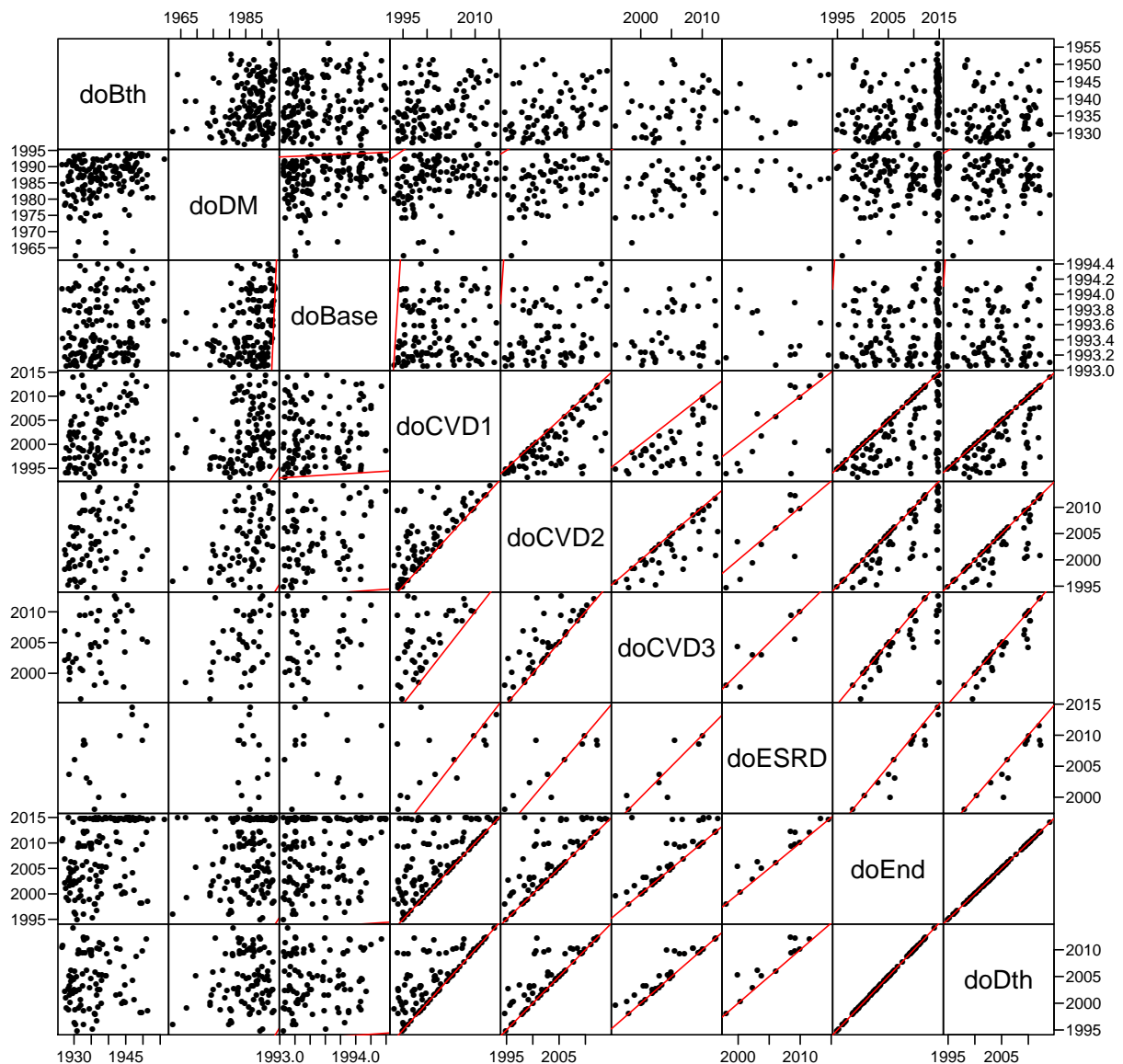


Figure 1.2: Pairwise plots of all dates in the data frame. The red lines are the identity lines. With the exception of doCVD and doESRD, points in the upper panels should be right / below the identity lines.

```
> ev <- st2[,c("doBase", "doCVD1", "doCVD2", "doCVD3", "doEnd", "doDth")]
> head(ev)
  doBase doCVD1 doCVD2 doCVD3 doEnd doDth
1 1993.357 2014.457      NA      NA 2014.778      NA
2 1993.357 2009.231 2009.570 2010.162 2014.589      NA
3 1993.362 2001.639      NA      NA 2001.639 2001.639
4 1993.362 1995.415 1997.461 2003.443 2003.443 2003.443
5 1993.162 1993.959 1994.657 1998.047 1998.047 1998.047
6 1993.201 1998.769      NA      NA 2014.684      NA
> wh <- apply(ev[,2:4], 1, function(x) sum(is.na(x))) < 3
> round( cbind( ev[wh,], t(apply(ev[wh,2:4], 1, diff)) ), 3 )
  doBase doCVD1 doCVD2 doCVD3 doEnd doDth doCVD2 doCVD3
1 1993.357 2014.457      NA      NA 2014.778      NA      NA      NA
```

2	1993.357	2009.231	2009.570	2010.162	2014.589	NA	0.339	0.591
3	1993.362	2001.639	NA	NA	2001.639	2001.639	NA	NA
4	1993.362	1995.415	1997.461	2003.443	2003.443	2003.443	2.045	5.982
5	1993.162	1993.959	1994.657	1998.047	1998.047	1998.047	0.698	3.389
6	1993.201	1998.769	NA	NA	2014.684	NA	NA	NA
7	1993.203	2001.058	2006.238	NA	2006.238	2006.238	5.180	NA
8	1993.209	2011.925	2012.149	NA	2012.149	2012.149	0.225	NA
9	1993.209	1995.123	1995.952	NA	1995.952	1995.952	0.830	NA
10	1993.075	2011.405	NA	NA	2011.405	2011.405	NA	NA
11	1993.066	2005.666	2006.068	NA	2006.068	2006.068	0.402	NA
12	1993.146	1994.000	1995.306	2002.427	2002.739	2002.739	1.306	7.121
14	1993.105	2014.038	NA	NA	2014.038	2014.038	NA	NA
16	1993.053	2000.034	2002.112	2009.680	2009.680	2009.680	2.078	7.567
18	1993.373	1998.260	1998.471	1998.594	2014.997	NA	0.211	0.123
21	1993.379	1994.274	NA	NA	2014.789	NA	NA	NA
23	1993.069	1993.132	1998.994	NA	1998.994	1998.994	5.862	NA
24	1993.187	2005.431	2007.667	2010.156	2010.156	2010.156	2.237	2.489
26	1993.225	1999.780	1999.985	2001.855	2001.855	2001.855	0.205	1.870
27	1993.222	2002.531	2006.175	2006.334	2009.228	2009.228	3.644	0.159
29	1993.280	2005.222	NA	NA	2005.222	2005.222	NA	NA
30	1993.392	2004.300	2005.543	NA	2005.543	2005.543	1.243	NA
31	1993.220	2006.112	2007.235	2008.513	2010.342	2010.342	1.123	1.279
32	1993.220	2000.168	NA	NA	2000.168	2000.168	NA	NA
34	1993.105	1996.045	2005.431	2012.546	2014.745	NA	9.385	7.116
35	1993.496	2001.710	2002.865	2003.018	2005.091	2005.091	1.155	0.153
36	1993.203	2001.912	NA	NA	2014.901	NA	NA	NA
37	1993.069	2008.697	NA	NA	2008.697	2008.697	NA	NA
38	1993.129	1998.742	2000.198	2000.407	2003.322	2003.322	1.457	0.208
39	1993.110	2010.838	NA	NA	2014.654	NA	NA	NA
40	1993.239	1997.765	2000.012	2000.012	2000.012	2000.012	2.248	0.000
41	1993.066	2002.471	NA	NA	2002.471	2002.471	NA	NA
42	1993.244	2002.758	2002.928	2004.902	2005.838	2005.838	0.170	1.974
43	1993.107	1994.052	2002.181	2002.367	2002.367	2002.367	8.129	0.186
45	1993.132	2010.233	NA	NA	2014.745	NA	NA	NA
46	1993.398	1997.409	2001.715	2001.869	2002.517	2002.517	4.307	0.153
47	1993.414	2010.789	NA	NA	2010.789	2010.789	NA	NA
48	1993.417	2002.589	2002.605	NA	2002.605	2002.605	0.016	NA
49	1993.417	2010.564	2011.199	NA	2014.767	NA	0.635	NA
50	1993.168	1996.880	2006.402	2006.780	2014.665	NA	9.522	0.378
51	1993.181	1996.957	2001.904	2002.145	2002.145	2002.145	4.947	0.241
54	1993.206	2012.283	2012.305	NA	2012.305	2012.305	0.022	NA
55	1993.072	1995.602	1996.042	2000.223	2000.223	2000.223	0.441	4.181
56	1993.124	2005.387	NA	NA	2005.387	2005.387	NA	NA
57	1993.431	2004.719	NA	NA	2004.719	2004.719	NA	NA
58	1993.431	1997.647	1997.937	NA	1997.937	1997.937	0.290	NA
60	1993.433	1999.328	2003.035	2004.209	2009.975	2009.975	3.707	1.175
61	1993.242	1999.040	1999.139	2000.743	2000.743	2000.743	0.099	1.604
62	1993.143	2006.786	NA	NA	2006.786	2006.786	NA	NA
64	1993.170	1996.045	1996.045	NA	1996.045	1996.045	0.000	NA
65	1993.299	1994.093	2001.001	NA	2001.001	2001.001	6.908	NA
66	1993.296	2000.710	NA	NA	2000.710	2000.710	NA	NA
68	1993.113	1995.166	2009.203	NA	2009.203	2009.203	14.037	NA
69	1993.302	2004.626	2009.368	2010.181	2014.953	NA	4.742	0.813
71	1993.124	1997.080	1998.695	NA	1998.695	1998.695	1.615	NA
72	1993.318	1997.211	1998.400	1998.999	2001.595	2001.595	1.188	0.600
73	1993.151	2008.867	NA	NA	2008.867	2008.867	NA	NA
74	1993.181	1996.689	NA	NA	1996.689	1996.689	NA	NA
77	1993.335	1994.841	1995.684	1995.875	1999.577	1999.577	0.843	0.192
78	1993.206	2008.305	NA	NA	2014.997	NA	NA	NA
79	1993.231	1999.955	NA	NA	1999.955	1999.955	NA	NA
80	1993.050	1999.194	NA	NA	1999.194	1999.194	NA	NA
81	1993.050	1994.868	1994.873	NA	1994.873	1994.873	0.005	NA
82	1993.165	2009.233	2009.373	NA	2009.373	2009.373	0.140	NA
83	1993.321	2009.710	2009.841	2010.071	2010.071	2010.071	0.131	0.230
85	1993.316	1993.825	2009.422	2009.436	2009.444	2009.444	15.598	0.014
86	1993.201	1998.057	2005.318	2009.515	2014.479	NA	7.261	4.197

87	1993.469	1995.407	2009.636	NA	2014.805	NA	14.229	NA
88	1993.570	2001.521	2004.229	NA	2004.229	2004.229	2.708	NA
89	1993.570	2001.567	2004.998	NA	2004.998	2004.998	3.431	NA
91	1993.584	1998.621	NA	NA	1998.621	1998.621	NA	NA
92	1993.584	2007.728	2007.760	NA	2007.760	2007.760	0.033	NA
93	1993.661	2004.281	NA	NA	2004.281	2004.281	NA	NA
94	1993.587	1998.969	2001.499	NA	2001.499	2001.499	2.530	NA
96	1993.592	1997.340	2000.828	2012.223	2012.223	2012.223	3.488	11.395
99	1993.606	1995.243	NA	NA	1995.243	1995.243	NA	NA
101	1993.622	2014.457	NA	NA	2014.827	NA	NA	NA
103	1993.644	2001.450	NA	NA	2001.450	2001.450	NA	NA
105	1993.663	2009.890	NA	NA	2009.890	2009.890	NA	NA
108	1993.762	1999.188	NA	NA	1999.188	1999.188	NA	NA
109	1993.759	1998.496	1999.374	2002.964	2002.964	2002.964	0.879	3.589
111	1993.778	1998.608	1999.413	2000.872	2003.218	2003.218	0.805	1.459
112	1993.781	2006.260	NA	NA	2006.260	2006.260	NA	NA
113	1993.781	1998.230	NA	NA	1998.230	1998.230	NA	NA
114	1993.775	1996.505	2005.053	2005.179	2005.179	2005.179	8.548	0.126
115	1993.795	1997.001	1998.175	2006.895	2006.895	NA	1.175	8.720
116	1993.838	2013.097	2014.153	NA	2014.668	NA	1.057	NA
118	1993.836	1995.938	2003.552	2004.366	2005.381	2005.381	7.614	0.813
119	1993.836	2002.841	2003.522	NA	2003.522	2003.522	0.682	NA
120	1993.838	1996.064	2000.330	2007.095	2009.351	2009.351	4.266	6.765
121	1993.841	2003.988	2003.905	2009.376	2009.376	2009.376	4.917	0.471
122	1994.205	2007.963	2010.277	2011.109	2014.997	NA	2.313	0.832
123	1993.855	1994.975	1996.371	NA	1996.371	1996.371	1.396	NA
124	1993.855	1997.373	2002.632	NA	2002.632	2002.632	5.259	NA
125	1993.890	2000.483	2000.606	2005.529	2009.770	2009.770	0.123	4.923
126	1993.910	1998.632	NA	NA	1998.632	1998.632	NA	NA
128	1993.910	1995.503	1998.230	NA	1998.230	1998.230	2.727	NA
129	1993.929	2001.236	2001.921	2005.159	2014.665	NA	0.684	3.239
130	1993.915	2007.708	2011.643	2012.081	2012.081	2012.081	3.934	0.438
131	1993.932	2002.487	NA	NA	2002.487	2002.487	NA	NA
132	1993.929	2001.444	2011.914	NA	2011.914	2011.914	10.470	NA
133	1994.047	1999.240	NA	NA	1999.240	1999.240	NA	NA
134	1994.066	1994.496	1996.330	1997.723	2000.338	2000.338	1.834	1.394
135	1994.082	2012.713	NA	NA	2014.997	NA	NA	NA
136	1994.011	2012.522	NA	NA	2014.997	NA	NA	NA
138	1994.052	2004.122	NA	NA	2004.122	2004.122	NA	NA
139	1994.049	2009.214	2012.678	NA	2014.750	NA	3.463	NA
140	1994.074	1995.706	2004.442	2004.700	2004.700	2004.700	8.736	0.257
142	1994.066	2002.331	NA	NA	2014.457	NA	NA	NA
143	1994.205	2007.522	2011.076	NA	2011.076	2011.076	3.554	NA
144	1994.104	2009.967	NA	NA	2009.967	2009.967	NA	NA
145	1994.071	2010.465	NA	NA	2010.465	2010.465	NA	NA
146	1994.071	1994.983	1998.701	NA	1998.701	1998.701	3.718	NA
148	1994.085	2002.107	2004.439	NA	2004.439	2004.439	2.333	NA
149	1994.140	2002.186	NA	NA	2002.186	2002.186	NA	NA
150	1994.120	2007.314	2008.494	2008.494	2010.378	2010.378	1.180	0.000
152	1994.142	2002.266	2013.792	NA	2014.824	NA	11.526	NA
156	1994.339	2012.051	NA	NA	2012.051	2012.051	NA	NA
159	1994.397	1998.742	2013.099	NA	2014.611	NA	14.357	NA

1.3 Lexis objects of follow-up

In order to get an overview of follow-up of persons we set up a Lexis object with 4 timescales: calendar time, age, diabetes duration and time since baseline:

```
> names( st2 )
[1] "ID"           "allocation"  "sex"         "baseCVD"    "deathCVD"   "doBth"
[7] "doDM"        "doBase"     "doCVD1"     "doCVD2"    "doCVD3"     "doESRD"
[13] "doEnd"       "doDth"
```

```
> L0 <- Lexis( entry = list( per = doBase,
+                             age = doBase-doBth,
+                             dur = doBase-doDM,
+                             tsb = 0 ),
+             exit = list( per = doEnd ),
+             exit.status = factor( (!is.na(doDth))+deathCVD,
+                                   labels=c("DM","Dead","CV-D") ),
+             data = st2 )
```

NOTE: entry.status has been set to "DM" for all.

```
> summary( L0 )
```

Transitions:

		To						
From	DM	Dead	CV-D	Records:	Events:	Risk time:	Persons:	
	DM	67	55	38	160	93	2417.6	160

We then cut the follow-up at the CVD event dates:

```
> L1 <- cutLexis( L0, cut=L0$doCVD1,
+                 new.state="1st CVD", split.states=TRUE,
+                 precursor.states=c("DM") )
> L1 <- cutLexis( L1, cut=L1$doCVD2,
+                 new.state="2nd CVD", split.states=TRUE,
+                 precursor.states=c("DM","1st CVD") )
> L1 <- cutLexis( L1, cut=L1$doCVD3,
+                 new.state="3rd CVD", split.states=TRUE,
+                 precursor.states=c("DM","1st CVD","2nd CVD") )
```

When keeping track of the states *from* which death occurs, we get awfully long state names by default, so we groom these and finally reorder them to facilitate plotting and summarizing;

```
> levels( L1 )
 [1] "DM" "1st CVD"
 [3] "2nd CVD" "3rd CVD"
 [5] "Dead" "CV-D"
 [7] "Dead(1st CVD)" "CV-D(1st CVD)"
 [9] "Dead(1st CVD)(2nd CVD)" "CV-D(1st CVD)(2nd CVD)"
[11] "CV-D(1st CVD)(2nd CVD)(3rd CVD)" "Dead(1st CVD)(2nd CVD)(3rd CVD)"

> # A function to tease out the number of previous events from the state
> # names, basically fishing out the 8th last character
> subrv8 <- function(x) ifelse( (wh<-(nchar(x)-7))>0, substr(x,wh,wh), "0" )
> cbind(
+ levels( L1$lex.Xst )[-(1:4)],
+ paste( substr( levels( L1$lex.Xst )[-(1:4)], 1, 4 ),
+         subrv8( levels( L1$lex.Xst )[-(1:4)] ) ) )
      [,1] [,2]
 [1,] "Dead" "Dead 0"
 [2,] "CV-D" "CV-D 0"
 [3,] "Dead(1st CVD)" "Dead 1"
 [4,] "CV-D(1st CVD)" "CV-D 1"
 [5,] "Dead(1st CVD)(2nd CVD)" "Dead 2"
 [6,] "CV-D(1st CVD)(2nd CVD)" "CV-D 2"
 [7,] "CV-D(1st CVD)(2nd CVD)(3rd CVD)" "CV-D 3"
 [8,] "Dead(1st CVD)(2nd CVD)(3rd CVD)" "Dead 3"

> levels( L1$lex.Cst )[-(1:4)] <-
+ levels( L1$lex.Xst )[-(1:4)] <-
+ paste( substr( levels( L1$lex.Xst )[-(1:4)], 1, 4 ),
+         subrv8( levels( L1$lex.Xst )[-(1:4)] ) )
> L1 <- Relevel( L1, c(1:10,12,11) )
> summary( L1 )
```

Transitions:

From	To	DM	1st CVD	2nd CVD	3rd CVD	Dead 0	CV-D 0	Dead 1	CV-D 1	Dead 2	CV-D 2	Dead 3
DM		41	86	0	0	24	9	0	0	0	0	0
1st CVD		0	11	48	0	0	0	14	13	0	0	0
2nd CVD		0	0	7	24	0	0	0	0	12	4	0
3rd CVD		0	0	0	8	0	0	0	0	0	0	5
Sum		41	97	55	32	24	9	14	13	12	4	5

Transitions:

From	To	CV-D 3	Records:	Events:	Risk time:	Persons:
DM		0	160	119	1870.64	160
1st CVD		0	86	75	342.61	86
2nd CVD		0	47	40	112.27	47
3rd CVD		12	25	17	92.08	25
Sum		12	318	251	2417.60	160

With this in order we can now show how persons fare through the study, separately for the two randomization groups:

```
> par( mfrow=c(1,2) )
> for( i in 1:2 ){
+ boxes( subset(L1,allocation==levels(allocation)[i]),
+       boxpos=list(x=c(rep(20,4),rep(80,8)),
+                   y=c(seq(90,10,,4),
+                       rep(seq(92,8,,4),each=2)+rep(c(1,-1)*4,4) ) ),
+       pos.arr=rep(c(5,4,6)/10,4)[-12],#[-15+i],
+       show.BE="nz", DR.sep=c("(", ")"), eq.ht=FALSE, hmult=1.3,
+       scale.R=1000)
+ text(10,par("usr")[4],levels(L1$allocation)[i],adj=c(0,1),font=2,cex=2)
+ }
```

1.3.1 Follow-up after end of intervention

According to Gæde *et al.*[1] the trial intervention ended at a mean of 7.8 years after intervention, so we define this as the end of study:

```
> as.Date.cal.yr( doStEnd <- mean(st2$doBase) + 7.8 )
[1] "2001-05-09"
```

First we construct the dataset P1 restricted to follow-up after end of study:

```
> S1 <- splitLexis( L1, breaks=doStEnd, time.scale="per" )
> S1[S1$lex.id<4,1:9]
  lex.id  per      age      dur      tsb      lex.dur lex.Cst lex.Xst ID
1      1 1993.357 61.05681  2.263567  0.000000  7.9942505      DM      DM  1
2      1 2001.351 69.05106 10.257817  7.994251 13.1063655      DM 1st CVD  1
3      1 2014.457 82.15743 23.364183 21.100616  0.3203285 1st CVD 1st CVD  1
4      2 1993.357 46.54346 11.226522  0.000000  7.9942505      DM      DM  2
5      2 2001.351 54.53771 19.220772  7.994251  7.8798084      DM 1st CVD  2
6      2 2009.231 62.41752 27.100581 15.874059  0.3394935 1st CVD 2nd CVD  2
7      2 2009.570 62.75702 27.440074 16.213552  0.5913758 2nd CVD 3rd CVD  2
8      2 2010.162 63.34839 28.031450 16.804928  4.4271047 3rd CVD 3rd CVD  2
9      3 1993.362 49.98494 10.801806  0.000000  7.9887748      DM      DM  3
10     3 2001.351 57.97372 18.790580  7.988775  0.2877481      DM  Dead 0  3

> P1 <- subset( S1, per>=doStEnd )
> P1[P1$lex.id<4,1:9]
  lex.id  per      age      dur      tsb      lex.dur lex.Cst lex.Xst ID
2      1 2001.351 69.05106 10.25782  7.994251 13.1063655      DM 1st CVD  1
3      1 2014.457 82.15743 23.36418 21.100616  0.3203285 1st CVD 1st CVD  1
5      2 2001.351 54.53771 19.22077  7.994251  7.8798084      DM 1st CVD  2
```

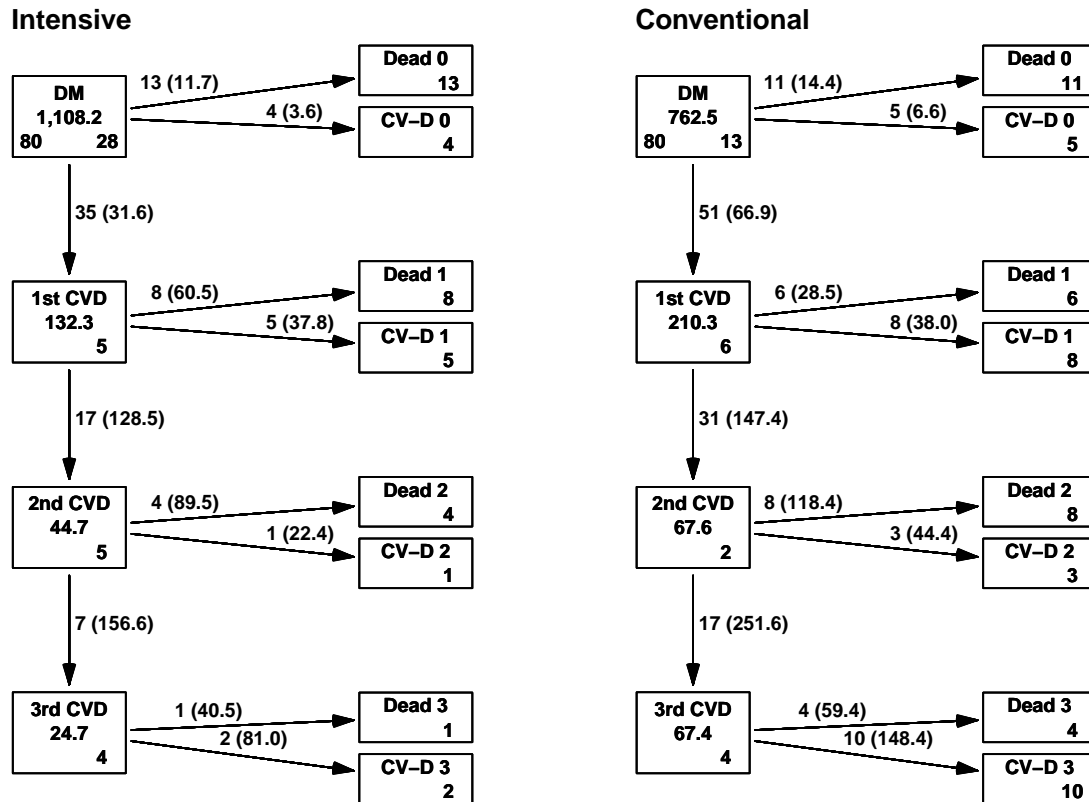


Figure 1.3: Flow of patients from the Steno2 study through CVD states to death by intervention group.

The numbers in the middle of the boxes are the person-years, the number at the bottom of the boxes are the number of persons starting (left), resp. ending (right) their follow-up in each state. The numbers on the arrows are the number of transitions (transition-rates per 1000 PY).

```
6      2 2009.231 62.41752 27.10058 15.874059 0.3394935 1st CVD 2nd CVD 2
7      2 2009.570 62.75702 27.44007 16.213552 0.5913758 2nd CVD 3rd CVD 2
8      2 2010.162 63.34839 28.03145 16.804928 4.4271047 3rd CVD 3rd CVD 2
10     3 2001.351 57.97372 18.79058 7.988775 0.2877481      DM  Dead 0  3
```

We observe that the first record for each person has the same date of start (that is, value of `per`).

We then show the plot of the states and transitions but only for the follow-up starting at end of intervention (that is the date from which the groups are on identical regimens):

```
> par( mfrow=c(1,2) )
> for( whl in levels(P1$allocation) )
+ {
+ boxes( subset( P1, allocation==whl ),
+         boxpos=list(x=c(rep(20,4),rep(80,8)),
+                       y=c( seq(90,10,,4),
+                             rep(seq(92,8,,4),each=2)+rep(c(1,-1)*4,4) ) ),
+         # pos.arr=rep(c(5,4,6)/10,5)[-13][-15+i],
+         show.BE="nz", DR.sep=c("(", ")"), eq.ht=FALSE, hmult=1.3,
+         scale.R=1000)
+ text(20,par("usr")[4],whl,adj=c(0.5,1),font=2,cex=2)
+ }
```

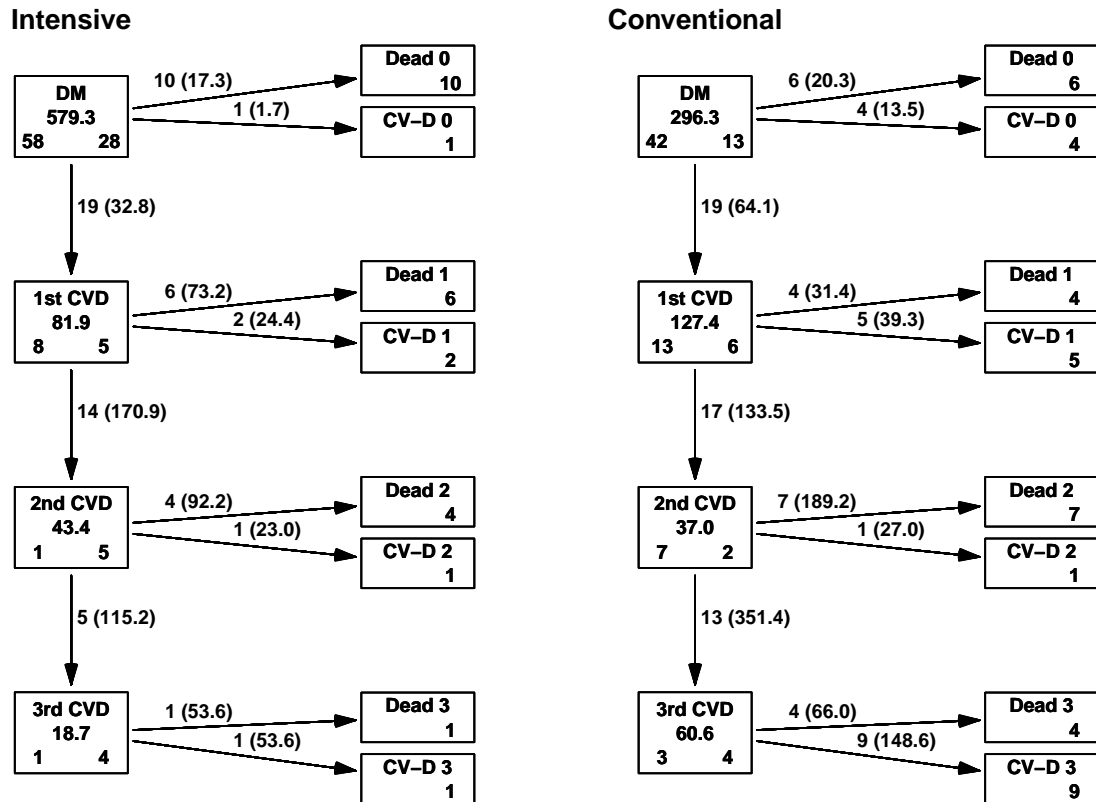



Figure 1.4: Flow of patients from the Steno2 study by randomization group, restricted to the period **after** end of intervention (May 2001).

The numbers in the middle of the boxes are the person-years, the number at the bottom of the boxes are the number of persons starting (left), resp. ending (right) their follow-up in each state. The numbers on the arrows are the number of transitions (transition-rates per 1000 PY).

Note that the dataset we have created here is only made for illustrational purposes, and will not be used in any analyses; there is no use for the date at which the two groups were merged as the effect of merging the group should be expected to be smooth and is thus automatically modeled by non-linear effects of time since baseline.

1.4 Datasets saved for analysis

Finally we save the relevant datasets, namely L0 (only one record of total follow-up for each person), and L1 where follow-up is cut by CVD event. In both datasets we have a subdivision of

```
> save( st2, file="../data/st2.Rda" )
> save( L0, L1, file="../data/Lx.Rda")
```

1.5 Datasets saved for public use

In order to give the public access to the data from the trial we anonymize data by adding a random quantity between -7 and $+7$ days to all dates recorded. This means that no dates or combinations of dates in the publicly available datasets occur in any real individual. Moreover we randomly assign new ID numbers to persons, so persons now have IDs in the range 1001–1160.

1.5.1 Merge to one data set

```
> clear()
> load( "../data/mic.Rda" )
> load( "../data/st2.Rda" )
> ( whd <- match(c("doEye","doPhC","doBlind"),names(mic)) )
[1] 7 8 9
> dmic <- aggregate( mic[,whd],
+                   mic[, "ID", drop=FALSE],
+                   FUN=function(x) mean(x, na.rm=TRUE) )
> str( dmic )
'data.frame':
  160 obs. of  4 variables:
 $ ID      : num  1 2 3 4 5 6 7 8 9 10 ...
 $ doEye   : num  2015 2015 NaN NaN NaN ...
 $ doPhC   : num  2015 2008 NaN NaN NaN ...
 $ doBlind : num  NaN NaN NaN NaN NaN ...
> st2 <- merge( st2, dmic )
> # Locate all date variables
> names(st2)[dn <- grep( "do", names(st2) )]
 [1] "doBth" "doDM" "doBase" "doCVD1" "doCVD2" "doCVD3" "doESRD" "doEnd"
 [9] "doDth" "doEye" "doPhC" "doBlind"
> # Add +/- 7 days to the dates
> for( i in dn ) st2[,i] <- st2[,i] + runif(160,-7,7)/365
> # Check and fix the internal consistency of dates:
> # doBth < doDM < doBase < doCVD1 < doCVD2 < doCVD3 < doDth
> # doBase < (doEye / doPhC / doBlind / doESRD) < doEnd < doDth
> # doBase < (=for visit1) doVis < (=for last visit) doEnd
>
> # The variables not constant within persons
> ( whm <- c( "ID", setdiff(names(mic), names(st2)) ) )
 [1] "ID" "doVis" "visit" "aer_level" "med_aer" "eds_l" "eds_r"
 [8] "autoprog" "periprog"
> tot <- merge( mic[,whm], st2, all.x=TRUE )
```

Generate bogus ids:

```
> ID <- unique(tot$ID)
> ID <- ID[sample(1:length(ID))]
> id <- data.frame( ID=ID, id=1000+1:160 )
> dim( id )
[1] 160  2
> dim( tot )
[1] 748 25
> tot <- merge( id, tot, all=TRUE )[, -1]
> visits <- tot[,c("id","allocation","sex","visit","doVis",
+                "aer_level","med_aer","eds_l","eds_r","autoprog","periprog",
+                "doEye","doPhC","doBlind","baseCVD","deathCVD",
+                "doBth","doDM","doBase","doCVD1","doCVD2","doCVD3",
+                "doESRD","doEnd","doDth")]
> base <- aggregate( visits[,c("baseCVD","deathCVD",
```

```
+           "doBth", "doDM", "doBase", "doCVD1", "doCVD2", "doCVD3",
+           "doESRD", "doEnd", "doDth")],
+   visits[,c("id", "allocation", "sex")],
+   FUN=function(x) mean(x, na.rm=TRUE) )
> names( visits )
 [1] "id"           "allocation"  "sex"         "visit"       "doVis"       "aer_level"
 [7] "med_aer"     "eds_l"      "eds_r"       "autoprog"    "periprog"    "doEye"
[13] "doPhC"       "doBlind"    "baseCVD"     "deathCVD"   "doBth"       "doDM"
[19] "doBase"      "doCVD1"     "doCVD2"     "doCVD3"     "doESRD"      "doEnd"
[25] "doDth"
> names( base )
 [1] "id"           "allocation"  "sex"         "baseCVD"     "deathCVD"    "doBth"
 [7] "doDM"        "doBase"     "doCVD1"     "doCVD2"     "doCVD3"     "doESRD"
[13] "doEnd"       "doDth"
> package.skeleton( "St2", c("visits", "base"), path="../data/", force=TRUE )
```

Chapter 2

Statistical analysis of rates

2.1 Concepts

2.1.1 Outcomes

There are two broad categories of events:

- Deaths:
 - CVD-death (defined by *cause*)
 - Other
 - All (the union of the two previous)
- CVD-events:
 - First event
 - Any event
 - Number of events
- Life lost, quality years lived:

These measures are derivatives of transition intensities (transition rates), basically summaries of the time spent in different states — the simplest being in the “alive” state, the more complicated being “without CVD”, say. Since these are *absolute measures* and not relative measures, they are not easy to generalize from the patient population at hand, and so they are better modelled also including age, sex and other demographic characteristics, and reported for select values of these.

2.1.2 Predictors

Beyond the randomization class (Intensive / Conventional), there is a specific interest in describing how mortality changes after intervention end, where the control group were “lifted” to the level of the Intensive group.

Hence we would primarily be interested in the effect of calendar time (or time since baseline which is effectively the same) on the mortality and in particular interaction between Intensive group and calendar time. This is best addressed by a parametric model for the mortality as function of time.

It is not likely that we will have power in the study to address effects of age and diabetes duration in any detail. Hence analysis of rates (mortality or CVD) will be by allocation group and time since randomization, and possibly by CVD status (such as current number of CVD events).

2.1.3 Multistate models

The overview of the patient flow shown in the previous chapter lends itself to quantification of years lived in various states, by using estimated transition rates for all transitions to predict the probability of being in any of the states at any time, and subsequently quantifying how long patients in different groups have spent in these states. Typically, the intervention effect can be assessed on this scale as well by subtracting the years lived for patients in the two groups, but this difference is not likely to be generalizable beyond the specific patient group at hand, so we may consider modeling the rates using at least age and sex as predictors too, so that the intervention effect of years lived can be quantified as a function of these.

2.2 Age- and sex-distribution

The age-distribution at baseline is a bit ragged, and the sex-distribution is very skew (41 women, 119 men) as seen in figure 2.1:

```
> library( Epi )
> clear()
> load( file="../data/Lx.Rda" )
> lls()
  name mode class      size
1 L0  list Lexis data.frame 160 23
2 L1  list Lexis data.frame 318 23

> par( mfrow=c(1,2) )
> for( sx in levels(L0$sex)[2:1] )
+ {
+ with( subset(L0,sex==sx),
+       hist( age, col=if(sx=="M") "blue" else "red",
+             border=if(sx=="M") "blue" else "red",
+             breaks=seq(36,69,3), ylim=c(0,22),
+             main="", xlab="" ) )
+ par( new=TRUE )
+ with( subset(L0,sex==sx & allocation=="Intensive"),
+       hist( age, col="#AAAAAAA",
+             border="transparent",
+             breaks=seq(36,69,3), ylim=c(0,22),
+             main="", xlab="" ) )
+ }
> mtext( "Age at baseline", side=1, line=-3, outer=TRUE )
```

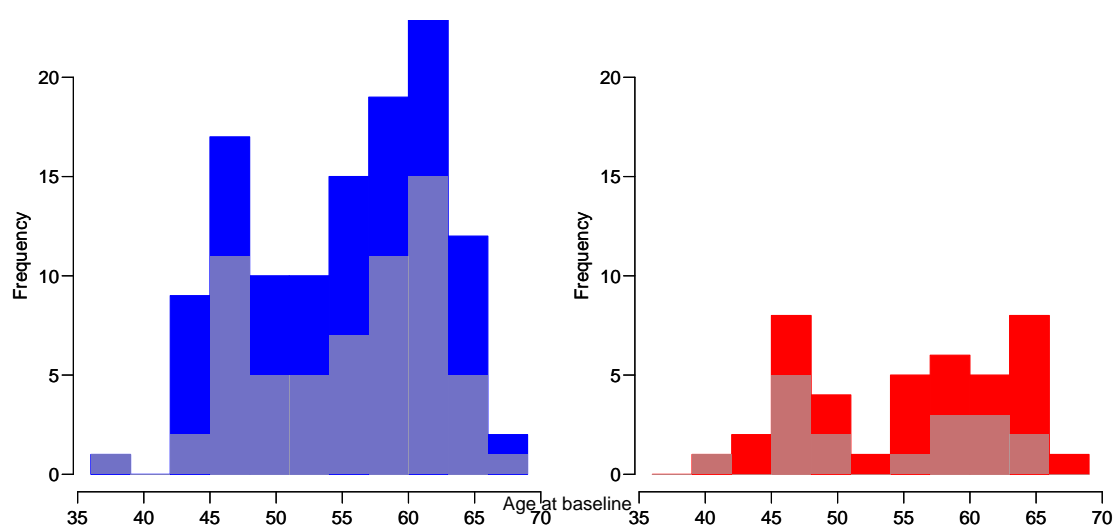


Figure 2.1: *Age-distribution among men (blue) and women (red), the light areas are the persons allocated to the intensive group. Age-classes are 3 years wide.*

Chapter 3

Mortality rates

3.1 Differences in median survival

First we compute the crude Kaplan-Meier-estimates of overall survival and CVD-free survival

To this end we take the previous analysis dataset but merge the two types of death:

```
> library( survival )
> library( Epi )
> clear()
> sessionInfo()
R version 3.3.0 (2016-05-03)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Ubuntu 14.04.4 LTS

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C              LC_TIME=en_US.UTF-8
 [4] LC_COLLATE=en_US.UTF-8   LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8     LC_NAME=C                 LC_ADDRESS=C
[10] LC_TELEPHONE=C           LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] utils      datasets  graphics  grDevices  stats      methods    base

other attached packages:
[1] Epi_2.5      survival_2.39-4

loaded via a namespace (and not attached):
 [1] cmprsk_2.2-7      MASS_7.3-44        plyr_1.8.3         Matrix_1.2-1
 [5] parallel_3.3.0   etm_0.6-2          Rcpp_0.11.6        splines_3.3.0
 [9] grid_3.3.0       numDeriv_2014.2-1 lattice_0.20-31
> load( file="../data/Lx.Rda" )
> lls()
  name mode class      size
1 L0  list Lexis data.frame 160 22
2 L1  list Lexis data.frame 318 22
> levels( L0 )
[1] "DM" "Dead" "CV-D"
> levels( L1 )
 [1] "DM" "1st CVD" "2nd CVD" "3rd CVD" "Dead 0" "CV-D 0" "Dead 1" "CV-D 1"
 [9] "Dead 2" "CV-D 2" "Dead 3" "CV-D 3"
> L1 <- Relevel( L1, list("Dead(no CVD)"=5:6,
+ "Dead(1 CVD)"=7:8,
+ "Dead(2 CVD)"=9:10,
+ "Dead(3 CVD)"=11:12), first=FALSE )
```

```

      type      old      new
1  lex.Cst      DM      DM
2  lex.Cst 1st CVD  1st CVD
3  lex.Cst 2nd CVD  2nd CVD
4  lex.Cst 3rd CVD  3rd CVD
5  lex.Cst Dead 0
6  lex.Cst CV-D 0
7  lex.Cst Dead 1
8  lex.Cst CV-D 1
9  lex.Cst Dead 2
10 lex.Cst CV-D 2
11 lex.Cst Dead 3
12 lex.Cst CV-D 3
13 lex.Xst      DM      DM
14 lex.Xst 1st CVD  1st CVD
15 lex.Xst 2nd CVD  2nd CVD
16 lex.Xst 3rd CVD  3rd CVD
17 lex.Xst Dead 0 Dead(no CVD)
18 lex.Xst CV-D 0 Dead(no CVD)
19 lex.Xst Dead 1 Dead(1 CVD)
20 lex.Xst CV-D 1 Dead(1 CVD)
21 lex.Xst Dead 2 Dead(2 CVD)
22 lex.Xst CV-D 2 Dead(2 CVD)
23 lex.Xst Dead 3 Dead(3 CVD)
24 lex.Xst CV-D 3 Dead(3 CVD)
> levels( L1 )
[1] "DM"          "1st CVD"      "2nd CVD"      "3rd CVD"      "Dead(no CVD)"
[6] "Dead(1 CVD)" "Dead(2 CVD)" "Dead(3 CVD)"
> summary( L1 )
Transitions:
  To
From      DM 1st CVD 2nd CVD 3rd CVD Dead(no CVD) Dead(1 CVD) Dead(2 CVD) Dead(3 CVD)
DM          41      86        0         0          33           0           0           0
1st CVD     0       11       48         0           0          27           0           0
2nd CVD     0        0        7        24           0           0          16           0
3rd CVD     0        0        0         8           0           0           0          17
Sum         41      97       55        32          33          27          16          17

Transitions:
  To
From      Records: Events: Risk time: Persons:
DM          160      119    1870.64     160
1st CVD     86       75     342.61      86
2nd CVD     47       40     112.27      47
3rd CVD     25       17     92.08       25
Sum         318      251    2417.60    160

```

The Kaplan-Meier curve for overall death is based on all risk time in any fo the alive states, and the event is any death, whereas the event “death or CVD” is based only on risk time in “DM” and events being either “1st CVD” or “D0”:

```

> ( Xall <- levels(L0)[2:3] )
[1] "Dead" "CV-D"
> ( Ccvd <- levels(L1)[1] )
[1] "DM"
> ( Xcvd <- levels(L1)[c(2,5)] )
[1] "1st CVD"      "Dead(no CVD)"

```

With this fixed we can set up the dataset needed for specification of the combined endpoint of death and first CVD (CVD-free survival), and then the two Kaplan-Meier analyses:


```

> Lcvd <- subset( L1, lex.Cst %in% Ccvd )
> # The ids are all in the same order
> all( L0$lex.id == Lcvd$lex.id )
[1] TRUE
> KMall <- survfit( Surv(lex.dur,lex.Xst %in% Xall) ~ allocation,
+                 data = L0 )
> KMcvd <- survfit( Surv(lex.dur,lex.Xst %in% Xcvd) ~ allocation,
+                 data = Lcvd )
> RTall <- do.call( cbind, summary( KMall, times=0:4*5)[c("time","n.risk")] )
> RTCvd <- do.call( cbind, summary( KMcvd, times=0:4*5 ) [c("time","n.risk")] )

```

We can plot the two sets of survival curves:

```

> nt <- nrow(RTall)/2
> clr <- c("forestgreen","red")
> par( mfrow=c(1,2), mar=c(6,3,1,1), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> plot( KMall, conf.int=TRUE, col=clr, lwd=1, yaxs="i", ylim=c(0,1.01),
+       ylab="Overall survival" )
> abline( h=0.5, v=summary(KMall)$table["median"] )
> axis( side=1, at=1:21, labels=NA, tcl=-0.3 )
> axis( side=2, at=1:9/10, labels=NA, tcl=-0.3 )
> text( rep(1,2), c(9,5)/100, c("Intensive","Conventional"),
+       col=clr, font=2, cex=1.2, adj=0 )
> lines( KMall, col=clr, lwd=4 )
> for(j in 0:1) mtext( RTall[1:nt+nt*j,2], at= RTall[1:nt+nt*j,1],
+                   line=2.5+j, side=1, col=clr[j+1], font=2, cex=1.2 )
> plot( KMcvd, conf.int=TRUE, col=clr, lwd=1, yaxs="i", ylim=c(0,1.01),
+       ylab="CVD-free survival" )
> abline( h=0.5, v=summary(KMcvd)$table["median"] )
> axis( side=1, at=1:21, labels=NA, tcl=-0.3 )
> axis( side=2, at=1:9/10, labels=NA, tcl=-0.3 )
> lines( KMcvd, col=clr, lwd=4 )
> for(j in 0:1) mtext( RTCvd[1:nt+nt*j,2], at= RTCvd[1:nt+nt*j,1],
+                   line=2.5+j, side=1, col=clr[j+1], font=2, cex=1.2 )

```

We can make the same plot the other way round

```

> nt <- nrow(RTall)/2
> clr <- c("forestgreen","red")
> par( mfrow=c(1,2), mar=c(6,3,1,1), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> plot( KMall, conf.int=TRUE, col=clr, lwd=1,
+       yaxs="i", yaxt="n", ylim=c(1,0),
+       ylab="Cumulative mortality" )
> abline( h=0.5, v=summary(KMall)$table["median"] )
> axis( side=1, at=1:21, labels=NA, tcl=-0.3 )
> axis( side=2, at=0:5/5, labels=5:0/5 )
> axis( side=2, at=1:9/10, labels=NA, tcl=-0.3 )
> text( rep(1,2), c(9,5)/100, c("Intensive","Conventional"),
+       col=clr, font=2, cex=1.2, adj=0 )
> lines( KMall, col=clr, lwd=4 )
> for(j in 0:1) mtext( RTall[1:nt+nt*j,2], at= RTall[1:nt+nt*j,1],
+                   line=3.5-j, side=1, col=clr[j+1], font=2, cex=1.2 )
> plot( KMcvd, conf.int=TRUE, col=clr, lwd=1,
+       yaxs="i", yaxt="n", ylim=c(1:0),
+       ylab="Death or CVD event" )
> abline( h=0.5, v=summary(KMcvd)$table["median"] )
> axis( side=1, at=1:21, labels=NA, tcl=-0.3 )
> axis( side=2, at=0:5/5, labels=5:0/5 )
> axis( side=2, at=1:9/10, labels=NA, tcl=-0.3 )
> lines( KMcvd, col=clr, lwd=4 )
> for(j in 0:1) mtext( RTCvd[1:nt+nt*j,2], at= RTCvd[1:nt+nt*j,1],
+                   line=3.5-j, side=1, col=clr[j+1], font=2, cex=1.2 )

```

The `summary.survfit` gives the median survival as well as the confidence intervals for these:

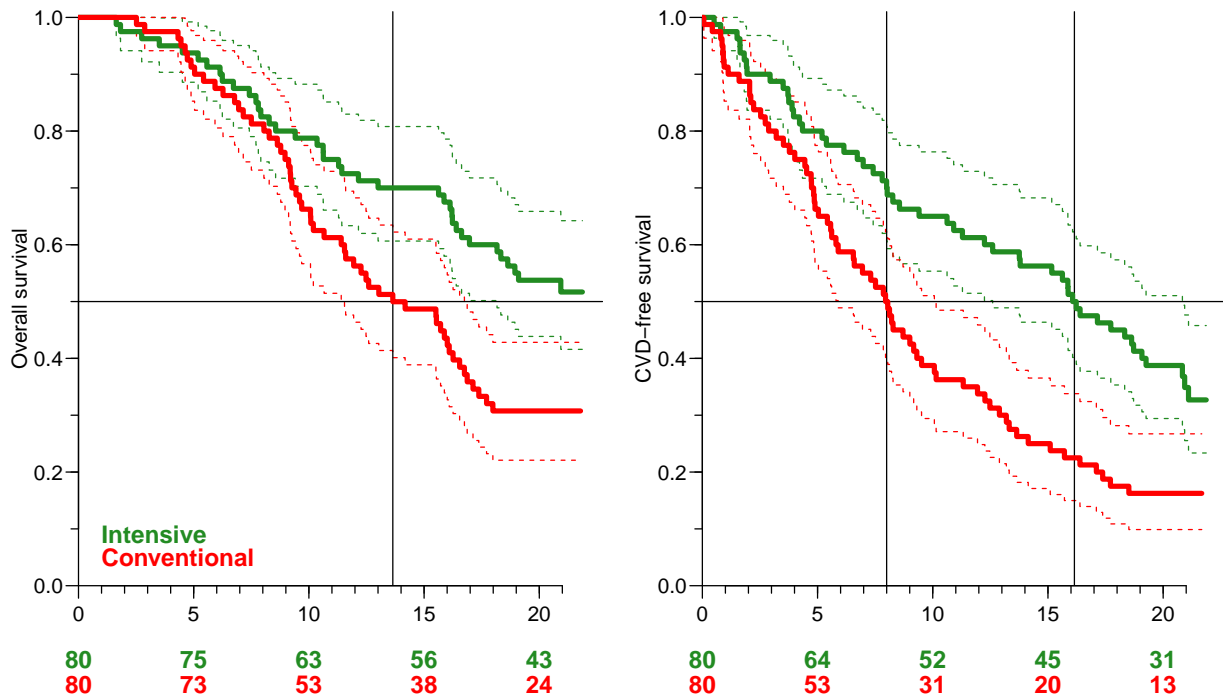


Figure 3.1: Kaplan-Meier estimators for overall and CVD-free survival, numbers at the bottom is the number at risk at times 0, 5, etc.

```
> round( summary( KMall )$table[-(5:6)], 1 )
              records n.max n.start events median 0.95LCL 0.95UCL
allocation=Intensive      80   80    80   38    NA    18.2    NA
allocation=Conventional   80   80    80   55   13.6   11.5   16.8
> round( summary( KMcvd )$table[-(5:6)], 1 )
              records n.max n.start events median 0.95LCL 0.95UCL
allocation=Intensive      80   80    80   52   16.1   12.6   20.8
allocation=Conventional   80   80    80   67    8.0    5.9   10.1
```

However we would like to see the confidence intervals for the *difference* between the groups. This is done by a simple bootstrap of the original dataset; we devise a function that computes the median survival in the two groups for the two types of outcome, for a resampled dataset and we then run this function 5000 times to get a bootstrap sample from the distribution of median survival times:

```
> Difmed <-
+ function()
+ {
+ wh <- sample(1:160,160,replace=TRUE)
+ KMall <- survfit( Surv(tsb,tsb+lex.dur,lex.Xst %in% Xall) ~ allocation,
+                   data = LO[wh,] )
+ KMcvd <- survfit( Surv(tsb,tsb+lex.dur,lex.Xst %in% Xcvd) ~ allocation,
+                   data = Lcvd[wh,] )
+ rbind( summary( KMall )$table[,"median"],
+         summary( KMcvd )$table[,"median"] )
+ }
```

We set up an array to hold the values of the median survival time for the two groups, and separately for the two types of outcome.

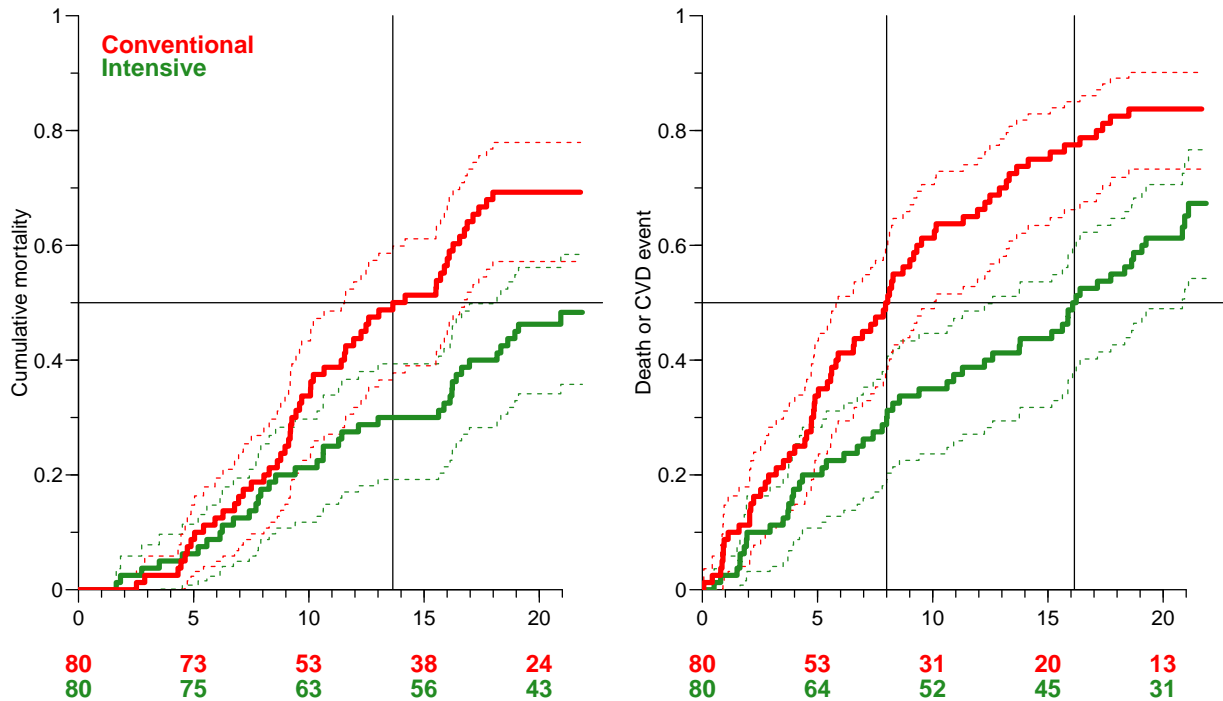


Figure 3.2: Kaplan-Meier estimators for overall and CVD-free survival, numbers at the bottom is the number at risk at times 0, 5, etc.

```
> meds <- NArray( list( 1:5000,
+                       type = c("All","CVD"),
+                       allocation = c("Int","Conv") ) )
> set.seed( 57976895 )
> system.time( for( i in 1:dim(meds)[1] ) meds[i,,] <- Difmed() )
  user system elapsed
 73.997  0.004 73.998
> org <- meds
> meds <- org
> round( apply( meds, 2:3, function(x) mean(is.na(x) ) ) * 100, 2 )
      allocation
type   Int Conv
All  60.80  0
CVD   0.14  0
```

We see that there quite a substantial fraction of the simulations that produce a missing median survival time for the intensive group for all-cause mortality. If we replace these with 21, we get an under-estimate of the median survival, and since the single imputation is mainly for one group also of the difference in median survival times:

```
> meds[is.na(meds)] <- 21
> meds <- meds[,c(1,2,2)]
> dimnames(meds)[[3]][3] <- "I - C"
> dimnames(meds)[-1]
$type
[1] "All" "CVD"
$allocation
[1] "Int" "Conv" "I - C"
```

```

> meds[, ,3] <- meds[, , "Int"] - meds[, , "Conv"]
> round( ftable( apply( meds,
+                     2:3,
+                     quantile,
+                     probs=c(0.5,0.025,0.975) ),
+                     col.vars=c(3,1) ), 1 )

```

	allocation			Int			Conv			I - C		
	50%	2.5%	97.5%	50%	2.5%	97.5%	50%	2.5%	97.5%	50%	2.5%	97.5%
type												
All	21.0	16.8	21.0	13.6	10.8	16.5	6.4	2.2	9.6			
CVD	16.1	12.2	19.7	8.0	5.8	9.5	8.3	4.0	12.6			

Thus we see that the difference in median survival is (at least) 6.4 years (2.2;9.6), whereas the difference in median CVD-free survival is

3.2 All-cause mortality

First we model all-cause mortality as a function of time since randomization separately in the two groups with the intent of inspecting how mortality between the two groups develop.

In terms of the multistate figures in the previous chapter, this analysis corresponds to pooling of all alive states and pooling all dead states, and just analyzing transitions from alive to dead.

3.2.1 Splitting the dataset by time

First we split the dataset in intervals of one month (that is 365.25/12 days) in order to model mortality by time from baseline as a smooth function of time. This is because we shall assume that mortality varies continuously with time since baseline.

```

> library( survival )
> library( Epi )
> library( splines )
> clear()
> print( sessionInfo(), l=F )
R version 3.3.0 (2016-05-03)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Ubuntu 14.04.4 LTS

attached base packages:
[1] splines    utils      datasets  graphics  grDevices  stats      methods    base

other attached packages:
[1] Epi_2.5      survival_2.39-4

loaded via a namespace (and not attached):
 [1] cmprsk_2.2-7    MASS_7.3-44    plyr_1.8.3     Matrix_1.2-1
 [5] parallel_3.3.0  etm_0.6-2      Rcpp_0.11.6    grid_3.3.0
 [9] numDeriv_2014.2-1 lattice_0.20-31

> load( file=" ../data/Lx.Rda" )
> lls()

  name mode class      size
1 L0  list Lexis data.frame 160 22
2 L1  list Lexis data.frame 318 22

> S1 <- splitLexis( L1, breaks=seq(0,25,1/12), time.scale="tsb" )
> summary( S1 )

```

Transitions:

From	To	DM	1st CVD	2nd CVD	3rd CVD	Dead 0	CV-D 0	Dead 1	CV-D 1	Dead 2	CV-D 2	Dead 3
DM		22407	86	0	0	24	9	0	0	0	0	0
1st CVD		0	4120	48	0	0	0	14	13	0	0	0
2nd CVD		0	0	1358	24	0	0	0	0	12	4	0
3rd CVD		0	0	0	1114	0	0	0	0	0	0	5
Sum		22407	4206	1406	1138	24	9	14	13	12	4	5

Transitions:

From	To	CV-D 3	Records:	Events:	Risk time:	Persons:
DM		0	22526	119	1870.64	160
1st CVD		0	4195	75	342.61	86
2nd CVD		0	1398	40	112.27	47
3rd CVD		12	1131	17	92.08	25
Sum		12	29250	251	2417.60	160

Note that the events enumerated in the above summary are *not* the deaths; the outcome of interest in the analysis of all-cause mortality are the transitions to any of the death states:

```
> ( dst <- levels(L1)[- (1:4)] )
[1] "Dead 0" "CV-D 0" "Dead 1" "CV-D 1" "Dead 2" "CV-D 2" "Dead 3" "CV-D 3"
```

We start by modeling mortality by a fairly detailed function of time since baseline (`tsb`), and common linear effects of age and sex between the two groups. First we devise the knots for the `tsb`-effect:

```
> ( t.kn <- with( subset(S1, lex.Xst %in% dst),
+               c(0, quantile( tsb+lex.dur, probs=(1:5-0.5)/5 ) ) )
0.000000  4.648323  7.869678 10.193018 14.711020 17.091855
10%      30%      50%      70%      90%
> m0 <- glm( (lex.Xst %in% dst) ~ allocation + Ns(tsb, kn=t.kn),
+           family = poisson,
+           offset = log(lex.dur),
+           data = S1 )
> mi <- update( m0, . ~ . - Ns(tsb, kn=t.kn) +
+             allocation:Ns(tsb, kn=t.kn) )
> ma <- update( m0, . ~ . + I(age-tsb) + sex )
> mA <- update( m0, . ~ . + age + sex )
> mia <- update( mi, . ~ . + I(age-tsb) + sex )
```

`m0` is the proportional hazards model, `mi` is the model where hazards varies separately for the two randomization groups, `ma` a proportional hazards model with effects of sex and (linear effect of) age at baseline (which, because the linear effect of time since baseline is in the model, will be the same as the effect of current age):

```
> rbind( ci.exp( ma, subset="age" ),
+        ci.exp( mA, subset="age" ) )
exp(Est.)      2.5%      97.5%
I(age - tsb)  1.109305  1.072611  1.147254
age           1.109305  1.072611  1.147254
> c( ma$deviance, mA$deviance )
[1] 1191.187 1191.187
```

With these models fitted we can now test for proportionality of hazards along the time since baseline with and without age and sex included, and we can test for the effect of age and sex with and without the proportional hazards assumption:

```
> zz <- as.matrix( anova( ma, m0, mi, mia, ma, test="Chisq" ) )
> rownames( zz ) <- c( "", "age+sex|ph", "ph|NULL", "age+sex|!ph", "ph|age+sex" )
> round(zz,3)
      Resid. Df Resid. Dev Df Deviance Pr(>Chi)
age+sex|ph    29241  1191.187 NA      NA      NA
ph|NULL       29243  1234.257 -2  -43.070  0.000
age+sex|!ph   29238  1229.505  5    4.752  0.447
age+sex|!ph   29236  1186.012  2   43.492  0.000
ph|age+sex    29241  1191.187 -5   -5.174  0.395
```

We see that there is no evidence whatsoever of non-proportional hazards, so we can compute the estimated HR in the two different proportional hazards models, with and without control for age and sex:

```
> HR.IC <- rbind( ci.exp( m0, subset="allo", pval=TRUE ),
+               ci.exp( ma, subset="allo", pval=TRUE ) )
> rownames(HR.IC) <- c("Allo|ph", "Allo|ph+age+sex")
> round( cbind( HR.IC, 1/HR.IC[,c(1,3,2)] ), 3 )
      exp(Est.) 2.5% 97.5%      P exp(Est.) 97.5% 2.5%
Allo|ph      1.815 1.198 2.750 0.005      0.551 0.364 0.835
Allo|ph+age+sex 1.899 1.249 2.888 0.003      0.526 0.346 0.800
```

We put these estimates in the collect structure for all cause mortality:

```
> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][1]
[1] "All cause mortality"
> zz <- HR.IC
> zz[,1:3] <- 1/HR.IC[,c(1,3,2)]
> mainCI[1,,] <- zz
> round( ftable( mainCI, row.vars=1 ), 3 )
      model      raw      age/sex
      what Estimate 2.5% 97.5%      P Estimate 2.5% 97.5%      P
outcome
All cause mortality      0.551 0.364 0.835 0.005      0.526 0.346 0.800 0.003
CVD mortality            0.379 0.191 0.754 0.006      0.353 0.177 0.705 0.003
non-CVD mortality       0.703 0.413 1.197 0.195      0.686 0.401 1.172 0.168
Death or 1st CVD        NA      NA      NA      NA      NA      NA      NA      NA
Death | CVD state       0.835 0.536 1.301 0.425      0.874 0.564 1.355 0.548
CVD event | CVD state   0.552 0.394 0.772 0.001      0.575 0.412 0.803 0.001
Retinopathy progression 0.668 0.507 0.881 0.004      0.673 0.511 0.887 0.005
Autonomic neuropathy     0.594 0.397 0.890 0.011      0.573 0.382 0.859 0.007
Peripheral neuropathy    1.120 0.707 1.774 0.630      1.101 0.694 1.747 0.683
Macroalbuminuria        0.516 0.316 0.842 0.008      0.495 0.302 0.811 0.005
> save( mainCI, file="../data/mainCI.Rda" )
```

Finally we can extract the estimates of the age and sex effects:

```
> as <- cbind(
+ ci.exp( ma, subset=c("age","sex"), pval=TRUE ),
+ ci.exp( mia, subset=c("age","sex"), pval=TRUE ) )
> colnames(as)[c(1,5)] <- c("effect|ph", "effect|!ph")
> round( as, 3 )
      effect|ph 2.5% 97.5%      P effect|!ph 2.5% 97.5%      P
I(age - tsb)    1.109 1.073 1.147 0.000      1.110 1.073 1.148 0.000
sexM            1.477 0.898 2.428 0.124      1.493 0.908 2.456 0.114
```

We see that the effect of age and sex on mortality is pretty much the same as that in the general Danish population, with an annual increase in mortality by age in the vicinity of 10% and a male/female mortality ratio close to 1.5.

3.2.2 Danish population mortality

To substantiate this, we load the Danish population mortality data (available in the Epi package) and make a quick analysis of mortality in the age-range 45–100 in the period 1995–2012. It is well-known that population all-cause mortality increases exponentially by age after age 40 in most populations and that the M/F mortality RR is almost constant over the age-range, so we fit a simple model to the Danish population data to tease out the population-effects of age and sex:

```
> data( M.dk )
> summary( M.dk )
```

	A	sex	P	D	Y
Min.	: 0.00	Min. :1.0	Min. :1974	Min. : 0.0	Min. : 55.83
1st Qu.:	24.75	1st Qu.:1.0	1st Qu.:1983	1st Qu.: 23.0	1st Qu.:18292.58
Median :	49.50	Median :1.5	Median :1993	Median : 122.0	Median :29515.00
Mean :	49.50	Mean :1.5	Mean :1993	Mean : 284.5	Mean :26267.56
3rd Qu.:	74.25	3rd Qu.:2.0	3rd Qu.:2003	3rd Qu.: 489.0	3rd Qu.:36592.42
Max. :	99.00	Max. :2.0	Max. :2012	Max. :1276.0	Max. :45979.33

```

rate
Min. : 0.0000
1st Qu.: 0.6393
Median : 4.6487
Mean : 52.3246
3rd Qu.: 44.2793
Max. :717.9487

> M.dk <- transform( subset( M.dk, A>44 & P>1994 ),
+                   sex=factor(sex,levels=2:1,labels=c("F","M")) )
> head( M.dk )
```

	A	sex	P	D	Y	rate
3553	45	M	1995	127	37734.50	3.365620
3554	45	F	1995	85	37032.83	2.295261
3555	45	M	1996	128	37342.50	3.427730
3556	45	F	1996	82	36362.83	2.255050
3557	45	M	1997	114	37024.50	3.079042
3558	45	F	1997	92	35947.00	2.559323

```

> mp <- glm( D ~ I(A+0.5) + I(P-1994.5) + sex,
+           offset = log(Y/1000),
+           family = poisson,
+           data = M.dk )
> round( cbind( ci.exp( mp, Exp=F ),
+             ci.exp( mp, pval=T ) ), 4 )
```

	Estimate	2.5%	97.5%	exp(Est.)	2.5%	97.5%	P
(Intercept)	-3.7948	-3.8090	-3.7806	0.0225	0.0222	0.0228	0
I(A + 0.5)	0.0997	0.0995	0.0999	1.1048	1.1046	1.1050	0
I(P - 1994.5)	-0.0197	-0.0200	-0.0193	0.9805	0.9802	0.9809	0
sexM	0.3869	0.3829	0.3910	1.4725	1.4666	1.4784	0

showing that the annual increase in mortality by age is 10%, annual decrease in mortality by calendar time is 2% and the M/F mortality rate-ratio is 1.47. Specifically, the mortality (per 1000 PY) among men aged 55 in 1995 (that is born 1940) as a function of age is:

$$\mu_M(a) = \exp(-3.7948 + 0.0997 \times a - 0.0197 \times (a - 55) + 0.3869)$$

which we shall use to overlay in the display of the observed mortality rates in the Steno 2 population, by computing the population mortality for men in ages 55 and 75 in years 1995 and 2015 respectively — here computed two different ways:

```

> p.mort <- ci.exp( mp, ctr.mat=cbind( c(1, 1),
+                                     c(0,20)+55,
+                                     c(0,20),
+                                     c(1, 1)) )
> ci.pred( mp, data.frame(A = 55+c(0,20)-0.5, # The coding in the data is 0.5 years younger
+                          P = 1995+c(0,20)-0.5, # and 0.5 years earlier than the real time
+                          sex = "M",
+                          Y = 1000) )
  Estimate      2.5%      97.5%
1 7.962197 7.917715 8.006929
2 39.452057 39.251706 39.653430
> p.mort
      exp(Est.)      2.5%      97.5%
[1,] 7.962197 7.917715 8.006929
[2,] 39.452057 39.251706 39.653430

```

Since the modeling of the (log)population rates is linear in age and period we only need the rates at the endpoints 55 years, 1995 and 75 years, 2015 in order to be able to draw the predicted mortality rates as a function of time since baseline.

3.2.3 Mortality curves

We can now plot the two mortality curves and the HR between them; for the sex-age-controlled model we use men aged 55 to show the mortality rates (the majority of patients are men and the median age at baseline is 56.6 years):

```

> t.pt <- seq(0,20,0.1)
> CM <- Ns( t.pt, kn=t.kn )
> nd <- data.frame(tsb=t.pt,lex.dur=1000,age=55+t.pt,sex="M")
> rS.t <- ci.pred( mi, newdata=cbind(nd,allocation="Conventional") )
> rI.t <- ci.pred( mi, newdata=cbind(nd,allocation="Intensive") )
> rS.a <- ci.pred( mia,newdata=cbind(nd,allocation="Conventional") )
> rI.a <- ci.pred( mia,newdata=cbind(nd,allocation="Intensive") )
> pS.t <- ci.pred( m0, newdata=cbind(nd,allocation="Conventional") )
> pI.t <- ci.pred( m0, newdata=cbind(nd,allocation="Intensive") )
> pS.a <- ci.pred( ma, newdata=cbind(nd,allocation="Conventional") )
> pI.a <- ci.pred( ma, newdata=cbind(nd,allocation="Intensive") )
> # Check parameter sequencing using subset=
> round( ci.exp( mi , subset=c("Intensive","Conventional") ), 3 )
      exp(Est.)      2.5%      97.5%
allocationIntensive:Ns(tsb, kn = t.kn)1      9.259 0.974      88.033
allocationIntensive:Ns(tsb, kn = t.kn)2      2.877 0.143      57.847
allocationIntensive:Ns(tsb, kn = t.kn)3      4.478 0.671      29.895
allocationIntensive:Ns(tsb, kn = t.kn)4     14.891 0.085     2603.902
allocationIntensive:Ns(tsb, kn = t.kn)5      3.044 0.967      9.582
allocationConventional      0.136 0.002      10.014
allocationConventional:Ns(tsb, kn = t.kn)1    72.673 2.391     2209.022
allocationConventional:Ns(tsb, kn = t.kn)2    88.787 1.539     5121.615
allocationConventional:Ns(tsb, kn = t.kn)3    52.265 3.398      803.813
allocationConventional:Ns(tsb, kn = t.kn)4   2736.016 1.193   6274265.608
allocationConventional:Ns(tsb, kn = t.kn)5      5.225 1.569      17.397
> RR.t <- ci.exp( mi , subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )
> RR.a <- ci.exp( mia, subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )

```

Note that since we assume a proportional hazards model by age and sex, age at baseline and sex are not part of the RR calculations — the assumption is that the intervention effect is the same across all ages and for both sexes.

We now have the predicted mortality rates and the estimated HRs from the two models, so we can plot them side by side and see to what extent adjustment for age at baseline influences the estimates. First we define the colors and the corresponding transparent colors:


```

> clr <- c("forestgreen","red","black",gray(0.6))
> slr <- rgb( t( col2rgb(clr[1:2]) ), alpha=40, max=255 )
> par( mfrow=c(1,2), mar=c(0,0,0,0), oma=c(3,4,1,2), mgp=c(3,1,0)/1.6 )
> yl <- c(0.1,250)
> matplot( t.pt, cbind(rI.t,rS.t,RR.t),#pI.t,pS.t),
+         log="y", ylim=yl, xlab="",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,
+         lwd=c(3,1,1), col=rep(clr[1:3],each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[1,2:3], col=clr[3], lty=1 )
> abline( h=1/HR.IC[1, 1], col=clr[3], lwd=3, lty=1 )
> matlines( t.pt, cbind(pI.t,pS.t),
+          type="l", lty=1, lwd=c(3,1,1), col=rep(clr[1:2],each=3) )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),#pI.a,pS.a),
+         log="y", ylim=yl, yaxt="n", ylab="",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,
+         lwd=c(3,1,1), col=rep(clr[1:3],each=3) )
> matlines( t.pt, cbind(pI.a,pS.a),
+          type="l", lty=1, lwd=c(3,1,1), col=rep(clr[1:2],each=3) )
> lines( c(0,20), p.mort[,1], lwd=3, lty=1, col=clr[4] )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2,2:3], col=clr[3], lty=1 )
> abline( h=1/HR.IC[2, 1], col=clr[3], lwd=3, lty=1 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( rep(11,3), 10*0.70^c(2,1,3,4), c("Intensive","Conventional","Rate ratio","Population"),
+       col=clr, font=2, cex=1.2, adj=0 )
> mtext( "Mortality rate per 1000 PY / Hazard ratio Intensive vs Conventional",
+       side=2, line=3, las=0, outer=TRUE )

```

In order to plot the rightmost panel for the ESM together with the same for the CVD mortality we save the relevant structures:

```

> save( t.pt, HR.IC,
+       rI.t, rS.t,
+       rI.a, rS.a,
+       pI.t, pS.t, RR.t,
+       pI.a, pS.a, RR.a,
+       file = "../data/all-mort.Rda" )

```

From figure 3.3 it is seen that the intervention effect on overall mortality is not formally manifest till some 10 years after baseline, and there also seems to be a diminishing effect some 10 years after end of intervention. Comparison with the population rates shows that the intensive group has a mortality that is roughly mid-way between that of the conventional group and the population mortality.

Also, although there is no formal interaction between time since randomization and treatment group, when using the interaction model, the mortality in the intensive group tends to be larger in the first 2 years, but with very large confidence intervals for the HR.

3.2.4 DM duration

It is of some interest to assess to what extent diabetes duration has an effect on mortality, and in particular if the intervention as claimed by some is potentially harmful for persons with long duration of diabetes at entry. This amounts to investigation of the effect of duration and its interaction with allocation. Thus we expand the model for total mortality with an effect of duration, and further with duration terms separately for the two intervention groups, and finally with quadratic terms of duration for the two groups, to allow more flexible differences between duration effects in the two randomization groups.

```

> round( ci.exp(ma), 3 )

```

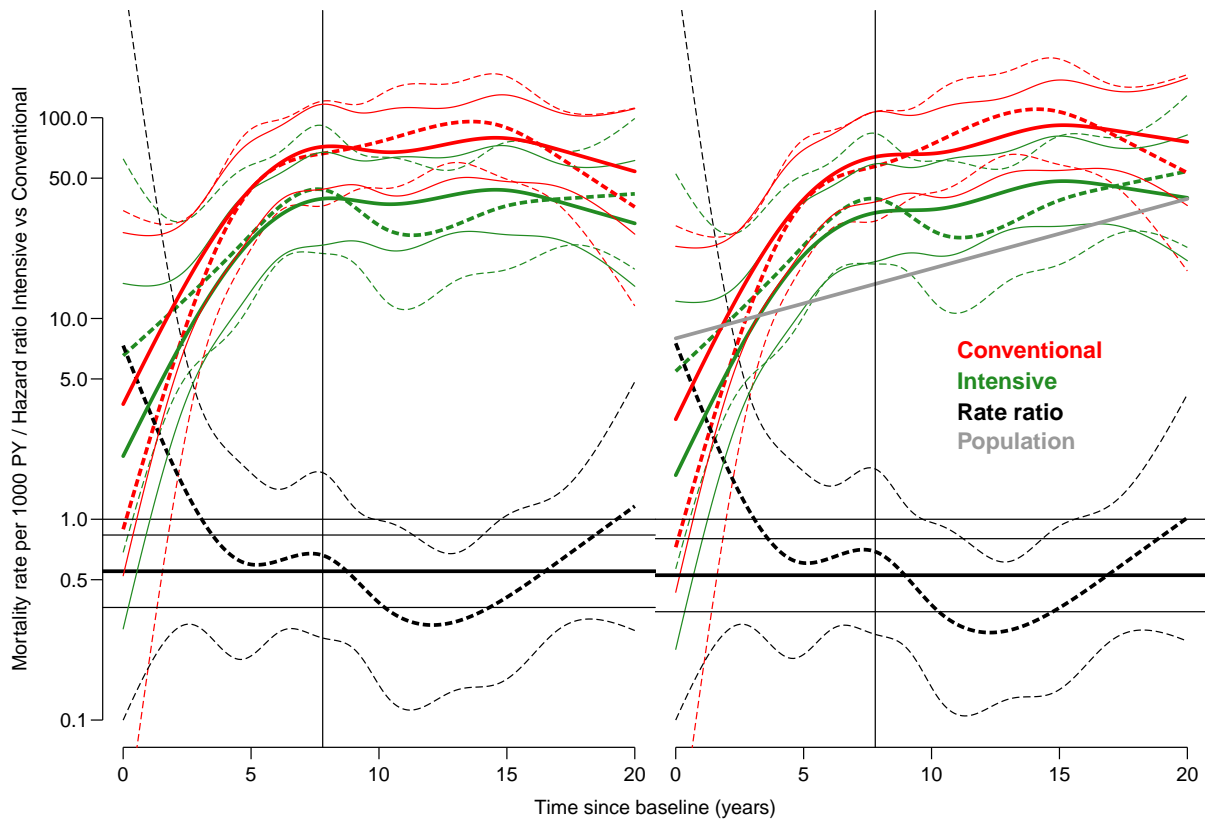


Figure 3.3: Overall mortality rates and hazard ratio in the two groups. The full lines are hazards and HRs assuming proportional hazards, and the vertical line indicates the intervention end, broken lines are the hazards without the proportional hazards assumption. Left panel is models with only group and time since baseline, the models in the right panel also includes sex and age at baseline as a linear term on the log-mortality scale, and mortality rates are shown for a man aged 55 at baseline. The full black line is the population mortality for men aged 55 at start, aging to 75, over the period 1995–2015.

```

exp(Est.)  2.5%    97.5%
(Intercept)          0.000 0.000  0.000
allocationConventional  1.899 1.249  2.888
Ns(tsb, kn = t.kn)1   21.956 3.362 143.382
Ns(tsb, kn = t.kn)2   19.638 1.908 202.077
Ns(tsb, kn = t.kn)3   18.422 3.960  85.697
Ns(tsb, kn = t.kn)4  235.790 3.143 17691.556
Ns(tsb, kn = t.kn)5    5.720 2.573  12.717
I(age - tsb)          1.109 1.073  1.147
sexM                  1.477 0.898  2.428
> md <- update( ma , . ~ . + I( dur-tsb ) )
> md1 <- update( ma , . ~ . + allocation:I( dur-tsb ) )
> md2 <- update( md1, . ~ . + allocation:I((dur-tsb)^2) )
> anova( ma, md, md1, md2, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  1  3.03918  0.08128
3  1  1.13584  0.28653
4  2  0.52607  0.76871

```

Thus we see that diabetes duration at baseline has a marginally significant effect ($p=0.081$), but no evidence of differential effect between the intensive and conventional groups. The estimated trend by duration, jointly and separately are here:

```

> round( ci.exp( md , subset=c("all","dur") ), 4 )
              exp(Est.)  2.5%  97.5%
allocationConventional  1.7976 1.1772 2.7451
I(dur - tsb)           1.0342 0.9968 1.0731

> round( ci.exp( md1, subset= "all"           ), 4 )
              exp(Est.)  2.5%  97.5%
allocationConventional  2.4691 1.1949 5.1021
allocationIntensive:I(dur - tsb)  1.0594 1.0012 1.1209
allocationConventional:I(dur - tsb)  1.0189 0.9717 1.0683

> round( ( rbind( ci.exp( md , subset="dur" ),
+               ci.exp( md1, subset="dur" ) )-1)*100, 1 )
              exp(Est.) 2.5% 97.5%
I(dur - tsb)          3.4 -0.3  7.3
allocationIntensive:I(dur - tsb)  5.9 0.1 12.1
allocationConventional:I(dur - tsb)  1.9 -2.8  6.8

```

We see a borderline significant effect of duration (HR=3.4%/year (-0.3,7.3), $p=0.081$), but no significant difference in this between the two groups, $p=0.286$, although there is a very weak indication that DM duration at baseline has different effects in the two groups. In the intensive group there is an increase of mortality of 5.2%/year of DM duration, in the conventional group only 1.9%/year, not significantly different though.

It is however more informative to see the HR between intensive and conventional by duration as predicted by the two interaction models (the linear and the quadratic):

```

> CD <- cbind( -1, 0:20, -(0:20), (0:20)^2, -(0:20)^2 )
> rownames(CD) <- 0:20
> head( CD )
  [,1] [,2] [,3] [,4] [,5]
0  -1   0   0   0   0
1  -1   1  -1   1  -1
2  -1   2  -2   4  -4
3  -1   3  -3   9  -9
4  -1   4  -4  16 -16
5  -1   5  -5  25 -25

> round( cbind( RRl <- ci.exp( md1, subset="all", ctr.mat=CD[,1:3], pval=TRUE ),
+             RRq <- ci.exp( md2, subset="all", ctr.mat=CD           , pval=TRUE ) ), 4 )
  exp(Est.)  2.5%  97.5%  P exp(Est.)  2.5%  97.5%  P
0  0.4050 0.1960 0.8369 0.0147 0.4428 0.1620 1.2102 0.1123
1  0.4211 0.2158 0.8218 0.0112 0.4451 0.1921 1.0312 0.0590
2  0.4379 0.2369 0.8094 0.0084 0.4491 0.2225 0.9066 0.0255
3  0.4553 0.2590 0.8003 0.0063 0.4548 0.2506 0.8255 0.0096
4  0.4734 0.2818 0.7953 0.0047 0.4623 0.2736 0.7812 0.0039
5  0.4922 0.3046 0.7954 0.0038 0.4718 0.2895 0.7689 0.0026
6  0.5118 0.3266 0.8020 0.0035 0.4832 0.2981 0.7832 0.0032
7  0.5321 0.3467 0.8166 0.0039 0.4968 0.3018 0.8178 0.0059
8  0.5533 0.3640 0.8410 0.0056 0.5126 0.3033 0.8665 0.0126
9  0.5753 0.3775 0.8766 0.0101 0.5310 0.3050 0.9246 0.0253
10 0.5981 0.3869 0.9247 0.0208 0.5521 0.3082 0.9888 0.0457
11 0.6219 0.3923 0.9860 0.0434 0.5762 0.3139 1.0575 0.0752
12 0.6467 0.3941 1.0610 0.0844 0.6036 0.3223 1.1304 0.1148
13 0.6724 0.3930 1.1502 0.1473 0.6346 0.3332 1.2089 0.1667
14 0.6991 0.3897 1.2542 0.2300 0.6698 0.3461 1.2962 0.2342
15 0.7269 0.3846 1.3737 0.3260 0.7096 0.3601 1.3983 0.3216
16 0.7558 0.3783 1.5098 0.4278 0.7546 0.3736 1.5242 0.4325
17 0.7859 0.3711 1.6640 0.5290 0.8055 0.3846 1.6870 0.5663
18 0.8171 0.3633 1.8378 0.6253 0.8630 0.3909 1.9053 0.7154
19 0.8496 0.3550 2.0333 0.7143 0.9281 0.3908 2.2043 0.8658
20 0.8834 0.3464 2.2525 0.7951 1.0019 0.3832 2.6193 0.9969

```

```

> par( mar=c(3,3,1,1), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> matplot( 0:20, cbind( RRI[, -4], RRq[, -4] ),
+         log="y", type="l", lwd=c(4,1,1), lty=1,
+         col=rep(c("black", "gray"), each=3), ylim=c(0.2, 2),
+         xlab="Diabetes duration at baseline",
+         ylab="Mortality HR: Intensive vs Conventional" )
> abline( h=1 )

> CD <- cbind( -1, 0:20, -(0:20), (0:20)^2, -(0:20)^2 )
> rownames(CD) <- 0:20
> head( CD )
  [,1] [,2] [,3] [,4] [,5]
0  -1   0   0   0   0
1  -1   1  -1   1  -1
2  -1   2  -2   4  -4
3  -1   3  -3   9  -9
4  -1   4  -4  16 -16
5  -1   5  -5  25 -25
> round( cbind( RRI <- ci.exp( md1, subset="all", ctr.mat=CD[, 1:3], pval=TRUE ),
+             RRq <- ci.exp( md2, subset="all", ctr.mat=CD
+             , pval=TRUE ) ), 4 )
  exp(Est.)  2.5% 97.5%   P exp(Est.)  2.5% 97.5%   P
0    0.4050 0.1960 0.8369 0.0147  0.4428 0.1620 1.2102 0.1123
1    0.4211 0.2158 0.8218 0.0112  0.4451 0.1921 1.0312 0.0590
2    0.4379 0.2369 0.8094 0.0084  0.4491 0.2225 0.9066 0.0255
3    0.4553 0.2590 0.8003 0.0063  0.4548 0.2506 0.8255 0.0096
4    0.4734 0.2818 0.7953 0.0047  0.4623 0.2736 0.7812 0.0039
5    0.4922 0.3046 0.7954 0.0038  0.4718 0.2895 0.7689 0.0026
6    0.5118 0.3266 0.8020 0.0035  0.4832 0.2981 0.7832 0.0032
7    0.5321 0.3467 0.8166 0.0039  0.4968 0.3018 0.8178 0.0059
8    0.5533 0.3640 0.8410 0.0056  0.5126 0.3033 0.8665 0.0126
9    0.5753 0.3775 0.8766 0.0101  0.5310 0.3050 0.9246 0.0253
10   0.5981 0.3869 0.9247 0.0208  0.5521 0.3082 0.9888 0.0457
11   0.6219 0.3923 0.9860 0.0434  0.5762 0.3139 1.0575 0.0752
12   0.6467 0.3941 1.0610 0.0844  0.6036 0.3223 1.1304 0.1148
13   0.6724 0.3930 1.1502 0.1473  0.6346 0.3332 1.2089 0.1667
14   0.6991 0.3897 1.2542 0.2300  0.6698 0.3461 1.2962 0.2342
15   0.7269 0.3846 1.3737 0.3260  0.7096 0.3601 1.3983 0.3216
16   0.7558 0.3783 1.5098 0.4278  0.7546 0.3736 1.5242 0.4325
17   0.7859 0.3711 1.6640 0.5290  0.8055 0.3846 1.6870 0.5663
18   0.8171 0.3633 1.8378 0.6253  0.8630 0.3909 1.9053 0.7154
19   0.8496 0.3550 2.0333 0.7143  0.9281 0.3908 2.2043 0.8658
20   0.8834 0.3464 2.2525 0.7951  1.0019 0.3832 2.6193 0.9969
> par( mfrow=c(1,2), mar=c(3,3,1,1), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> hist( LO$dur, breaks=seq(0,32,2), col="black", main="",
+       xlab="Diabetes duration at baseline" )
> matplot( 0:20, cbind( RRI[, -4], RRq[, -4] ),
+         log="y", type="l", lwd=c(4,1,1), lty=1, col="black", ylim=c(0.2, 2),
+         xlab="Diabetes duration at baseline",
+         ylab="HR: Intensive vs Conventional" )
> abline( h=1 )

> round( RRI[seq(1,21,5),], 2 )
  exp(Est.) 2.5% 97.5%   P
0    0.41 0.20 0.84 0.01
5    0.49 0.30 0.80 0.00
10   0.60 0.39 0.92 0.02
15   0.73 0.38 1.37 0.33
20   0.88 0.35 2.25 0.80

```

Thus the linear effect model corresponds to a HR between intensive and standard of 0.41 at baseline duration 0; 0.49 at 5 years; 0.60 at 10 years and 0.88 at 20 years - only significantly less than 1 for durations less than 10 years. Very similar effect is seen for the quadratic extension; figure ???. In conclusion, there is no significant effect of duration at baseline and neither any sign of differential intervention effect by diabetes duration.

Hence we shall not explore this further.

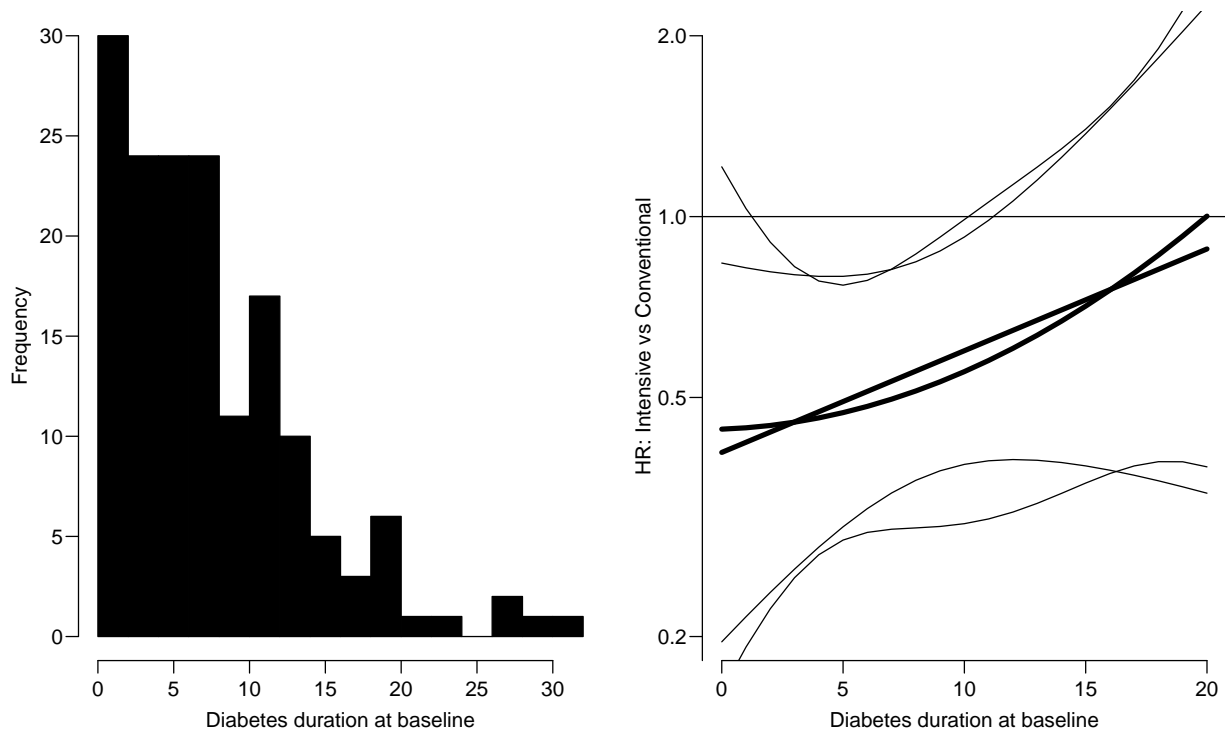


Figure 3.4: *Left: Distribution of duration of diabetes at baseline. Right: Hazard ratio between intensive and conventional by diabetes duration at baseline, estimated from two different interaction models, one with log-linear effects and another with log-quadratic effects of DM duration on HR.*

3.2.5 Survival

The analyses of mortality rates immediately translate into survival curves, the model with age-correction of course to survival curves for different ages at baseline — for the latter we shall use age 45,50,...,65; corresponding approximately to the 10,30,50,70 and 90 age-percentiles at baseline, and of course give estimates separately for men and women:

```
> round( quantile( L0$age, probs=1:9/10 ), 1 )
 10% 20% 30% 40% 50% 60% 70% 80% 90%
45.4 47.4 50.0 54.1 56.6 57.9 60.4 62.2 64.2
> round( do.call( rbind,
+               with( L0, tapply( age, sex,
+                               quantile, probs=seq(1,9,2)/10 ) ) ), 1 )
      10% 30% 50% 70% 90%
F 46.1 50.3 56.8 60.7 64.6
M 45.3 50.0 56.6 60.1 63.5
```

We initially use the model `mi` from above; it is a model with a separate mortality for each group, modelled by a smooth function of time since baseline. Thus it is *not* a proportional hazards model; it is a model that basically models the mortality rates for each group separately, pretty much the machinery used to generate Kaplan-Meier curves for each group.

First we check that we get what we want from the `subset=` argument to `ci.exp`, and then we use it to compute the cumulative hazards with confidence bands

```

> ci.exp( mi, subset=c("cept","ensive") )
              exp(Est.)      2.5%      97.5%
(Intercept) 0.006534735 0.0006839742 0.0624333
allocationIntensive:Ns(tsb, kn = t.kn)1 9.258952868 0.9738221988 88.0327110
allocationIntensive:Ns(tsb, kn = t.kn)2 2.876667150 0.1430540711 57.8467556
allocationIntensive:Ns(tsb, kn = t.kn)3 4.477914618 0.6707393510 29.8949499
allocationIntensive:Ns(tsb, kn = t.kn)4 14.891396861 0.0851620683 2603.9022416
allocationIntensive:Ns(tsb, kn = t.kn)5 3.043918250 0.9669421010 9.5822059
> ci.exp( mi, subset=c("cept","tional") )
              exp(Est.)      2.5%      97.5%
(Intercept) 6.534735e-03 0.0006839742 6.243330e-02
allocationConventional 1.364352e-01 0.0018588457 1.001404e+01
allocationConventional:Ns(tsb, kn = t.kn)1 7.267304e+01 2.3908186795 2.209022e+03
allocationConventional:Ns(tsb, kn = t.kn)2 8.878695e+01 1.5391867900 5.121615e+03
allocationConventional:Ns(tsb, kn = t.kn)3 5.226498e+01 3.3983404357 8.038125e+02
allocationConventional:Ns(tsb, kn = t.kn)4 2.736016e+03 1.1930935186 6.274266e+06
allocationConventional:Ns(tsb, kn = t.kn)5 5.225328e+00 1.5694436068 1.739728e+01
> ( intl <- mean(diff(t.pt)) )
[1] 0.1
> chI <- ci.cum( mi, subset=c("cept","ensive"), ctr.mat=cbind(1, CM), intl=intl, ci.Exp=TRUE )
> chC <- ci.cum( mi, subset=c("cept","tional"), ctr.mat=cbind(1,1,CM), intl=intl, ci.Exp=TRUE )
> round( cbind(t.pt,chI,chC)[20+1:10,], 5 )
      t.pt Estimate      2.5%      97.5%      Erf Estimate      2.5%      97.5%      Erf
[1,] 2.0 0.01810 0.00417 0.07858 4.34025 0.00590 0.00065 0.05371 9.10043
[2,] 2.1 0.01925 0.00460 0.08056 4.18584 0.00657 0.00078 0.05541 8.42769
[3,] 2.2 0.02042 0.00505 0.08248 4.03948 0.00731 0.00094 0.05708 7.81007
[4,] 2.3 0.02162 0.00554 0.08435 3.90089 0.00811 0.00112 0.05875 7.24392
[5,] 2.4 0.02286 0.00606 0.08619 3.76979 0.00898 0.00134 0.06041 6.72569
[6,] 2.5 0.02414 0.00662 0.08800 3.64592 0.00993 0.00159 0.06209 6.25198
[7,] 2.6 0.02545 0.00721 0.08980 3.52901 0.01096 0.00188 0.06380 5.81952
[8,] 2.7 0.02679 0.00784 0.09159 3.41881 0.01208 0.00223 0.06554 5.42521
[9,] 2.8 0.02817 0.00850 0.09340 3.31506 0.01329 0.00262 0.06735 5.06610
[10,] 2.9 0.02960 0.00920 0.09523 3.21751 0.01460 0.00308 0.06922 4.73945

```

We now have the cumulative hazards evaluated at the points `t.pt` for the two groups (with confidence intervals), and we can thus change them into survival curves with `c.i.` which we plot with shaded confidence bands:

```

> pls <-
+ function(yaxt="s",ci=TRUE,xl="Time since baseline (years)")
+ {
+ matplot( t.pt, exp( -cbind(chI[,1],chC[,1]) ),
+          col=clr[1:2], lty=1, lwd=4, type="l",
+          yaxt=yaxt, yaxs="i", ylim=c(0,1), ylab="",
+          xlim=c(0,20), xaxs="i", xlab=xl )
+ if(ci)
+ {
+ polygon( c(t.pt,rev(t.pt)), exp(-c(chI[,2],rev(chI[,3])) ), col=slr[1], border="transparent" )
+ polygon( c(t.pt,rev(t.pt)), exp(-c(chC[,2],rev(chC[,3])) ), col=slr[2], border="transparent" )
+ }
+ }
> par( mar=c(3,3,1,2), bty="n", mgp=c(3,1,0)/1.6, las=1 )
> pls()
> axis( side=4, at=0:5*20 /100)
> axis( side=4, at=0:10*10/100, tcl=-0.4, labels=NA )
> axis( side=4, at=0:20*5 /100, tcl=-0.3, labels=NA )
> axis( side=4, at=0:100 /100, tcl=-0.2, labels=NA )
> mtext( "Survival", side=2, las=0, line=2 )

```

We can compare the smoothed version of the survival curve with the Kaplan-Meier-estimates for the two groups, and we see that they are pretty much alike:

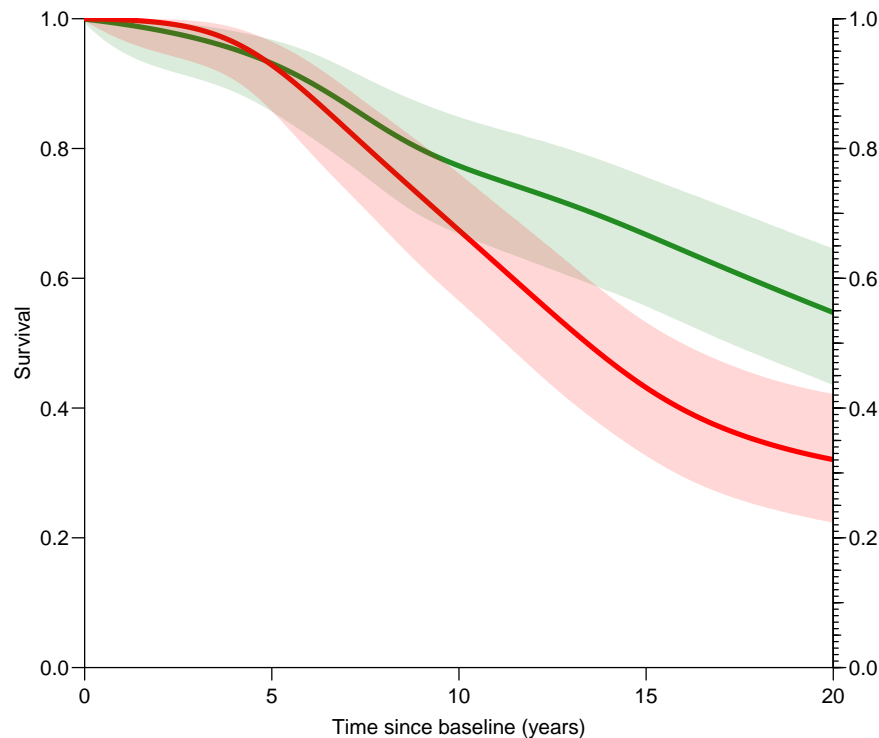


Figure 3.5: Overall survival in the Steno 2 study. The smooth curves are from a model assuming a smoothly varying mortality rate by time since baseline.

```
> par( mfrow=c(1,2), mar=c(3,0,1,1), oma=c(0,3,0,0), bty="n",
+      mgp=c(3,1,0)/1.6, las=1 )
> km <- survfit( Surv(tsb,tsb+lex.dur,lex.Xst %in% dst) ~ allocation, data=L1 )
> pls(ci=FALSE)
> mtext( "Survival", side=2, las=0, line=2 )
> abline( v=seq(0,20,5), h=0:10/10, col=gray(0.8) )
> par( new=TRUE )
> pls()
> pls(yaxt="n")
> lines(km,col=clr[1:2],mark.time=FALSE,conf.int=FALSE,lwd=2)
> lines(km,col=clr[1:2],mark.time=FALSE,conf.int="only",lwd=1,lty=3)
```

The curves in figure 3.6 refer to survival in a population of type 2 DM patients with an age-composition as the patient population in the Steno 2 study. The right hand panel has overlaid the traditional Kaplan-Meier curves that estimate the same quantities (the proportion of the patient population surviving to a given time) but clearly in a clinically much less credible form.

The difference between the intensive and standard groups is best summarized by the HR, either as constant or as time-varying as shown in figure 3.3. The *absolute* difference between regimens only makes sense if referred to a specific age and sex, because these two variables are such powerful predictors of mortality.

We therefore show the *estimated* survival for men and women entering the study at ages 45, 50, 55, 60 and 65 in order to make the absolute magnitudes of the survival probabilities interpretable. This will also make it possible to show the intervention effect relative to the general aging effect:

```
> # Array to collect estimated survival probabilities
> tabSurv <- NArray( list( grp = c("Int","Cnv"),
```

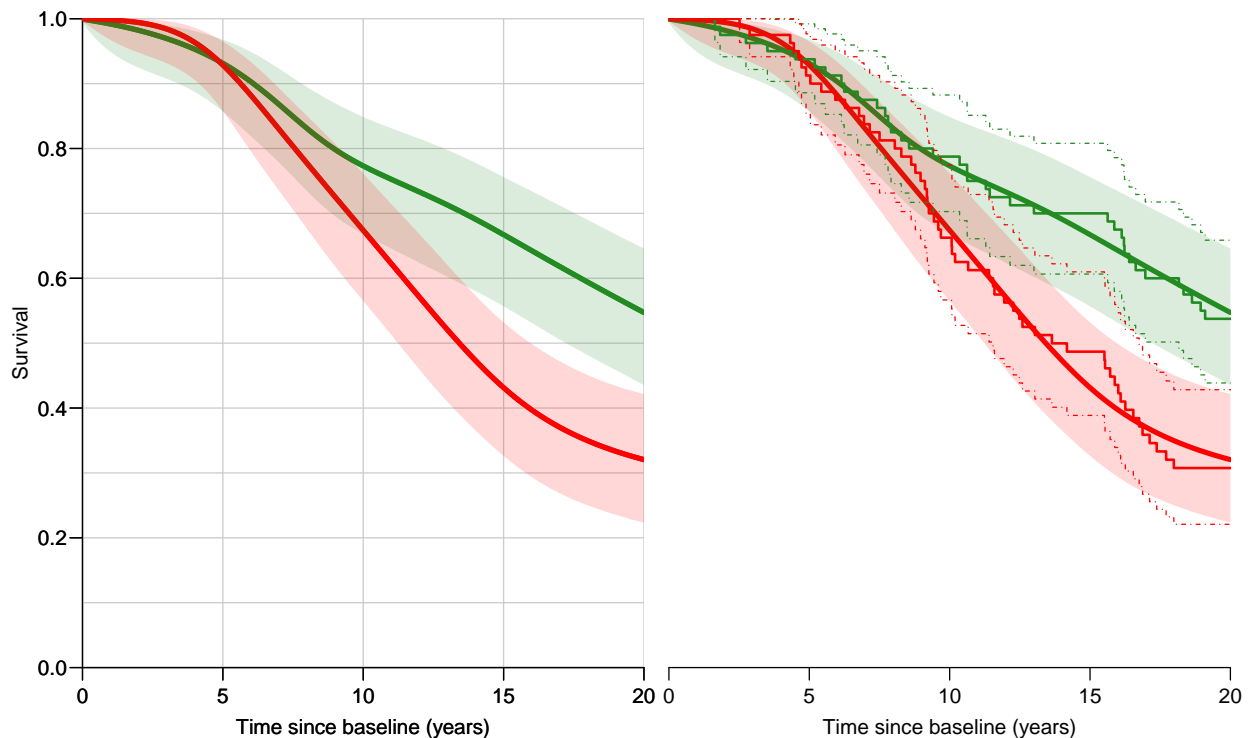


Figure 3.6: Overall survival in the Steno 2 study. The smooth curves with shaded confidence bands (identical in the two panels) are from a model assuming a smoothly varying mortality rate by time since baseline, derived from the mortalities shown in the left panel of figure 3.3. The ragged curves overlaid in the right panel are the traditional Kaplan-Meier estimates, conveying a lot of unsubstantiated noise to the survival curves.

```

+           sex = c("M", "F"),
+           age = seq(45,65,5),
+           tfr = t.pt,
+           what = c("est", "lo", "up") ) )
> ci.exp( mia, subset=c("sex", "age", "cept", "sive") )
              exp(Est.)      2.5%      97.5%
sexM          1.493121e+00  9.077945e-01  2.455854e+00
I(age - tsb)  1.110024e+00  1.073230e+00  1.148079e+00
(Intercept)  1.174012e-05  5.527660e-07  2.493467e-04
allocationIntensive:Ns(tsb, kn = t.kn)1  9.834423e+00  1.029889e+00  9.390903e+01
allocationIntensive:Ns(tsb, kn = t.kn)2  3.356022e+00  1.663345e-01  6.771227e+01
allocationIntensive:Ns(tsb, kn = t.kn)3  5.399008e+00  8.080748e-01  3.607251e+01
allocationIntensive:Ns(tsb, kn = t.kn)4  2.075319e+01  1.179995e-01  3.649973e+03
allocationIntensive:Ns(tsb, kn = t.kn)5  4.071501e+00  1.291699e+00  1.283357e+01
> ci.exp( mia, subset=c("sex", "age", "cept", "onal") )
              exp(Est.)      2.5%      97.5%
sexM          1.493121e+00  9.077945e-01  2.455854e+00
I(age - tsb)  1.110024e+00  1.073230e+00  1.148079e+00
(Intercept)  1.174012e-05  5.527660e-07  2.493467e-04
allocationConventional
allocationConventional:Ns(tsb, kn = t.kn)1  7.324541e+01  2.348622e+00  2.284271e+03
allocationConventional:Ns(tsb, kn = t.kn)2  1.143154e+02  1.931006e+00  6.767462e+03
allocationConventional:Ns(tsb, kn = t.kn)3  7.235271e+01  4.618208e+00  1.133538e+03
allocationConventional:Ns(tsb, kn = t.kn)4  4.615768e+03  1.904531e+00  1.118665e+07
allocationConventional:Ns(tsb, kn = t.kn)5  7.922665e+00  2.358872e+00  2.660959e+01
> par( mfrow=c(2,5), mar=c(3,0,1,1), oma=c(2,4,2,0) )
> for( sx in 1:0 )
+ for( a in seq(45,65,5) )

```



```

+   {
+   chI <- ci.cum( mia, subset=c("sex","age","cept","sive"),
+                 ctr.mat=cbind(sx,a,1, CM), intl=intl, ci.Exp=TRUE )
+   chC <- ci.cum( mia, subset=c("sex","age","cept","onal"),
+                 ctr.mat=cbind(sx,a,1,1,CM), intl=intl, ci.Exp=TRUE )
+   tabSurv["Int",2-sx,paste(a),,] <- exp(-chI[,c(1,3,2)])
+   tabSurv["Cnv",2-sx,paste(a),,] <- exp(-chC[,c(1,3,2)])
+   pls(yaxt=if(a>47) "n" else "s",xl="")
+   if( sx==1 )
+   text( 18, 0.99, paste(a), font=1, adj=c(1,1), cex=1.5 )
+   abline(h=1:19/20,col="white")
+   }
> mtext( "Survival", side=2, line=2, las=0, outer=TRUE )
> mtext( "Time since baseline (years)", side=1, line=0, las=0, outer=TRUE )
> mtext( "Age at entry", side=3, line=0, las=0, outer=TRUE )
> mtext( "Men", at=0.75, side=2, line=2, las=0, outer=TRUE )
> mtext( "Women", at=0.25, side=2, line=2, las=0, outer=TRUE )

```

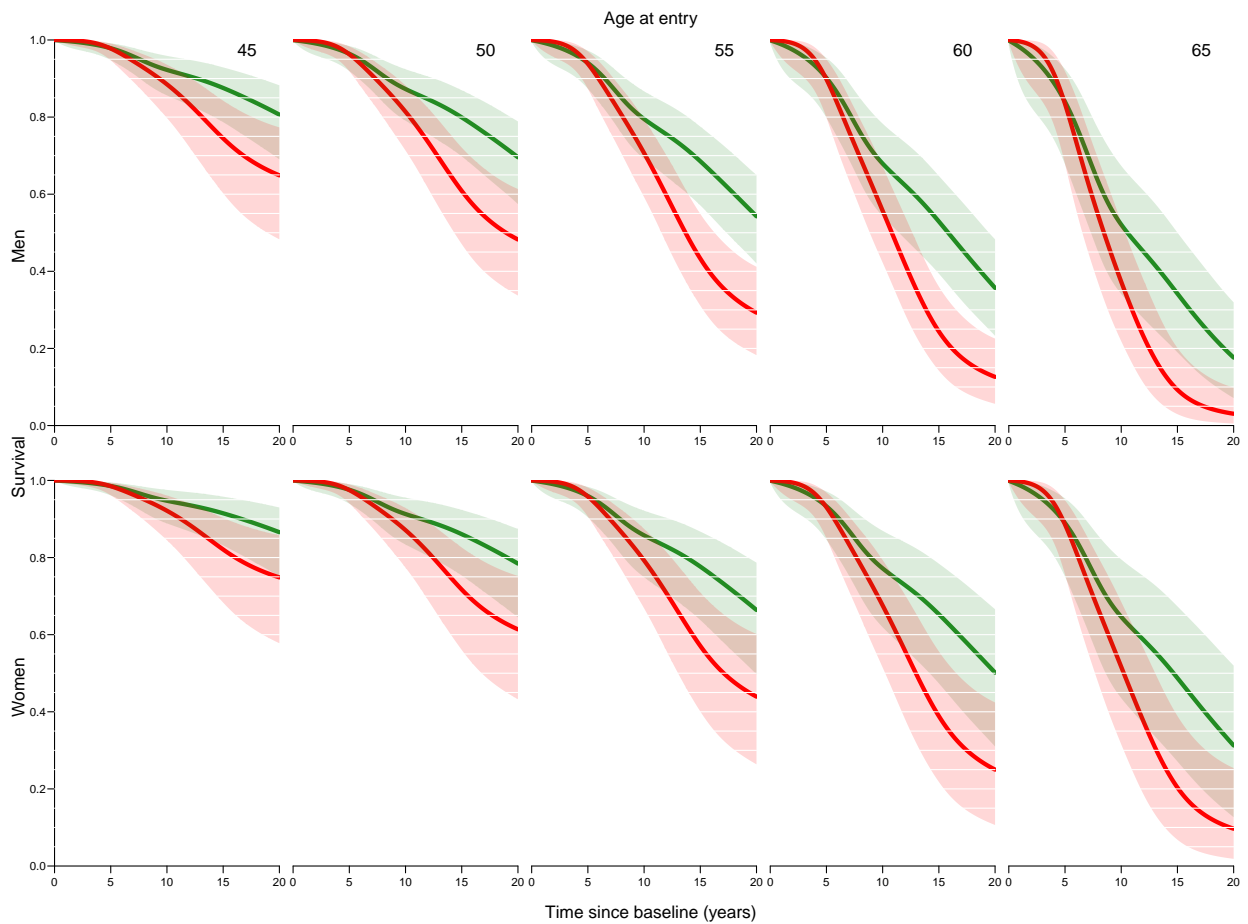


Figure 3.7: The effect of intervention and age on survival. Estimated survival probabilities with 95% c.i. for patients entering the study in different ages.

3.2.6 Change in mortality after intervention

Formally we have already tested whether the hazard ratio changed at the end of the intervention period — it was the test of proportionality of hazard rates between the groups,

but we could of course do a more naive test by expanding the model with proportional hazards with an interaction between allocation and an indicator of intervention period:

```
> ms <- update( m0, . ~ . + I(tsb>7.8 & allocation=="Conventional") )
> round( ci.exp( ms, subset="Conventional" ), 3 )
```

	exp(Est.)	2.5%	97.5%
allocationConventional	1.149	0.567	2.326
I(tsb > 7.8 & allocation == "Conventional")TRUE	1.932	0.850	4.393

```
> zz <- ci.exp( ms, subset="Conventional",
+             ctr.mat=rbind("0-7.8"=c(1,0),
+                          " 7.8+"=c(1,1),
+                          "ratio"=c(0,1)) )
> round( cbind( zz, 1/zz[,c(1,3,2)] ), 3 )
```

	exp(Est.)	2.5%	97.5%	exp(Est.)	97.5%	2.5%
0-7.8	1.149	0.567	2.326	0.870	0.430	1.762
7.8+	2.220	1.366	3.606	0.450	0.277	0.732
ratio	1.932	0.850	4.393	0.518	0.228	1.177

So we see that if anything, the gap between intensive and conventional becomes wider after the end of intervention, although they are not formally statistically significantly different — the confidence interval for the ratio of HRs before and after is (0.85;4.39).

However, the basic assumption in this type of model is that there is a constant mortality ratio between the two groups, and that this ratio suddenly changes to a different value exactly the time when the two groups are offered the same treatment. This basically amounts to an averaging of the HR in the two intervals defined as during (0–7.8 years) and after (7.8+ years) intervention, which is apparently what was done in the NEJM paper from 2008 [1]. However the estimated mortality curves and the HR curve as a function of time since baseline as shown in figure 3.3 does not indicate any dramatic change in mortality post intervention, although it seems that the mortality rates do converge after the intervention period.

3.2.7 CVD events and all-cause mortality

It is of course also of interest to see whether CVD events post baseline have any influence on mortality rates. However, by subdividing follow-up by CVD-event status we are conditioning on intermediate events that are also influenced by the intervention, and by that token we are likely to underestimate the effect of allocation.

So we try to add indicators of intermediate CVD to the PH model for all-cause mortality:

- m1 effect any CVD after baseline
- m1 additional effect of subsequent CVD events, the same effect of each extra CVD event (up to 3)
- m3 separate effects of extra CVDs — that is no assumption of the same effect of each new CVD

```
> ( anyCVD <- levels(S1$lex.Cst)[2:4] )
[1] "1st CVD" "2nd CVD" "3rd CVD"
> m1 <- update( m0, . ~ . + I(lex.Cst %in% anyCVD) )
> m1 <- update( m1, . ~ . + pmax(as.integer(lex.Cst)-2,0) )
> m3 <- update( m1, . ~ . + lex.Cst )
> round( ci.exp( m1, subset="Cst" ), 4 )
```

```

                                exp(Est.)  2.5% 97.5%
I(lex.Cst %in% anyCVD)TRUE      4.9295 3.155 7.702
> round( ci.exp( m1, subset="Cst" ), 4 )

                                exp(Est.)  2.5% 97.5%
I(lex.Cst %in% anyCVD)TRUE      3.8140 2.3035 6.3150
pmax(as.integer(lex.Cst) - 2, 0)  1.5218 1.1190 2.0694
> round( ci.exp( m3, subset="Cst" ), 4 )

                                exp(Est.)  2.5% 97.5%
lex.Cst1st CVD                   3.6949 2.1884 6.2384
lex.Cst2nd CVD                    6.3278 3.3742 11.8666
lex.Cst3rd CVD                     7.9542 4.1698 15.1730

```

We can summarize the HR of death versus the group with no post-baseline CVD from the three models:

```

> cmpeff <- cbind( ci.exp( m3, subset="Cst" ),
+                 ci.exp( m1, subset="Cst",
+                         ctr.mat=rbind(c(1,0),
+                                       c(1,1),
+                                       c(1,2)) ),
+                 ci.exp( m1, subset="Cst" )[c(1,1,1),] )
> colnames( cmpeff )[c(1,4,7)] <- c("Grouped", "Linear", "Uniform")
> round( cmpeff, 3 )

      Grouped  2.5% 97.5% Linear  2.5% 97.5% Uniform  2.5% 97.5%
lex.Cst1st CVD  3.695 2.188 6.238  3.814 2.303 6.315  4.929 3.155 7.702
lex.Cst2nd CVD  6.328 3.374 11.867  5.804 3.663 9.196  4.929 3.155 7.702
lex.Cst3rd CVD  7.954 4.170 15.173  8.832 4.854 16.070  4.929 3.155 7.702

```

We can also compare the models, first by a Wald test for linearity of effects and also by comparing different models using a likelihood ratio test:

```

> Wald( m3, subset="Cst", ctr.mat=rbind(c(1,-2,1)) )
      Chisq      d.f.      P
0.2536462 1.0000000 0.6145193
> anova( m0, m1, m1, m3, test="Chisq" )

Analysis of Deviance Table

Model 1: (lex.Xst %in% dst) ~ allocation + Ns(tsb, kn = t.kn)
Model 2: (lex.Xst %in% dst) ~ allocation + Ns(tsb, kn = t.kn) + I(lex.Cst %in%
anyCVD)
Model 3: (lex.Xst %in% dst) ~ allocation + Ns(tsb, kn = t.kn) + I(lex.Cst %in%
anyCVD) + pmax(as.integer(lex.Cst) - 2, 0)
Model 4: (lex.Xst %in% dst) ~ allocation + lex.Cst + allocation:Ns(tsb,
kn = t.kn)

  Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1     29243     1234.3
2     29242     1182.2  1    52.038 5.445e-13
3     29241     1175.3  1     6.884 0.008698
4     29235     1172.3  6     2.982 0.811160

```

The three p-values are for the hypotheses of no effect of CVD, linear effect of no. CVD event, and additional non-linear effect of CVD.

Thus we see that the increase in mortality is the same from 0 to 1, from 1 to 2 and from 2 to 3 CVD events after baseline, namely a HR of 1.52 (1.12;2.07) (from model m1), and the likelihood ratio test for linearity, $p = 0.811$.

However, it might be of more interest to see if these effects are the same in the two intervention groups:

```

> mx1 <- update( m0, . ~ . + allocation:I(lex.Cst %in% levels(lex.Cst)[2:4]) )
> mx1 <- update( mx1, . ~ . + allocation:pmax(as.integer(lex.Cst)-2,0) )
> mx3 <- update( m0, . ~ . + allocation:lex.Cst )
> round( ci.exp( m1 ), 4 )

      exp(Est.)   2.5%   97.5%
(Intercept)      0.0026 0.0004  0.0192
allocationConventional  1.1395 0.7360  1.7640
Ns(tsb, kn = t.kn)1   10.3812 1.5536  69.3676
Ns(tsb, kn = t.kn)2    7.8597 0.7476  82.6293
Ns(tsb, kn = t.kn)3    6.5515 1.3786  31.1337
Ns(tsb, kn = t.kn)4   40.7542 0.5228 3176.6372
Ns(tsb, kn = t.kn)5    2.0692 0.9116  4.6968
I(lex.Cst %in% anyCVD)TRUE  3.8140 2.3035  6.3150
pmax(as.integer(lex.Cst) - 2, 0) 1.5218 1.1190  2.0694
> round( ci.exp( mx1, subset=c("base","Cst") ), 4 )

      exp(Est.)   2.5%   97.5%
allocationIntensive:I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE  5.8950 3.0955 11.2265
allocationConventional:I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE 4.2163 2.3348  7.6141
> round( ci.exp( mx1, subset=c("base","Cst") ), 4 )

      exp(Est.)   2.5%   97.5%
allocationIntensive:I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE  5.5162 2.7060 11.2451
allocationConventional:I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE 2.8012 1.4066  5.5786
allocationIntensive:pmax(as.integer(lex.Cst) - 2, 0)  1.1895 0.6485  2.1819
allocationConventional:pmax(as.integer(lex.Cst) - 2, 0)  1.7054 1.1842  2.4560
> round( ci.exp( mx3, subset=c("base","Cst") ), 4 )

      exp(Est.)   2.5%   97.5%
allocationIntensive:lex.Cst1st CVD  5.5537 2.6919 11.4578
allocationConventional:lex.Cst1st CVD  2.6120 1.2672  5.3841
allocationIntensive:lex.Cst2nd CVD  6.2803 2.2685 17.3868
allocationConventional:lex.Cst2nd CVD  5.8345 2.6752 12.7245
allocationIntensive:lex.Cst3rd CVD  7.8775 2.2323 27.7987
allocationConventional:lex.Cst3rd CVD  7.5113 3.5537 15.8763
> anova( m1, mx1, test="Chisq" )[2,-(1:2),drop=F]
  Df Deviance Pr(>Chi)
2  1  0.57058  0.45
> anova( m1, mx1, test="Chisq" )[2,-(1:2),drop=F]
  Df Deviance Pr(>Chi)
2  2  1.9986  0.3681
> anova( m3, mx3, test="Chisq" )[2,-(1:2),drop=F]
  Df Deviance Pr(>Chi)
2 -2 -0.43183  0.8058

...and indeed they are; so the relevant effect is still the overall hazard ratio from a
proportional hazards model that takes these different effects into account; here we show the
allocation effects both ways as well as the effects of post-baseline CVD:

> msb <- update( m0 , . ~ . + I(age-tsb) + sex )
> msl <- update( msb, . ~ . + I(lex.Cst %in% levels(lex.Cst)[2:4])
+
+ pmax(as.integer(lex.Cst)-2,0) )
> msf <- update( msb, . ~ . + lex.Cst )
> zz <- ci.exp( msb, subset=c("allo","Cst") )
> round( rbind( 1/zz[1,c(1,3,2)], zz ), 3 )

      exp(Est.) 97.5%  2.5%
      0.526 0.346 0.800
allocationConventional  1.899 1.249 2.888
> zz <- ci.exp( msl, subset=c("allo","Cst") )
> round( rbind( 1/zz[1,c(1,3,2)], zz ), 3 )

      exp(Est.) 97.5%  2.5%
      0.830 0.533 1.293
allocationConventional  1.205 0.774 1.877
I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE  3.003 1.799 5.012
pmax(as.integer(lex.Cst) - 2, 0)  1.577 1.151 2.160

```

```
> zz <- ci.exp( msf, subset=c("allo","Cst") )
> round( rbind( 1/zz[1,c(1,3,2)], zz ), 3 )
              exp(Est.) 97.5%  2.5%
allocationConventional  1.201 0.771  1.872
lex.Cst1st CVD          3.063 1.814  5.172
lex.Cst2nd CVD          4.422 2.360  8.286
lex.Cst3rd CVD          7.743 4.103 14.612
```

We see that as soon as post-baseline CVD is included in the model in one form or other, the effect of allocation group is changed from a HR of 1.9 to about 1.2, the latter non-significant. This is because the major part of the intervention effect is exercised on the transitions from DM to CVD and to subsequent CVD events, and these transitions are not modelled in the overall mortality analysis.

We shall return to a more detailed analysis of the CVD event rates later.

3.2.8 Summary of all-cause mortality

For all-cause mortality there is no indication of non-proportional hazards over the follow-up range ($P=0.447$), and the hazard ratio between the conventional and the intensive group is 1.94 (1.28;2.96).

Persons who see a post-baseline CVD have a higher mortality (HR 3.00 (1.80;5.58)) and an increase in HR of 1.54 (1.12;2.10) per extra CVD event up to 3; these effects are the same for the two intervention groups too.

The M/F HR was 1.49 (0.90;2.48) and effect of age was a HR of 1.11 (1.07;1.15) per year, corresponding to a doubling time of mortality of 6.8 (5.9;10.9) years. The latter shows that summary survival figures are heavily influenced by the particular age-distribution in the study population, *i.e.* they cannot easily be generalized to other settings, because their very strong dependency on the particular age and sex-distribution in the Steno 2 study. Hence the reporting of survival by age and sex.

The 20-year survival in the intensive group is 55%, and in the standard group 32%. These figures are however specific for the particular age-composition in the Steno 2 patient population. It is more relevant to state the 20-year survival of for example men entering at age 50 as 72% and 52% respectively; a more complete overview of survival probabilities is here:

```
> ftable( round(tabSurv[,,,c("5","10"),]*100,1),
+         col.vars=c(4,2,5),
+         row.vars=c(3,1) )
      tfr      5                                10
      sex      M      lo      up      F      lo      up      M      lo      up      F      lo      up
      what  est  lo  up  est  lo  up  est  lo  up  est  lo  up
age grp
45  Int      97.9 94.8 99.1 98.6 96.1 99.5 92.2 85.6 95.9 94.7 88.9 97.6
     Cnv      97.8 94.7 99.1 98.5 96.2 99.4 88.5 79.8 93.6 92.1 84.5 96.1
50  Int      96.4 91.9 98.4 97.6 94.0 99.0 87.3 79.1 92.4 91.3 83.4 95.5
     Cnv      96.3 91.9 98.3 97.5 94.1 99.0 81.4 71.3 88.2 87.1 77.4 92.8
55  Int      94.0 87.3 97.2 96.0 90.4 98.3 79.5 69.2 86.7 85.8 74.9 92.2
     Cnv      93.8 87.3 97.0 95.8 90.6 98.2 70.7 59.2 79.4 79.2 66.8 87.4
60  Int      90.2 79.8 95.3 93.3 84.6 97.2 67.9 54.3 78.3 77.2 61.9 86.9
     Cnv      89.8 79.8 95.0 93.0 85.0 96.9 55.7 42.2 67.2 67.6 51.5 79.4
65  Int      84.0 67.8 92.5 89.0 75.0 95.3 52.1 34.1 67.4 64.6 43.5 79.5
     Cnv      83.4 67.8 91.9 88.6 75.6 94.9 37.3 21.9 52.7 51.7 31.5 68.5

> ftable( round(tabSurv[,,,c("15","20"),]*100,1),
+         col.vars=c(4,2,5),
+         row.vars=c(3,1) )
```

age	grp	tfr sex what	15				20						
			M	F		M	F						
		est	lo	up	est	lo	up	est	lo	up	est	lo	up
45	Int	87.5	78.7	92.9	91.5	83.1	95.8	80.6	69.0	88.2	86.6	75.1	93.0
	Cnv	74.5	60.2	84.3	82.1	68.3	90.3	64.9	48.3	77.3	74.9	57.8	85.8
50	Int	79.9	69.8	87.0	86.1	75.3	92.4	69.6	57.4	78.9	78.4	64.5	87.4
	Cnv	60.9	47.0	72.1	71.7	56.0	82.6	48.2	33.7	61.4	61.4	43.3	75.3
55	Int	68.5	57.1	77.5	77.7	63.8	86.7	54.3	42.0	65.0	66.4	49.7	78.7
	Cnv	43.3	31.1	54.9	57.1	40.1	70.9	29.3	18.3	41.2	43.9	26.4	60.2
60	Int	52.9	39.4	64.7	65.3	47.4	78.4	35.7	23.3	48.3	50.2	31.0	66.6
	Cnv	24.4	14.2	36.0	38.9	21.8	55.7	12.6	5.6	22.6	25.0	10.6	42.4
65	Int	34.2	19.0	50.0	48.8	27.1	67.3	17.6	7.1	32.0	31.3	12.7	52.0
	Cnv	9.3	2.9	20.1	20.4	6.8	38.9	3.1	0.5	9.9	9.7	1.9	25.4

3.3 CVD and other cause mortality

Unlike the analysis of overall survival there is not a simple transformation of rates that yields the survival function when we consider multiple causes of death (CVD / non-CVD). Moreover, the survival function might not be the target of interest when analyzing CVD and other mortality separately — more likely the probabilities of interest would be the fraction of patients that die from each cause; or possibly the fraction of patients that see 0, 1, 2 or 3 CVD events before death. We shall return to these in the next chapter, as they depend on full modeling of all transitions shown in figure 1.3.

The analysis of the cause-specific mortalities is in principle exactly parallel to that of all-cause mortality, except that we use two different subsets of the death states as events:

```
> ( dst <- levels( S1 )[-(1:4)] )
[1] "Dead 0" "CV-D 0" "Dead 1" "CV-D 1" "Dead 2" "CV-D 2" "Dead 3" "CV-D 3"
```

3.3.1 CVD mortality

For the CVD mortality, the events are every other of the states:

```
> dst
[1] "Dead 0" "CV-D 0" "Dead 1" "CV-D 1" "Dead 2" "CV-D 2" "Dead 3" "CV-D 3"
> ( dcv <- dst[1:4*2] )
[1] "CV-D 0" "CV-D 1" "CV-D 2" "CV-D 3"
> sum( S1$lex.Xst %in% dcv )
[1] 38
```

and this is used in the modeling, but note that we only have 38 events, so the modeling possibilities are smaller, and we therefore only use 4 knots for the hazard rate function:

```
> ( t.kn <- with( subset(S1,lex.Xst %in% dcv),
+               c(0,quantile( tsb+lex.dur, probs=(1:3-0.5)/3) )) )
      16.66667%      50% 83.33333%
0.000000 4.935889 9.639973 16.059776
> m0 <- glm( (lex.Xst %in% dcv) ~ allocation + Ns(tsb, kn=t.kn),
+           family = poisson,
+           offset = log(lex.dur),
+           data = S1 )
> mi <- update( m0, . ~ . - Ns(tsb, kn=t.kn) +
+             allocation:Ns(tsb, kn=t.kn) )
> ma <- update( m0, . ~ . + I(age-tsb) + sex )
> mia <- update( mi, . ~ . + I(age-tsb) + sex )
> anova( ma, m0, mi, mia, ma, test="Chisq" )[-1,-(1:2)]
```

```

      Df Deviance Pr(>Chi)
2 -2 -23.7778 6.866e-06
3 3 8.2851 0.04047
4 2 24.3871 5.063e-06
5 -3 -8.8944 0.03073

> HR.IC <- rbind( ci.exp( m0, subset="allo", pval=TRUE ),
+               ci.exp( ma, subset="allo", pval=TRUE ) )
> round( cbind( 1/HR.IC[,c(1,3,2)], HR.IC ), 3 )

              exp(Est.) 97.5% 2.5% exp(Est.) 2.5% 97.5% P
allocationConventional 0.379 0.191 0.754 2.637 1.327 5.240 0.006
allocationConventional 0.353 0.177 0.705 2.832 1.419 5.652 0.003

> round( cbind( ci.exp( ma , subset=c("age","sex") ),
+               ci.exp( mia, subset=c("age","sex") ) ), 3 )

              exp(Est.) 2.5% 97.5% exp(Est.) 2.5% 97.5%
I(age - tsb) 1.123 1.064 1.186 1.126 1.066 1.189
sexM 2.256 0.940 5.415 2.266 0.944 5.442

```

We see that there is a borderline significant interaction, so formally we can not maintain the CVD mortality rates are proportional throughout the study period. Ignoring this, the estimated overall HR between intensive and conventional is 0.38 (0.19;0.75); and even with the rather small number of events this is significantly different from 1.

We then update the HR collector

```

> dimnames( mainCI )[[1]][2]
[1] "CVD mortality"

> zz <- HR.IC
> zz[,1:3] <- 1/HR.IC[,c(1,3,2)]
> mainCI[2,] <- zz
> save( mainCI, file="../data/mainCI.Rda" )

```

It is of course also of interest to see whether CVD event post baseline only influence CVD mortality rates; so we try to add these to the model:

```

> mx <- update( mia, . ~ . + I(lex.Cst %in% levels(lex.Cst)[2:4]) )
> round( ci.exp( mx ), 3 )

              exp(Est.) 2.5% 97.5%
(Intercept) 0.000 0.000 0.001
allocationConventional 0.402 0.010 16.487
I(age - tsb) 1.110 1.045 1.178
sexM 2.118 0.879 5.104
I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE 7.182 3.191 16.164
allocationIntensive:Ns(tsb, kn = t.kn)1 0.238 0.024 2.371
allocationConventional:Ns(tsb, kn = t.kn)1 8.159 1.121 59.412
allocationIntensive:Ns(tsb, kn = t.kn)2 1.242 0.008 193.489
allocationConventional:Ns(tsb, kn = t.kn)2 12.365 0.024 6368.653
allocationIntensive:Ns(tsb, kn = t.kn)3 0.438 0.102 1.875
allocationConventional:Ns(tsb, kn = t.kn)3 4.002 0.988 16.207

> anova( mia, mx, test="Chisq" )

Analysis of Deviance Table

Model 1: (lex.Xst %in% dcv) ~ allocation + I(age - tsb) + sex + allocation:Ns(tsb,
kn = t.kn)
Model 2: (lex.Xst %in% dcv) ~ allocation + I(age - tsb) + sex + I(lex.Cst %in%
levels(lex.Cst)[2:4]) + allocation:Ns(tsb, kn = t.kn)
Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1 29240 555.60
2 29239 527.83 1 27.771 1.365e-07

```

We see that there is a significant increased CVD mortality in persons with CVD events post baseline.

We now plot the two mortality curves and the RR between them from the models where follow-up CVD events are not taken into account; for the sex-age controlled model we use men aged 55 to show the mortality rates (median age at baseline is 56.6 years):

```
> CM <- Ns( t.pt, kn=t.kn )
> nd <- data.frame(tsb=t.pt,lex.dur=1000,age=55+t.pt,sex="M")
> rS.t <- ci.pred( mi, newdata=cbind(nd,allocation="Conventional") )
> rI.t <- ci.pred( mi, newdata=cbind(nd,allocation="Intensive") )
> rS.a <- ci.pred( mia, newdata=cbind(nd,allocation="Conventional") )
> rI.a <- ci.pred( mia, newdata=cbind(nd,allocation="Intensive") )
> pS.t <- ci.pred( m0, newdata=cbind(nd,allocation="Conventional") )
> pI.t <- ci.pred( m0, newdata=cbind(nd,allocation="Intensive") )
> pS.a <- ci.pred( ma, newdata=cbind(nd,allocation="Conventional") )
> pI.a <- ci.pred( ma, newdata=cbind(nd,allocation="Intensive") )
> # Check parameter sequencing using subset=
> round( ci.exp( mia, subset=c("Intensive","Conventional") ), 3 )

```

	exp(Est.)	2.5%	97.5%
allocationIntensive:Ns(tsb, kn = t.kn)1	0.388	0.039	3.865
allocationIntensive:Ns(tsb, kn = t.kn)2	5.676	0.043	757.559
allocationIntensive:Ns(tsb, kn = t.kn)3	0.661	0.158	2.764
allocationConventional	0.524	0.014	19.804
allocationConventional:Ns(tsb, kn = t.kn)1	20.276	2.918	140.910
allocationConventional:Ns(tsb, kn = t.kn)2	81.812	0.198	33882.038
allocationConventional:Ns(tsb, kn = t.kn)3	7.729	1.983	30.122

```
> RR.t <- ci.exp( mi, subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )
> RR.a <- ci.exp( mia, subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )
```

In order to plot the rightmost panel for the ESM together with the same for the all cause mortality we save the relevant structures:

```
> save( t.pt, HR.IC,
+       rI.t, rS.t,
+       rI.a, rS.a,
+       pI.t, pS.t, RR.t,
+       pI.a, pS.a, RR.a,
+       file = "../data/cvd-mort.Rda" )
```

We now have the predicted mortality rates and the estimated HRs from the two models, so we can plot them side by side and see to what extent adjustment for sex and age at baseline influences the estimates:

```
> par( mfrow=c(1,2), mar=c(0,0,0,0), oma=c(3,4,1,2), mgp=c(3,1,0)/1.6 )
> yl <- c(0.1,250)
> matplot( t.pt, cbind( rI.t, rS.t, RR.t),
+          log="y", ylim=yl, xlab="",
+          type="l", lty=1, lwd=c(3,1,1), col=rep(cclr,each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[1,-4], col="gray" )
> abline( h=1/HR.IC[1,1], col="gray", lwd=3 )
> matplot( t.pt, cbind( rI.a, rS.a, RR.a),
+          log="y", ylim=yl, yaxt="n", ylab="",
+          type="l", lty=1, lwd=c(3,1,1), col=rep(cclr,each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2,-4], col="gray" )
> abline( h=1/HR.IC[2,1], col="gray", lwd=3 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( rep(9,3), 20*0.75^c(2,1,3), c("Intensive","Conventional","Rate ratio"),
+       col=cclr, font=2, cex=1.2, adj=0 )
> mtext( "CVD mortality rate per 1000 PY / Hazard ratio",
+       side=2, line=3, las=0, outer=TRUE )
```

There is a decrease in the intensive vs. conventional HR of CVD mortality from around 5 years since randomization, consonant with the significant non-proportionality ($p=0.040$).

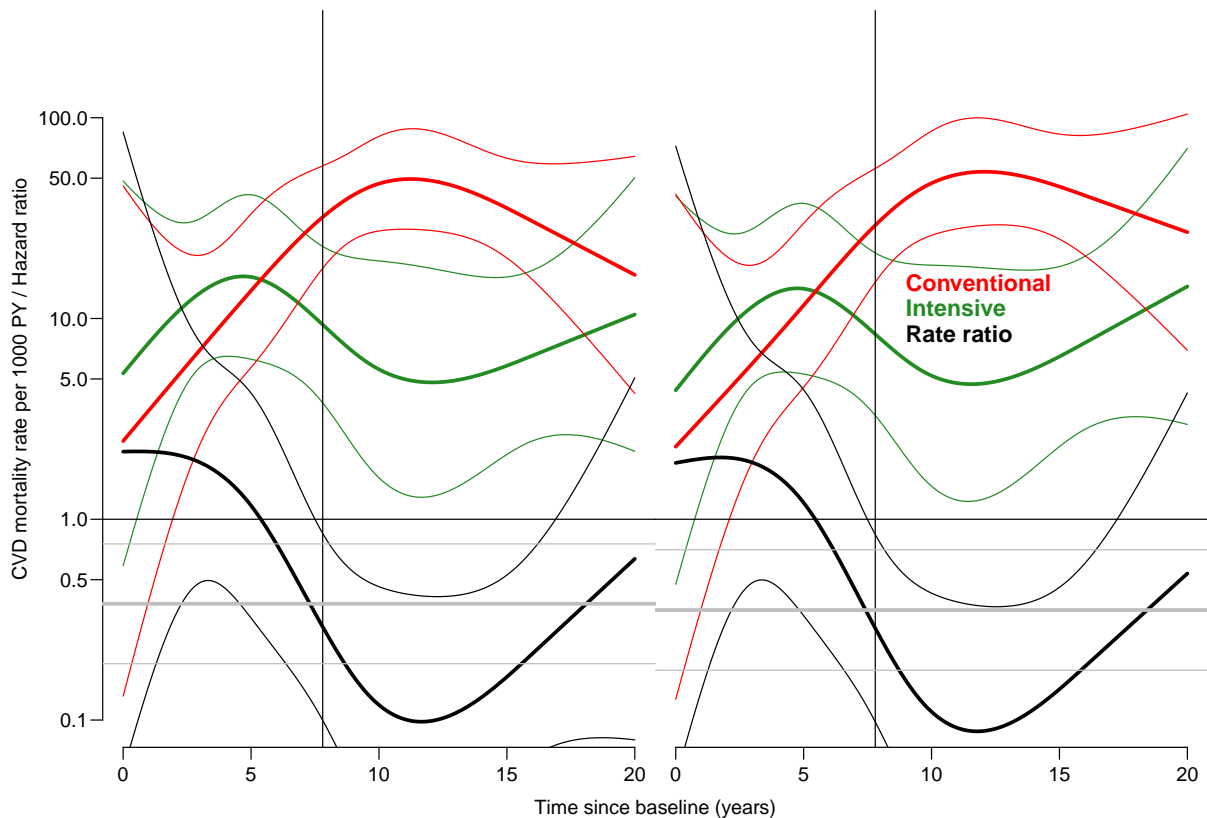


Figure 3.8: CVD mortality rates and hazard ratio between the two groups. The gray lines indicate the overall HR with 95% c.i., the vertical line indicates the intervention end. Left panel is a model with only group and time since baseline, the model in the right panel also includes sex and age at baseline as a linear term on the log-mortality scale, and mortality rates are shown for a man aged 55 at baseline. Horizontal gray lines indicate the overall HR from the (non-tenable) proportional hazards models.

3.3.2 Non-CVD mortality

For the non-CVD mortality, the events are also every other of the states:

```
> ( doth <- dst[0:3*2+1] )
[1] "Dead 0" "Dead 1" "Dead 2" "Dead 3"
> sum( S1$lex.Xst %in% doth )
[1] 55
```

and this is used in the modeling, but note that we only have 55 events, so the modeling possibilities are smaller than for all-cause mortality:

```
> ( t.kn <- with( subset(S1,lex.Xst %in% doth),
+               c(0,quantile( tsb+lex.dur, probs=(1:4-0.5)/4) )) )
      12.5%   37.5%   62.5%   87.5%
0.000000  5.535250  9.140999 12.841889 17.458590
> m0 <- glm( (lex.Xst %in% doth) ~ allocation + Ns(tsb, kn=t.kn),
+           family = poisson,
+           offset = log(lex.dur),
+           data = S1 )
> mi <- update( m0, . ~ . - Ns(tsb, kn=t.kn) +
+             allocation:Ns(tsb, kn=t.kn) )
> ma <- update( m0, . ~ . + I(age-tsb) + sex )
> mia <- update( mi, . ~ . + I(age-tsb) + sex )
> anova( ma, m0, mi, mia, ma, test="Chisq" )[-1,-(1:2)]
```

```

  Df Deviance Pr(>Chi)
2 -2 -21.0909 2.631e-05
3  4  1.7994  0.7726
4  2  20.9958 2.759e-05
5 -4 -1.7043  0.7899

> HR.IC <- rbind( ci.exp( m0, subset="allo", pval=TRUE ),
+               ci.exp( ma, subset="allo", pval=TRUE ) )
> rownames(HR.IC) <- c("Allo|ph", "Allo|ph+age+sex")
> round( cbind( HR.IC, 1/HR.IC[,c(1,3,2)] ), 3 )

      exp(Est.)  2.5% 97.5%      P exp(Est.) 97.5%  2.5%
Allo|ph      1.422 0.835 2.421 0.195      0.703 0.413 1.197
Allo|ph+age+sex 1.459 0.853 2.494 0.168      0.686 0.401 1.172

> round( cbind( ci.exp( ma , subset=c("age","sex") ),
+               ci.exp( mia, subset=c("age","sex") ) ), 3 )

      exp(Est.)  2.5% 97.5% exp(Est.)  2.5% 97.5%
I(age - tsb)      1.100 1.054 1.149      1.100 1.054 1.149
sexM              1.149 0.624 2.114      1.164 0.632 2.143

```

We see that there is no significant interaction, so formally we can maintain the the mortality rates are proportional throughout the study period, and the estimated HR between intensive and conventional is 0.70 (0.41;1.19) — not significantly different from 1.

We then update the HR collector

```

> dimnames( mainCI )[[1]][3]
[1] "non-CVD mortality"

> zz <- HR.IC
> zz[,1:3] <- 1/HR.IC[,c(1,3,2)]
> mainCI[3,,] <- zz
> save( mainCI, file="../data/mainCI.Rda" )

```

It is also of some interest to see whether CVD events post baseline also influences non-CVD mortality rates; so we try to add these to the model:

```

> mx <- update( mia, . ~ . + I(lex.Cst %in% levels(lex.Cst)[2:4]) )
> anova( ma, mx, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  5  15.288  0.0092

> round( ci.exp( mx, subset="Cst" ), 3 )

I(lex.Cst %in% levels(lex.Cst)[2:4])TRUE      exp(Est.)  2.5% 97.5%
      2.878 1.638 5.055

```

We see that there is a significant effect of CVD events post baseline; any CVD event holds a HR of 2.9 (1.6;5.1).

We now plot the two mortality curves and the HR between them from the models where post follow-up CVD events are not taken into account; for the sex-age-model we use men aged 55 to show the mortality rates (median age at baseline is 56.6 years):

```

> CM <- Ns( t.pt, kn=t.kn )
> nd <- data.frame( tsb=t.pt, lex.dur=1000, age=55+t.pt, sex="M" )
> rS.t <- ci.pred( mi , newdata=cbind(nd,allocation="Conventional") )
> rI.t <- ci.pred( mi , newdata=cbind(nd,allocation="Intensive") )
> rS.a <- ci.pred( mia, newdata=cbind(nd,allocation="Conventional") )
> rI.a <- ci.pred( mia, newdata=cbind(nd,allocation="Intensive") )
> # Check parameter sequencing using subset=
> round( ci.exp( mi , subset=c("Intensive","Conventional") ), 3 )

```

```

                                exp(Est.)  2.5%          97.5%
allocationIntensive:Ns(tsb, kn = t.kn)1    254.255 0.544 1.188090e+05
allocationIntensive:Ns(tsb, kn = t.kn)2     23.663 0.290 1.930249e+03
allocationIntensive:Ns(tsb, kn = t.kn)3    6575.244 0.007 6.102874e+09
allocationIntensive:Ns(tsb, kn = t.kn)4     12.715 1.173 1.378550e+02
allocationConventional                       0.922 0.000 3.639632e+03
allocationConventional:Ns(tsb, kn = t.kn)1   153.879 1.403 1.687202e+04
allocationConventional:Ns(tsb, kn = t.kn)2    50.276 1.523 1.660002e+03
allocationConventional:Ns(tsb, kn = t.kn)3  50603.940 0.964 2.655259e+09
allocationConventional:Ns(tsb, kn = t.kn)4    6.731 1.058 4.281300e+01

> RR.t <- ci.exp( mi , subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )
> RR.a <- ci.exp( mia, subset=c("Intensive","Conventional"), ctr.mat=cbind(CM,-1,-CM) )

```

We now have the predicted mortality rates and the estimated HRs from the two models, so we can plot them side by side and see to what extent adjustment for sex and age at baseline influences the estimates:

```

> par( mfrow=c(1,2), mar=c(0,0,0,0), oma=c(3,4,1,2), mgp=c(3,1,0)/1.6 )
> yl <- c(0.1,250)
> matplot( t.pt, cbind(rI.t,rS.t,RR.t),
+          log="y", ylim=yl, xlab="",
+          type="l", lty=1, lwd=c(3,1,1), col=rep(cclr,each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[1,-4], col="gray" )
> abline( h=1/HR.IC[1,1], col="gray", lwd=3 )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),
+          log="y", ylim=yl, yaxt="n", ylab="",
+          type="l", lty=1, lwd=c(3,1,1), col=rep(cclr,each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2,-4], col="gray" )
> abline( h=1/HR.IC[2,1], col="gray", lwd=3 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( rep(11,3), 10*0.75^c(2,1,3), c("Intensive","Conventional","Rate ratio"),
+       col=cclr, font=2, cex=1.2, adj=0 )
> mtext( "Non-CVD mortality rate per 1000 PY / Hazard ratio",
+       side=2, line=3, las=0, outer=TRUE )

```

We see from figure 3.9 that mortality increases substantially the first 5 years, and thereafter levels off; also it is pretty obvious that there is not much evidence of non-proportionality, the p-value for the null of proportional hazards along the time scale is 0.773.

3.4 Mortality summary

In terms of mortality the majority of the intervention effect is on CVD mortality; there is no statistically or clinically (figure ??) significant difference between the non-CVD mortality between intensive and conventional group.

The intensive effects is manifest as a crude HR of 2.6 (1.3;5.2) in the conventional group relative to the intensive group; if controlled for age and sex it is 2.8 (1.4;5.7). The HR in the intensive group relative to the conventional group is 0.37 (0.19,0.85) and 0.35 (0.18;0.70) respectively; significantly different from 1 but not very well determined.

3.4.1 Plot for the ESM

Here we just take the two summary plots for the ESM:

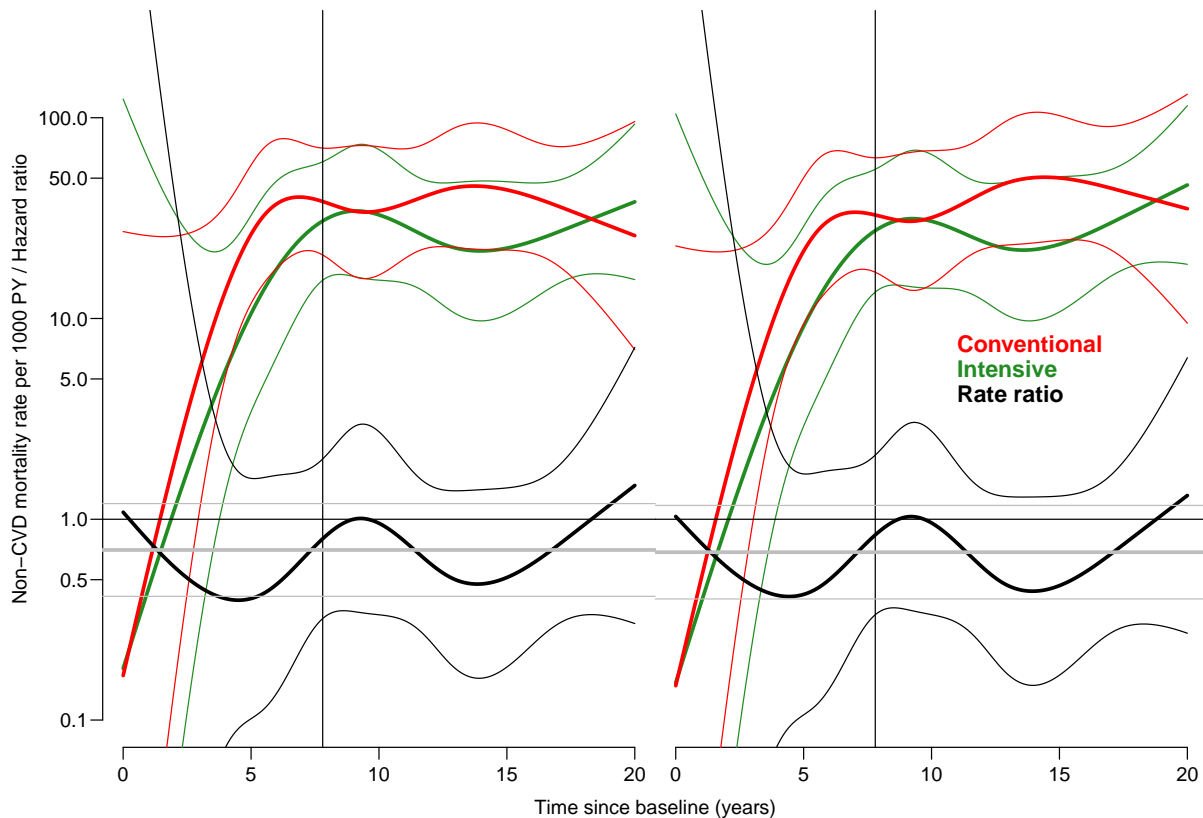


Figure 3.9: Non-CVD mortality rates and hazard ratio between the two groups. The gray lines indicate the overall HR with 95% c.i., the vertical line indicates the intervention end. Left panel is a model with only group and time since baseline, the model in the right panel also includes sex and age at baseline as a linear term on the log-mortality scale, and mortality rates are shown for a man aged 55 at baseline. The rate-scale is the same as that for CVD mortality, but not for all-cause mortality.

```

> clr <- c("forestgreen","red","black",gray(0.6))
> slr <- rgb( t( col2rgb(clr[1:2]) ), alpha=40, max=255 )
> par( mfrow=c(1,2), mar=c(0,2,0,0), oma=c(3,2,1,2), mgp=c(3,1,0)/1.6 )
> yl <- c(0.1,250)
> load( file = "../data/all-mort.Rda" )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),#pI.a,pS.a),
+         log="y", ylim=yl, ylab="", # yaxt="n",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,
+         lwd=c(3,1,1), col=rep(clr[1:3],each=3) )
> matlines( t.pt, cbind(pI.a,pS.a),
+          type="l", lty=1, lwd=c(3,1,1), col=rep(clr[1:2],each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2,2:3], col=clr[3], lty=1 )
> abline( h=1/HR.IC[2, 1], col=clr[3], lwd=3, lty=1 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( rep(11,3), 10*0.70~c(2,1,3), c("Intensive","Conventional","Rate ratio"),
+      col=clr[-4], font=2, cex=1.2, adj=0 )
> text( 0, 200, "All cause\nmortality", adj=0, font=2, cex=1.2 )
> mtext( "Mortality rate per 1000 PY / Hazard ratio Intensive vs Conventional",
+      side=2, line=2.5, las=0, outer=FALSE )
> load( file = "../data/cvd-mort.Rda" )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),
+         log="y", ylim=yl, ylab="", yaxt="n",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,

```

```

+           lwd=c(3,1,1), col=rep(clr[1:3],each=3) )
> matlines( t.pt, cbind(pI.a,pS.a),
+           type="l", lty=1, lwd=c(3,1,1), col=rep(clr[1:2],each=3) )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2,2:3], col=clr[3], lty=1 )
> abline( h=1/HR.IC[2, 1], col=clr[3], lwd=3, lty=1 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( 0, 200, "CVD\nmortality", adj=0, font=2, cex=1.2 )

> clr <- c("forestgreen","red","black",gray(0.6))
> slr <- rgb( t( col2rgb(clr[1:2]) ), alpha=40, max=255 )
> par( mfrow=c(1,2), mar=c(0,2,0,0), oma=c(3,2,1,2), mgp=c(3,1,0)/1.6 )
> yl <- c(0.1,250)
> rmcol <- c(clr[1:3],"transparent")[c(1,4,4,2,4,4,3,4,4)]
> load( file = "../data/all-mort.Rda" )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),#pI.a,pS.a),
+         log="y", ylim=yl, ylab="", # yaxt="n",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,
+         lwd=c(3,1,1), col=rmcol )
> matlines( t.pt, cbind(pI.a,pS.a),
+           type="l", lty=1, lwd=c(3,1,1), col=rmcol[1:6] )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2, 1], col=clr[3], lwd=3, lty=1 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
> text( rep(11,3), 10*0.70^c(2,1,3), c("Intensive","Conventional","Rate ratio"),
+       col=clr[-4], font=2, cex=1.2, adj=0 )
> text( 0, 200, "All cause\nmortality", adj=0, font=2, cex=1.2 )
> mtext( "Mortality rate per 1000 PY / Hazard ratio Intensive vs Conventional",
+       side=2, line=2.5, las=0, outer=FALSE )
> load( file = "../data/cvd-mort.Rda" )
> matplot( t.pt, cbind(rI.a,rS.a,RR.a),#pI.a,pS.a),
+         log="y", ylim=yl, ylab="", yaxt="n",
+         type="l", lty=rep(c("21","63","63"),3), lend=1,
+         lwd=c(3,1,1), col=rmcol )
> matlines( t.pt, cbind(pI.a,pS.a),
+           type="l", lty=1, lwd=c(3,1,1), col=rmcol[1:6] )
> abline( h=1, v=7.8 )
> abline( h=1/HR.IC[2, 1], col=clr[3], lwd=3, lty=1 )
> text( 0, 200, "CVD\nmortality", adj=0, font=2, cex=1.2 )
> mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )

```

Chapter 4

CVD status during follow-up

Since a large part of the intervention effect is mediated through the occurrence of CVD events, a natural way of showing the intervention effect is through the burden of CVD events in the two groups of patients. That is, using the status of the patients during follow-up as having 0, 1, 2 or 3 CVD events, and enumerating the time spent in each state and the fraction of patients dying with 0, 1, 2 and 3 CVD events.

4.1 Multistate set-up

To this end we take the previous analysis dataset but merge the two types of death:

```
> library( survival )
> library( Epi )
> clear()
> sessionInfo()
R version 3.3.0 (2016-05-03)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Ubuntu 14.04.4 LTS

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C              LC_TIME=en_US.UTF-8
 [4] LC_COLLATE=en_US.UTF-8   LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8     LC_NAME=C                 LC_ADDRESS=C
[10] LC_TELEPHONE=C          LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] utils      datasets  graphics  grDevices  stats      methods   base

other attached packages:
[1] Epi_2.5      survival_2.39-4

loaded via a namespace (and not attached):
 [1] cmprsk_2.2-7      MASS_7.3-44      plyr_1.8.3       Matrix_1.2-1
 [5] parallel_3.3.0   etm_0.6-2        Rcpp_0.11.6      splines_3.3.0
 [9] grid_3.3.0       numDeriv_2014.2-1 lattice_0.20-31
> load( file="../data/Lx.Rda" )
> lls()

  name mode class      size
1 L0  list Lexis data.frame 160 22
2 L1  list Lexis data.frame 318 22
> levels( L0 )
[1] "DM" "Dead" "CV-D"
> levels( L1 )
```

```

[1] "DM"      "1st CVD" "2nd CVD" "3rd CVD" "Dead 0"  "CV-D 0"  "Dead 1"  "CV-D 1"
[9] "Dead 2"  "CV-D 2"  "Dead 3"  "CV-D 3"
> L1 <- Relevel( L1, list("3+ CVD" = 4,
+                          "D(no CVD)" = 5:6,
+                          "D(1 CVD)" = 7:8,
+                          "D(2 CVD)" = 9:10,
+                          "D(3+ CVD)" = 11:12), first=FALSE )
  type      old      new
1 lex.Cst   DM      DM
2 lex.Cst  1st CVD  1st CVD
3 lex.Cst  2nd CVD  2nd CVD
4 lex.Cst  3rd CVD  3+ CVD
5 lex.Cst  Dead 0
6 lex.Cst  CV-D 0
7 lex.Cst  Dead 1
8 lex.Cst  CV-D 1
9 lex.Cst  Dead 2
10 lex.Cst CV-D 2
11 lex.Cst Dead 3
12 lex.Cst CV-D 3
13 lex.Xst   DM      DM
14 lex.Xst  1st CVD  1st CVD
15 lex.Xst  2nd CVD  2nd CVD
16 lex.Xst  3rd CVD  3+ CVD
17 lex.Xst  Dead 0  D(no CVD)
18 lex.Xst  CV-D 0  D(no CVD)
19 lex.Xst  Dead 1  D(1 CVD)
20 lex.Xst  CV-D 1  D(1 CVD)
21 lex.Xst  Dead 2  D(2 CVD)
22 lex.Xst  CV-D 2  D(2 CVD)
23 lex.Xst  Dead 3  D(3+ CVD)
24 lex.Xst  CV-D 3  D(3+ CVD)
> summary( L1 )
Transitions:
  To
From      DM 1st CVD 2nd CVD 3+ CVD D(no CVD) D(1 CVD) D(2 CVD) D(3+ CVD) Records:
DM         41    86     0      0      33      0      0      0      160
1st CVD   0     11    48     0      0      27     0      0      86
2nd CVD   0      0     7    24     0      0     16     0      47
3+ CVD    0      0     0     8      0      0      0     17     25
Sum       41    97    55    32    33     27    16    17    318

Transitions:
  To
From      Events: Risk time: Persons:
DM         119    1870.64    160
1st CVD    75     342.61     86
2nd CVD    40     112.27     47
3+ CVD     17     92.08      25
Sum        251    2417.60    160

```

We show the transitions separately for the two groups, using the same colouring of the states (DM, CVD, and death from each of state)

```

> clr <- c("forestgreen",heat.colors(5)[3:1])
> clr[3] <- rgb(t(col2rgb(clr[2])+col2rgb(clr[4]))/2,max=255)
> clr <- c(clr,rgb(t(col2rgb(clr)*0.6+255*0.4),max=255))
> names( clr ) <- levels(L1)
> clr
  DM      1st CVD      2nd CVD      3+ CVD      D(no CVD)      D(1 CVD)
"forestgreen" "#FFAA00FF" "#FF5500"  "#FF0000FF" "#7AB97A"    "#FFCC66"
  D(2 CVD)    D(3+ CVD)
"#FF9966"    "#FF6666"

```

```

> par( mfrow=c(1,2), mar=c(0,0,0,0) )
> for( lv in levels(L1$allocation) )
+ {
+ boxes( subset(L1,allocation==lv),
+        boxpos=list(x=c(rep(20,4),rep(80,4))+
+                        7-14*(lv=="Intensive"),
+                        y=rep(seq(90,10,,4),2) ),
+        pos.arr=c(rep(c(5,2),3),2)/10,
+        show.BE="nz", DR.sep=c(" ", " ")), hmult=1.3,
+        scale.R=100, lwd=3,
+        col.bg=clr,
+        col.border=c(clr[1:4],rep("black",4)),
+        col.txt=rep(c("white","black"),each=4) )
+ text( 10+5-10*(lv=="Intensive"), 98, lv, cex=1.5, font=2, adj=0 )
+ }

```

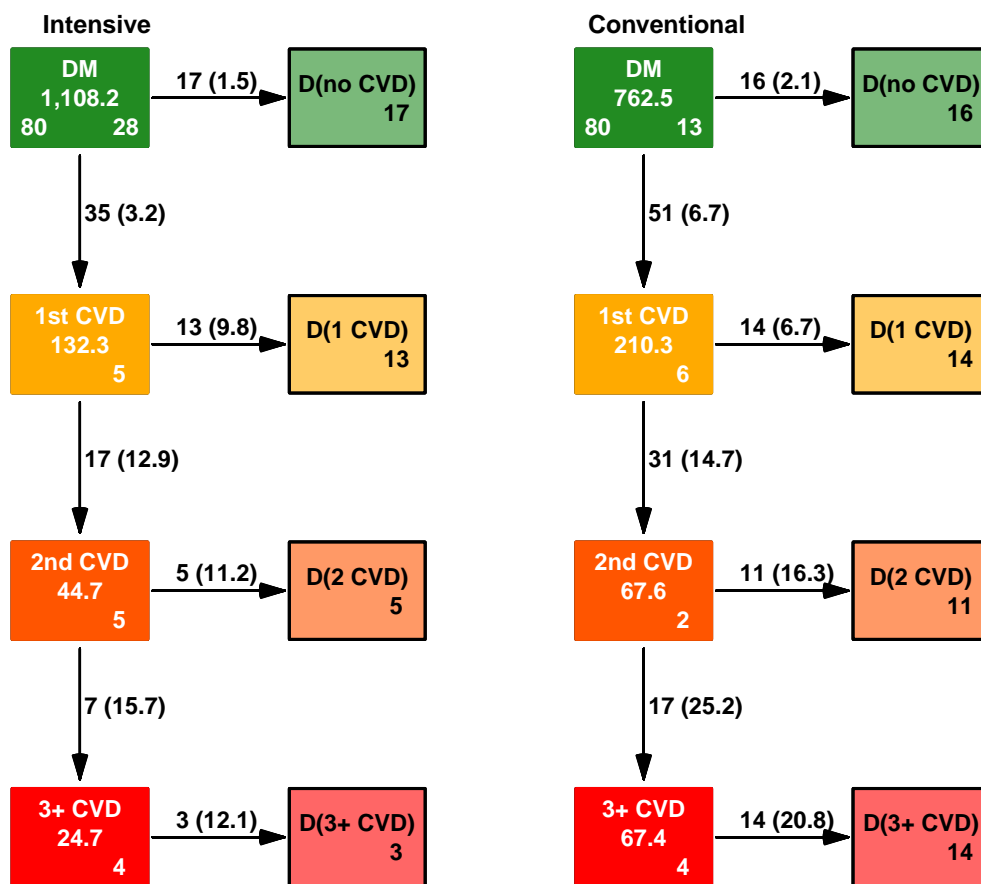


Figure 4.1: Transitions between states of CVD and death for the two randomization groups. The number in the center of the boxes are the person-years in each state, the number at the bottom left the number of persons starting follow up in the state and the bottom right is the number of persons ending follow-up in the state. The numbers on the arrows is the number of transitions and in parenthesis the event rates per 100 PY.

In order to answer the questions above we need a statistical models for all 7 transitions in figure 4.1.

4.2 Models for transitions

We shall model all transitions between the defined states by a function of time since baseline (baseline hazard), however only with separate effects for CVD transitions and for death. We shall moreover assume proportionality of hazards between the intervention and control groups.

The model we use in the first place has the following two separate baseline hazards:

- CVD events — this is 3 transitions which we shall assume proportional between different states of CVD progression, thus 3 proportionality parameters in addition to the baseline hazard.
- Death — this is 4 different transitions, assumed proportional between patients in different states of CVD; so three proportionality parameters on top of the baseline hazard.

Further to this model we will add a separate intervention hazard ratio for each of the 7 transitions; these will be one of the main results of the analysis.

This model will be expanded by separate sex- and linear age terms for CVD rates and mortality rates. As described the model is really two separate models; one for CVD events and one for death.

4.2.1 Modeling

We note that the transitions assumed proportional all originate in *different* states; and moreover that the mortality rates refer to *all* follow-up time.

Therefore we can do the modeling using a time-split dataset; because there are no two transitions *out* of the same box, there is no need to stack the data for analysis.

So initially we split the data in 1-month intervals of follow-up and show how the risk time and events are divided between the intervention and control groups:

```
> S1 <- splitLexis( L1, breaks=seq(0,25,1/12), time.scale="tsb" )
> summary( S1, by="allocation" )
```

```
$Intensive
```

```
Transitions:
```

From	To	DM	1st CVD	2nd CVD	3+ CVD	D(no CVD)	D(1 CVD)	D(2 CVD)	D(3+ CVD)	Records:
DM		13282	35	0	0	17	0	0	0	13334
1st CVD		0	1592	17	0	0	13	0	0	1622
2nd CVD		0	0	543	7	0	0	5	0	555
3+ CVD		0	0	0	299	0	0	0	3	302
Sum		13282	1627	560	306	17	13	5	3	15813

```
Transitions:
```

From	To	Events:	Risk time:	Persons:
DM		52	1108.17	80
1st CVD		30	132.29	35
2nd CVD		12	44.70	17
3+ CVD		3	24.70	7
Sum		97	1309.87	80

```
$Conventional
```

```
Transitions:
```

From	To									Records:
	DM	1st CVD	2nd CVD	3+ CVD	D(no CVD)	D(1 CVD)	D(2 CVD)	D(3+ CVD)		
DM	9125	51	0	0	16	0	0	0		9192
1st CVD	0	2528	31	0	0	14	0	0		2573
2nd CVD	0	0	815	17	0	0	11	0		843
3+ CVD	0	0	0	815	0	0	0	14		829
Sum	9125	2579	846	832	16	14	11	14		13437

Transitions:

From	To		
	Events:	Risk time:	Persons:
DM	67	762.47	80
1st CVD	45	210.31	51
2nd CVD	28	67.57	30
3+ CVD	14	67.38	18
Sum	154	1107.73	80

4.2.2 Mortality rates

We first devise a set of knots for the spline basis to use for the underlying mortality rates:

```
> levels(S1)
[1] "DM"          "1st CVD"    "2nd CVD"    "3+ CVD"    "D(no CVD)" "D(1 CVD)"  "D(2 CVD)"
[8] "D(3+ CVD)"
> ( d.kn <- c(0,with( subset(S1,lex.Xst %in% levels(lex.Xst)[5:8]),
+                   quantile(tsb+lex.dur,probs=1:4/5) ) ) )
                20%    40%    60%    80%
0.000000  6.254346  9.174538 11.664339 16.176318
```

With these we can now fit the relevant models for mortality, the base model (`m0`) for death being the model with proportional mortalities between states of CVD and a common intervention effect.

```
> ( dlev <- levels(S1$lex.Xst)[5:8] )
[1] "D(no CVD)" "D(1 CVD)"  "D(2 CVD)"  "D(3+ CVD)"
> m0 <- glm( (lex.Xst %in% dlev) ~
+           Ns( tsb, knots=d.kn ) + lex.Cst + allocation,
+           offset = log(lex.dur),
+           family = poisson,
+           data = S1 )
> m1 <- update( m0, . ~ . + sex + age )
> m1 <- update( m0, . ~ . - lex.Cst + I(as.integer(lex.Cst)) )
> m0i <- update( m0, . ~ . - allocation + allocation:lex.Cst )
> m1i <- update( m1, . ~ . - allocation + allocation:lex.Cst )
> mli <- update( m1, . ~ . + allocation:I(as.integer(lex.Cst)) )

> mcmp <- rbind(
+ anova( m1i, m1, m0, m0i, test="Chisq" )[-1,-(1:2)],
+ anova( mli, m1, m0, test="Chisq" )[-1,-(1:2)] )
> rownames( mcmp ) <- c("CVD*allo|s+a",
+                       "s+a|ph",
+                       "CVD*allo",
+                       "1CVD*allo",
+                       "1CVD|CVD")
> round( abs(mcmp), 3 )

                Df Deviance Pr(>Chi)
CVD*allo|s+a   3    2.711   0.438
s+a|ph         2   30.619  <2e-16
CVD*allo       3    2.392   0.495
1CVD*allo      1    0.427   0.513
1CVD|CVD       2    6.414   0.040
```

We see there is evidence that the CVD is non-linear in no of events, but also that there is no evidence that the effect of no. of CVD events is non-proportional between allocation groups.

```

> ests <- cbind( ci.exp(m1, pval=TRUE ),
+               rbind(ci.exp(m1, pval=TRUE ),NA)[c(1:5,7,8,8,6,8,8),],
+               rbind(ci.exp(m0, pval=TRUE ),NA)[c(1:9,10,10),] )
> colnames(ests)[c(1,5,9)] <- c(" m1"," m1"," m0")
> round( ests[-(1:5),], 3 )

```

	m1	2.5%	97.5%	P	m1	2.5%	97.5%	P	m0	2.5%
lex.Cst1st CVD	3.076	1.821	5.194	0.000	2.082	1.728	2.51	0.000	3.721	2.206
lex.Cst2nd CVD	4.419	2.356	8.288	0.000	NA	NA	NA	NA	6.319	3.415
lex.Cst3+ CVD	7.760	4.111	14.645	0.000	NA	NA	NA	NA	8.517	4.487
allocationConventional	1.198	0.769	1.867	0.425	1.178	0.758	1.83	0.466	1.144	0.738
sexM	1.380	0.828	2.300	0.216	NA	NA	NA	NA	NA	NA
age	1.101	1.062	1.142	0.000	NA	NA	NA	NA	NA	NA

```


```

	97.5%	P
lex.Cst1st CVD	6.274	0.000
lex.Cst2nd CVD	11.690	0.000
lex.Cst3+ CVD	16.168	0.000
allocationConventional	1.773	0.548
sexM	NA	NA
age	NA	NA

```

> round( cbind( ci.exp(m1 , subset=c("sex","age") ),
+               ci.exp(m1i, subset=c("sex","age") ) ), 3 )

```

	exp(Est.)	2.5%	97.5%	exp(Est.)	2.5%	97.5%
sexM	1.380	0.828	2.300	1.333	0.797	2.230
age	1.101	1.062	1.142	1.103	1.063	1.144

```

> HR <- cbind( ci.exp(m1i,subset="Cst"),
+               ci.exp(m0i,subset="Cst") )
> round( rbind(HR,1/HR[4:7,,drop=FALSE]), 3 )

```

	exp(Est.)	2.5%	97.5%	exp(Est.)	2.5%	97.5%
lex.Cst1st CVD	4.813	2.323	9.971	5.564	2.697	11.482
lex.Cst2nd CVD	4.678	1.693	12.924	6.334	2.288	17.536
lex.Cst3+ CVD	8.023	2.298	28.005	7.775	2.199	27.493
lex.CstDM:allocationConventional	1.664	0.835	3.315	1.487	0.749	2.951
lex.Cst1st CVD:allocationConventional	0.712	0.332	1.529	0.699	0.328	1.487
lex.Cst2nd CVD:allocationConventional	1.371	0.457	4.115	1.356	0.452	4.070
lex.Cst3+ CVD:allocationConventional	1.348	0.377	4.819	1.457	0.408	5.201
lex.CstDM:allocationConventional	0.601	1.197	0.302	0.673	1.335	0.339
lex.Cst1st CVD:allocationConventional	1.404	3.012	0.654	1.431	3.045	0.673
lex.Cst2nd CVD:allocationConventional	0.729	2.189	0.243	0.737	2.212	0.246
lex.Cst3+ CVD:allocationConventional	0.742	2.652	0.208	0.686	2.450	0.192

```

> HR <- cbind( ci.exp(m1 ,subset=c("Cst","allo"), pval=TRUE ),
+               ci.exp(m0 ,subset=c("Cst","allo"), pval=TRUE ) )
> IHR <- 1/HR[4,,drop=F]
> IHR[,c(4,8)] <- 1/IHR[,c(4,8)]
> round( rbind(HR,IHR), 3 )

```

	exp(Est.)	2.5%	97.5%	P	exp(Est.)	2.5%	97.5%	P
lex.Cst1st CVD	3.076	1.821	5.194	0.000	3.721	2.206	6.274	0.000
lex.Cst2nd CVD	4.419	2.356	8.288	0.000	6.319	3.415	11.690	0.000
lex.Cst3+ CVD	7.760	4.111	14.645	0.000	8.517	4.487	16.168	0.000
allocationConventional	1.198	0.769	1.867	0.425	1.144	0.738	1.773	0.548
allocationConventional	0.835	1.301	0.536	0.425	0.874	1.355	0.564	0.548

We then update the HR collector:

```

> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][5]
[1] "Death | CVD state"

```

```
> mainCI[5,] <- rbind(IHR[c(1,3,2,4)],IHR[c(1,3,2,4)+4])
> save( mainCI, file=" ../data/mainCI.Rda" )
```

It is seen that mortality increases by CVD status, as well as by age. Also it is seen that the CVD-effects are smaller in the model where age is accounted for; this is because the persons with more advanced CVD status are older. The HR between intervention and control is 0.83 (0.53;1.30) when controlled for sex and age.

A graphical overview of the estimates of the intervention effect:

```
> e0 <- ci.exp(m0 ,subset="allocation")
> e1 <- ci.exp(m1 ,subset="allocation")
> e0i <- ci.exp(m0i,subset="allocation")
> e1i <- ci.exp(m1i,subset="allocation")
> rownames( e0 ) <- "Overall"
> rownames( e0i ) <- gsub( "lex.Cst", "",
+   gsub( ":allocationConventional", "", rownames(e0i) ) )
> rownames( e1i ) <- gsub( "lex.Cst", "",
+   gsub( ":allocationConventional", "", rownames(e1i) ) )
> par( mar=c(3,3,1,1), mgp=c(3,1,0)/1.6 )
> yp <- c(5,3:0)
> plotEst( 1/rbind(e0,e0i), txtpos=yp, y=yp+0.1,
+   vref=1, xlog=TRUE, xlim=c(1/5,5), grid=c(2:10/10,1.5,2:5),
+   xlab="Mortality HR: Intensive vs. conventional", cex=2, lwd=5 )
> linesEst( 1/rbind(e1,e1i), y=yp-0.1, col="gray", cex=2, lwd=5 )
```

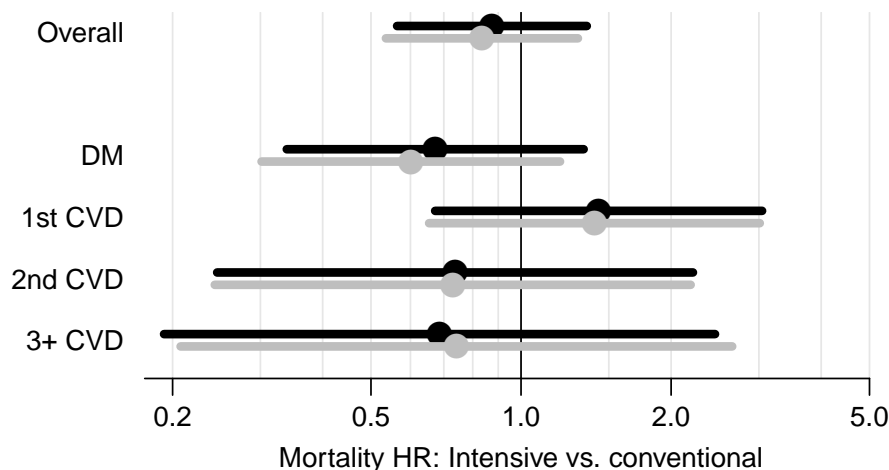


Figure 4.2: Ratio of mortality between intensive and conventional group, in different states of CVD progression. Gray estimates are from models with sex and age effect.

From figure 4.2 it is seen that there is no significant difference in mortality between the two groups when we condition on the CVD status. There seems to be a slightly **higher** mortality in the intervention group among persons with *one* CVD event than among persons with none or 2+ CVD events. As it is neither statistically significant nor biologically plausible, we shall use the proportional hazards approach; that is the one where we assume that intervention has the same effect (as measured by the HR) on all transition rates — the top estimate in the graph.

4.2.3 CVD rates

When we model the CVD occurrence rates (hazards) we must exclude the follow-up time in the state “3+ CVD” because these are not at risk of any further CVD event. Moreover the

specification of the events are a bit more tricky, because these are not just the exit status being equal to one of the CVD states, but *also* that `lex.Xst` is different from `lex.Cst`, because some of the split records will have both `lex.Cst` and `lex.Xst` equal to a given CVD state. Otherwise the analyses are along the same lines as the mortality analyses:

```
> ( clev <- levels(S1$lex.Xst)[2:4] )
[1] "1st CVD" "2nd CVD" "3+ CVD"

> c0 <- glm( ( lex.Xst %in% clev ) & ( lex.Cst!=lex.Xst ) ~
+           Ns( tsb, knots=d.kn ) + lex.Cst + allocation,
+           offset = log(lex.dur),
+           family = poisson,
+           data = subset( S1, lex.Cst!="3+ CVD" ) )
> c1 <- update( c0, . ~ . + sex + age )
> c1 <- update( c0, . ~ . - lex.Cst + I(as.integer(lex.Cst)) )
> c0i <- update( c0, . ~ . - allocation + allocation:lex.Cst )
> c1i <- update( c1, . ~ . - allocation + allocation:lex.Cst )
> c1i <- update( c1, . ~ . + allocation:I(as.integer(lex.Cst)) )
> ccmp <- rbind(
+ anova( c1i, c1, c0, c0i, test="Chisq" )[-1,-(1:2)],
+ anova( c1i, c1, c0, test="Chisq" )[-1,-(1:2)] )
> rownames( ccmp ) <- c("CVD*allo|s+a",
+                      "s+a|ph",
+                      "CVD*allo",
+                      "1CVD*allo",
+                      "1CVD|CVD")
> round( abs(ccmp), 3 )

              Df Deviance Pr(>Chi)
CVD*allo|s+a  2    2.685   0.261
s+a|ph        2   12.538   0.002
CVD*allo      2    2.712   0.258
1CVD*allo     1    0.988   0.320
1CVD|CVD      1    2.259   0.133

> cest <- cbind( ci.exp(c1, pval=TRUE ),
+               rbind( ci.exp(c1, pval=TRUE ), NA)[c(1:5,7,8,6,8,8),],
+               rbind( ci.exp(c0, pval=TRUE ), NA)[c(1:8,9,9),] )
> colnames(cest)[c(1,5,9)] <- c(" c1", " c1", " c0")
> round( cest[-(1:5),], 3 )

              c1  2.5% 97.5%    P    c1  2.5% 97.5%    P    c0  2.5% 97.5%
lex.Cst1st CVD    2.427 1.671 3.523 0.000 2.167 1.749 2.684 0.000 2.740 1.894 3.964
lex.Cst2nd CVD    3.481 2.148 5.643 0.000    NA    NA    NA    NA  4.260 2.661 6.818
allocationConventional 1.813 1.296 2.536 0.001 1.788 1.282 2.493 0.001 1.738 1.245 2.426
sexM              1.325 0.897 1.958 0.157    NA    NA    NA    NA    NA    NA    NA
age              1.042 1.016 1.068 0.001    NA    NA    NA    NA    NA    NA    NA
                P
lex.Cst1st CVD    0.000
lex.Cst2nd CVD    0.000
allocationConventional 0.001
sexM              NA
age              NA

> round( cbind( ci.exp(c1, subset=c("sex","age") ),
+             ci.exp(c1i, subset=c("sex","age") ) ), 3 )

              exp(Est.)  2.5% 97.5% exp(Est.)  2.5% 97.5%
sexM              1.325 0.897 1.958    1.296 0.876 1.919
age              1.042 1.016 1.068    1.043 1.017 1.069
```

From the models we can also extract both the HRs and the inverse HRs corresponding to intervention versus control — the latter are in the 3 last lines here:

```
> HR <- cbind( ci.exp(c1i,subset="Cst"),
+             ci.exp(c0i,subset="Cst") )
> round( rbind(HR,1/HR[3:5,,drop=FALSE]), 3 )
```

```

                                exp(Est.) 2.5% 97.5% exp(Est.) 2.5% 97.5%
lex.Cst1st CVD                   3.625 2.016 6.519    4.101 2.288 7.350
lex.Cst2nd CVD                   3.929 1.703 9.068    4.722 2.053 10.863
lex.CstDM:allocationConventional 2.229 1.443 3.444    2.134 1.383 3.291
lex.Cst1st CVD:allocationConventional 1.195 0.658 2.170    1.143 0.632 2.066
lex.Cst2nd CVD:allocationConventional 1.799 0.719 4.497    1.763 0.705 4.411
lex.CstDM:allocationConventional 0.449 0.693 0.290    0.469 0.723 0.304
lex.Cst1st CVD:allocationConventional 0.837 1.519 0.461    0.875 1.581 0.484
lex.Cst2nd CVD:allocationConventional 0.556 1.390 0.222    0.567 1.419 0.227

> HR <- cbind( ci.exp(c1, subset=c("Cst","allo"), pval=TRUE),
+             ci.exp(c0, subset=c("Cst","allo"), pval=TRUE) )
> round( rbind(HR,1/HR[,,drop=FALSE]), 3 )

                                exp(Est.) 2.5% 97.5%      P exp(Est.) 2.5% 97.5%      P
lex.Cst1st CVD                   2.427 1.671 3.523    0.000    2.740 1.894 3.964    0.000
lex.Cst2nd CVD                   3.481 2.148 5.643    0.000    4.260 2.661 6.818    0.000
allocationConventional            1.813 1.296 2.536    0.001    1.738 1.245 2.426    0.001
allocationConventional            0.552 0.772 0.394 1934.702    0.575 0.803 0.412 863.382

> IHR <- 1/HR[,,drop=F]
> IHR[,c(4,8)] <- 1/IHR[,c(4,8)]
> round( rbind(HR,IHR), 3 )

                                exp(Est.) 2.5% 97.5%      P exp(Est.) 2.5% 97.5%      P
lex.Cst1st CVD                   2.427 1.671 3.523    0.000    2.740 1.894 3.964    0.000
lex.Cst2nd CVD                   3.481 2.148 5.643    0.000    4.260 2.661 6.818    0.000
allocationConventional            1.813 1.296 2.536    0.001    1.738 1.245 2.426    0.001
allocationConventional            0.552 0.772 0.394    0.001    0.575 0.803 0.412    0.001

```

We then update the HR collector:

```

> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][6]
[1] "CVD event | CVD state"

> mainCI[6,] <- rbind(IHR[c(1,3,2,4)],IHR[c(1,3,2,4)+4])
> save( mainCI, file="../data/mainCI.Rda" )

```

As before we see smaller effect of the intervention after the first CVD; formally the intervention effect on CVD occurrence is non-significant among persons with one or two post-baseline CVD, but there is no consistently declining effect as shown in figure 4.3:

```

> e0 <- ci.exp(c0 ,subset="allocation")
> e1 <- ci.exp(c1 ,subset="allocation")
> e0i <- ci.exp(c0i,subset="allocation")
> e1i <- ci.exp(c1i,subset="allocation")
> rownames( e0 ) <- "Overall"
> rownames( e0i ) <- gsub( "lex.Cst", "",
+                         gsub( ":allocationConventional", "", rownames(e0i) ) )
> rownames( e1i ) <- gsub( "lex.Cst", "",
+                         gsub( ":allocationConventional", "", rownames(e1i) ) )
> par( mar=c(3,3,1,1), mgp=c(3,1,0)/1.6 )
> yp <- c(4,2:0)
> plotEst( 1/rbind(e0,e0i), txtpos=yp, y=yp+0.1,
+         vref=1, xlog=TRUE, xlim=c(1/5,5), grid=c(2:10/10,1.5,2:5),
+         xlab="Invervention vs. conventional CVD HR", cex=2, lwd=5 )
> linesEst( 1/rbind(e1,e1i), y=yp-0.1, col="gray", cex=2, lwd=5 )

```

As for mortality rates, for CVD rates there is no convincing evidence of interaction between allocation and state, so we use the model with common intervention HR for all CVD event rates.

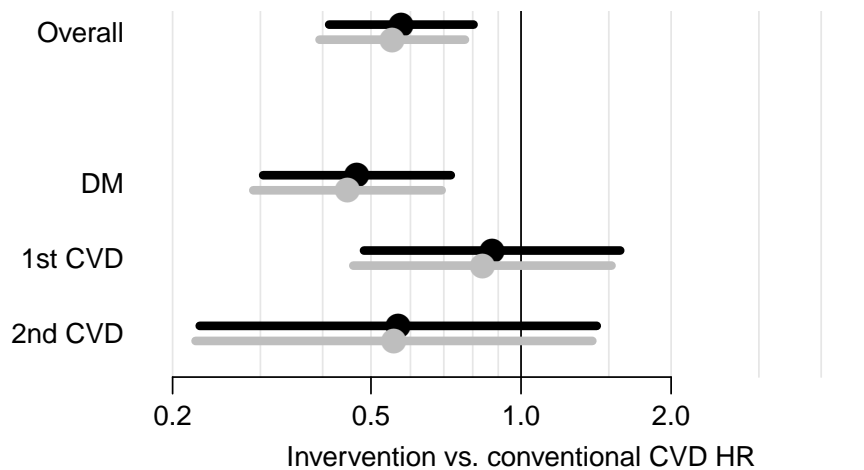


Figure 4.3: *Ratio of mortality between intervention and conventional group, in different states of CVD progression. The gray estimates are from a model with conventional for age and sex.*

4.2.4 Summary of intervention

In summary, we have thus two intervention effects; one for CVD-rates and the other for mortality rates — both of which are independent of the current CVD status of the patients (0, 1, 2 (or 3) CVD events).

For completeness we have also included HR from the analysis of overall mortality where CVD status was ignored:

Transition	Hazard ratio (95% c.i)				P
	Conv vs. Int		Int vs. Conv		
Crude					
Mortality	1.82	(1.20;2.75)	0.55	(0.36;0.83)	0.005
Mort CVD	1.14	(0.74;1.77)	0.87	(0.56;1.36)	0.548
CVD	1.74	(1.25;2.43)	0.58	(0.41;0.80)	0.001
Controlled for age and sex					
Mortality	1.90	(1.25;2.89)	0.53	(0.35;0.80)	0.003
Mort CVD	1.20	(0.77;1.87)	0.83	(0.54;1.30)	0.425
CVD	1.81	(1.30;2.54)	0.55	(0.39;0.77)	0.001

Note that the effect of intervention on mortality given CVD status (Mort|CVD) is formally not significant; this is because the effect shown is the effect on the *conditional* mortality *given* CVD state — the 4 horizontal transitions shown in figure 4.1. The intervention is only significant for the overall mortality *disregarding* CVD status.

Thus it seems that the major effect of the intervention on mortality is mediated by (prevention of) CVD occurrence.

4.3 Predictions

We now have (several) statistical models for all 7 transitions shown in figure 4.1 and this enables us to estimate the state occupancy probabilities for a given base population under these models. With state occupancy probabilities estimated as continuous functions of time we can compute the fraction of persons that see 0, 1, 2 or 3 CVDs during the first 20 years of follow-up; and we can compute the years spent alive with 0, 1, 2 and 3 CVD events. These quantities can then be compared between the persons in the two groups.

Both the mortality models and the CVD models we have fitted are cross-classified by whether they include a sex and an age term and whether they include an interaction between CVD state and intervention.

There are two substantially different scenarios we want to address:

- How does the follow-up look for the Steno2 population, and how is it influenced by the intervention?

This can be addressed using either the models with or without age and sex-effects, since the prediction anyway will be for a population with the same age- and sex-composition as the Steno2 population.

- How does the follow-up look for T2D patients of a given sex and age?

This can only be addressed using the models with age and sex-effects, since the prediction otherwise will be the same regardless of the population's age- and sex-composition.

In view of this it seems most sensible to use the models with age- and sex effects as basis for all predictions. Also in the light of the very significant effect of these two variables on the rates. Note that even if we have a randomized study, the fact that CVD rates are influenced by age and sex implies that the age- and sex-distribution in the CVD-states is not comparable between allocation groups.

There is no formal evidence in data that that neither the mortality nor the CVD HRs between intervention and conventional vary across CVD states:

```
> round( cbind( rbind( ci.exp(m1 , subset="allo", pval=T),
+                   ci.exp(m1i, subset="allo", pval=T) ),
+             rbind( ci.exp(c1 , subset="allo", pval=T),
+                   ci.exp(c1i, subset="allo", pval=T), NA ) ), 3 )
      exp(Est.)  2.5% 97.5%  P exp(Est.)  2.5% 97.5%
allocationConventional      1.198 0.769 1.867 0.425      1.813 1.296 2.536
lex.CstDM:allocationConventional      1.664 0.835 3.315 0.148      2.229 1.443 3.444
lex.Cst1st CVD:allocationConventional      0.712 0.332 1.529 0.384      1.195 0.658 2.170
lex.Cst2nd CVD:allocationConventional      1.371 0.457 4.115 0.574      1.799 0.719 4.497
lex.Cst3+ CVD:allocationConventional      1.348 0.377 4.819 0.646      NA      NA      NA
      P
allocationConventional      0.001
lex.CstDM:allocationConventional      0.000
lex.Cst1st CVD:allocationConventional      0.558
lex.Cst2nd CVD:allocationConventional      0.209
lex.Cst3+ CVD:allocationConventional      NA
> anova( m1, m1i, test="Chisq")
Analysis of Deviance Table

Model 1: (lex.Xst %in% dlev) ~ Ns(tsb, knots = d.kn) + lex.Cst + allocation +
sex + age
Model 2: (lex.Xst %in% dlev) ~ Ns(tsb, knots = d.kn) + lex.Cst + sex +
```



```

      age + lex.Cst:allocation
  Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1      29239      1145.1
2      29236      1142.4 3    2.7111  0.4383
> anova( c1, cli, test="Chisq")
Analysis of Deviance Table

Model 1: ((lex.Xst %in% clev) & (lex.Cst != lex.Xst)) ~ Ns(tsb, knots = d.kn) +
  lex.Cst + allocation + sex + age
Model 2: ((lex.Xst %in% clev) & (lex.Cst != lex.Xst)) ~ Ns(tsb, knots = d.kn) +
  lex.Cst + sex + age + lex.Cst:allocation
  Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1      28109      1833.3
2      28107      1830.6 2    2.6854  0.2611

```

Also, the deviance statistics are both smaller than 3.84, so it can be safely excluded that the non-significance is a mere product of inflation of degrees of freedom. Hence we shall make predictions based on the models with intervention effects constant across CVD strata.

4.3.1 PH model predictions

In order to set up a prediction scenario we need a transition object, that is a list of lists indicating what models to be used for each of the transitions. We set up two transition objects (models for transition intensities), one using the same proportionality between randomization groups for all transitions of each type (death / CVD) and the other allowing differential allocation effects on rates across CVD states. Both sets of models used are however proportional hazards models along the time since baseline scale.

```

> levels( S1 )
[1] "DM"      "1st CVD"  "2nd CVD"  "3+ CVD"   "D(no CVD)" "D(1 CVD)" "D(2 CVD)"
[8] "D(3+ CVD)"
> TM1 <- list( "DM" = list( "D(no CVD)" = m1,
+                          "1st CVD" = c1 ),
+             "1st CVD" = list( "D(1 CVD)" = m1,
+                               "2nd CVD" = c1 ),
+             "2nd CVD" = list( "D(2 CVD)" = m1,
+                               "3+ CVD" = c1 ),
+             "3+ CVD" = list( "D(3+ CVD)" = m1 ) )
> TM1i <- list( "DM" = list( "D(no CVD)" = m1i,
+                          "1st CVD" = c1i ),
+             "1st CVD" = list( "D(1 CVD)" = m1i,
+                               "2nd CVD" = c1i ),
+             "2nd CVD" = list( "D(2 CVD)" = m1i,
+                               "3+ CVD" = c1i ),
+             "3+ CVD" = list( "D(3+ CVD)" = m1i ) )
> lapply( TM1, names )
$DM
[1] "D(no CVD)" "1st CVD"

$`1st CVD`
[1] "D(1 CVD)" "2nd CVD"

$`2nd CVD`
[1] "D(2 CVD)" "3+ CVD"

$`3+ CVD`
[1] "D(3+ CVD)"

```

Note that there is only two models used in each of the transition objects; because we are using the proportional hazards assumption for the CVD rates and for the mortality rates separately.


```

user system elapsed
124.118 3.743 129.355
> save( sim2, sim2i, file="../data/sim2.Rda" )

```

The Lexis objects `sim2` and `sim2i` now contain a simulated follow up of a cohort of 32,000 persons; 100 copies of each of the original Steno2 participants (that is of the age and sex of the persons) assigned to conventional and 100 copies assigned to intervention. Thus the observed state distribution in each of the two subsets will therefore be an estimate of the state probability distribution induced by the estimated transition intensities: Further, after simulation of the temporal distribution of state occupancy, we will show the distribution across states in a certain *order* of the states:

```

> prm <- c(1:4,8:5)
> levels( St2$lex.Cst )[prm]
[1] "DM"      "1st CVD"  "2nd CVD"  "3+ CVD"   "D(3+ CVD)" "D(2 CVD)" "D(1 CVD)"
[8] "D(no CVD)"

```

```

> load( file="../data/sim2.Rda" )
> summary( sim2, by="allocation" )

```

\$Intensive

Transitions:

From	To									Records:
	DM	1st CVD	2nd CVD	3+ CVD	D(no CVD)	D(1 CVD)	D(2 CVD)	D(3+ CVD)		
DM	4970	7374	0	0	3656	0	0	0	0	16000
1st CVD	0	1593	3263	0	0	2518	0	0	0	7374
2nd CVD	0	0	678	1425	0	0	1160	0	0	3263
3+ CVD	0	0	0	477	0	0	0	0	948	1425
Sum	4970	8967	3941	1902	3656	2518	1160	948		28062

Transitions:

From	To		
	Events:	Risk time:	Persons:
DM	11030	206419.26	16000
1st CVD	5781	36158.20	7374
2nd CVD	2585	10399.24	3263
3+ CVD	948	4648.26	1425
Sum	20344	257624.96	16000

\$Conventional

Transitions:

From	To									Records:
	DM	1st CVD	2nd CVD	3+ CVD	D(no CVD)	D(1 CVD)	D(2 CVD)	D(3+ CVD)		
DM	2647	10261	0	0	3092	0	0	0	0	16000
1st CVD	0	1115	6297	0	0	2849	0	0	0	10261
2nd CVD	0	0	675	3747	0	0	1875	0	0	6297
3+ CVD	0	0	0	1030	0	0	0	0	2717	3747
Sum	2647	11376	6972	4777	3092	2849	1875	2717		36305

Transitions:

From	To		
	Events:	Risk time:	Persons:
DM	13353	162159.43	16000
1st CVD	9146	39737.00	10261
2nd CVD	5622	15918.40	6297
3+ CVD	2717	12806.19	3747
Sum	30838	230621.02	16000

```

> system.time(
+ StC <- nState( subset(sim2,allocation=="Conventional"),
+               at=seq(0,20,0.1), from=0, time.scale="tsb" ) )

```

```

    user system elapsed
13.727  0.000 13.724
> StI <- nState( subset(sim2,allocation=="Intensive"),
+               at=seq(0,20,0.1), from=0, time.scale="tsb" )
> pC <- pState( StC, perm=prm )
> pI <- pState( StI, perm=prm )
> system.time(
+ StCi <- nState( subset(sim2i,allocation=="Conventional"),
+               at=seq(0,20,0.1), from=0, time.scale="tsb" ) )
    user system elapsed
11.942  0.000 11.936
> StIi <- nState( subset(sim2i,allocation=="Intensive"),
+               at=seq(0,20,0.1), from=0, time.scale="tsb" )
> pCi <- pState( StCi, perm=prm )
> pIi <- pState( StIi, perm=prm )
> save( pC, pI, pCi, pIi, file="../data/pSt2.Rda" )

> load( file="../data/pSt2.Rda" )
> head( pC )
      State
when    DM    1st CVD    2nd CVD    3+ CVD D(3+ CVD) D(2 CVD) D(1 CVD) D(no CVD)
0  1.0000000 1.0000000 1.0000000 1.0000000 1.0000000 1.0000000 1.0000000 1
0.1 0.9925000 0.9993125 0.9993125 0.9993750 0.9993750 0.9993750 0.9993750 1
0.2 0.9848125 0.9988125 0.9988750 0.9989375 0.9989375 0.9989375 0.9990000 1
0.3 0.9795000 0.9980625 0.9985000 0.9986250 0.9986250 0.9986250 0.9988125 1
0.4 0.9727500 0.9971875 0.9981250 0.9982500 0.9982500 0.9982500 0.9984375 1
0.5 0.9652500 0.9963125 0.9976875 0.9979375 0.9979375 0.9979375 0.9981875 1

> ppp <- function( pI=pI, pC=pC, rv=FALSE, ln=FALSE)
+ {
+ par( mfrow=c(1,2), mar=c(0,3,0,0), oma=c(3,0,2,3-2*rv), las=1 )
+
+ (c50I <- approx(pI[,1],as.numeric(rownames(pI)),0.5)$y)
+ (c50C <- approx(pC[,1],as.numeric(rownames(pI)),0.5)$y)
+ (d50C <- approx(pC[,4],as.numeric(rownames(pI)),0.5)$y)
+
+ plot( pI, col=clr[prm], xlab="" )
+ lines( as.numeric(rownames(pI)), pI[,4], col="black", lwd=4 )
+ if( ln ) {
+   abline( h=0.5, col="gray" )
+   segments(c50I,0,c50I,0.5,col="gray")
+ }
+ axis( side=4, at=0:5/5 )
+ axis( side=4, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Intensive", side=3, line=1 )
+
+ plot( pC, col=clr[prm], xlim=if(rv) c(20,0) else c(0,20),
+       xlab="", yaxt="n", ylab="", col.lab="transparent" )
+ lines( as.numeric(rownames(pC)), pC[,4], col="black", lwd=4 )
+ if( ln ) {
+   abline(h=0.5,col="gray")
+   segments(c50C,0,c50C,0.5,col="gray")
+   segments(d50C,0,d50C,0.5,col="gray")
+ }
+ axis( side=4-2*rv, at=1:5/5, if(rv) labels=NA )
+ axis( side=4-2*rv, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4-2*rv, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4-2*rv, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Conventional", side=3, line=1 )
+
+ mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
+ }
> ppp(pI,pC,ln=TRUE,rv=TRUE)

```

```

> ppp(pIi,pCi,rv=TRUE,ln=TRUE)

> postscript( file="../graph/Fig1.eps", height=6, width=8 )
> ppp(pI,pC)
> dev.off()
null device
  1

> postscript( file="../graph/Fig1r.eps", height=6, width=8 )
> ppp(pI,pC,rv=TRUE)
> dev.off()
null device
  1

> ppp(pIi,pCi,rv=TRUE)

```

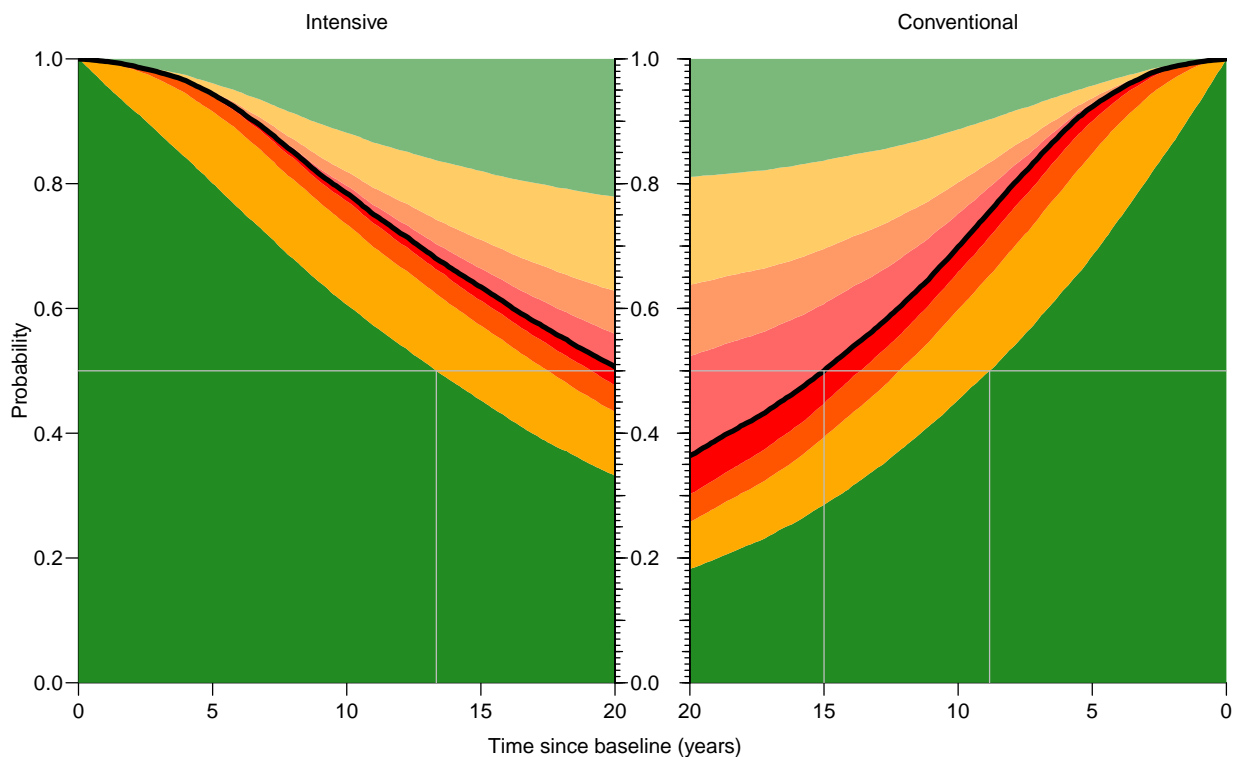


Figure 4.4: State distribution for a patient population with the same age- and sex-distribution as the Steno 2 population using models with constant intervention effect across CVD states. The colors of the states correspond to the colouring in figure 4.1.

4.4 Age- and sex specific results

We also want to make predictions conditional on the age at entry; and separately for men and women, because these two variables have major impact on both mortality and CVD-rates, and in particular because these are more likely to be generalizable — the overall figures derived above are essentially only valid for a patient population of the exact same age and sex composition as the Steno 2 population.

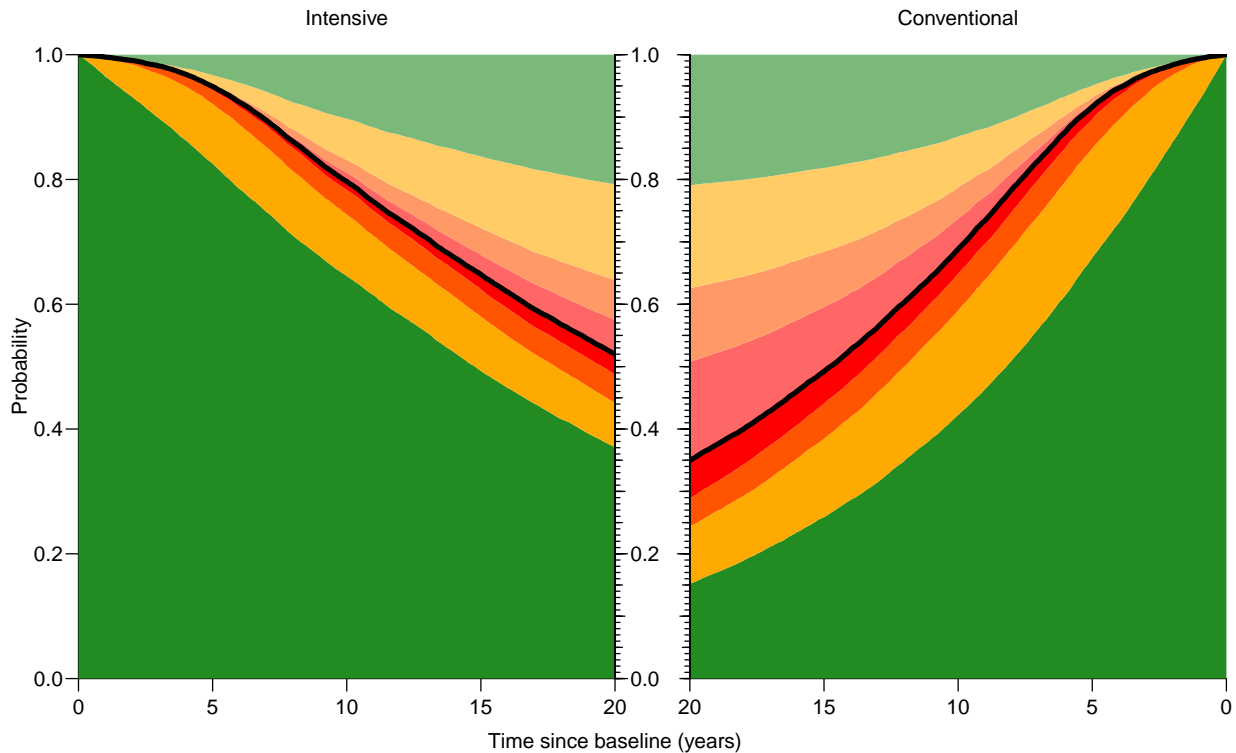


Figure 4.5: *State distribution for a patient population with the same age- and sex-distribution as the Steno 2 population using models with constant intervention effect across CVD states. The colors of the states correspond to the colouring in figure 4.1. This is the same plot as in figure 4.4, except that the time axis is reversed for the conventional group.*

Thus we simulate the life-course for a sample of 10,000 identical patients for each combination of allocation, sex and 5 different ages at entry (45, 50, 55, 60, 65) covering the range of entry age to the Steno2 study.

The results in the form of `pState` objects will be stored in an array, that is a list with a `dim` attribute of length 3, corresponding to the three variables (allocation, age and sex). The elements of the list is then filled in a loop over the dimensions:

```
> lp <- list()
> length( lp ) <- 20
> dim( lp ) <- c(5,2,2)
> dimnames( lp ) <- list( age = seq(45,65,5),
+                          sex = levels(L0$sex),
+                          grp = levels(L0$allocation) )
```

We make a copy of this to hold the results from simulations from the models with varying intervention effects:

```
> lpi <- lp
```

Then we can fill the two lists with `pState` objects from simulations, but we first need to define a starting Lexis object (a person with characteristics that we want to simulate). All persons are taken to start at 0 (since baseline) and in the DM state. Since the Lexis object also contains the timescales per and dur, we must define values for these too, but the particular values chosen for these will have no effect on the results, because the models for the transitions do not depend on these.

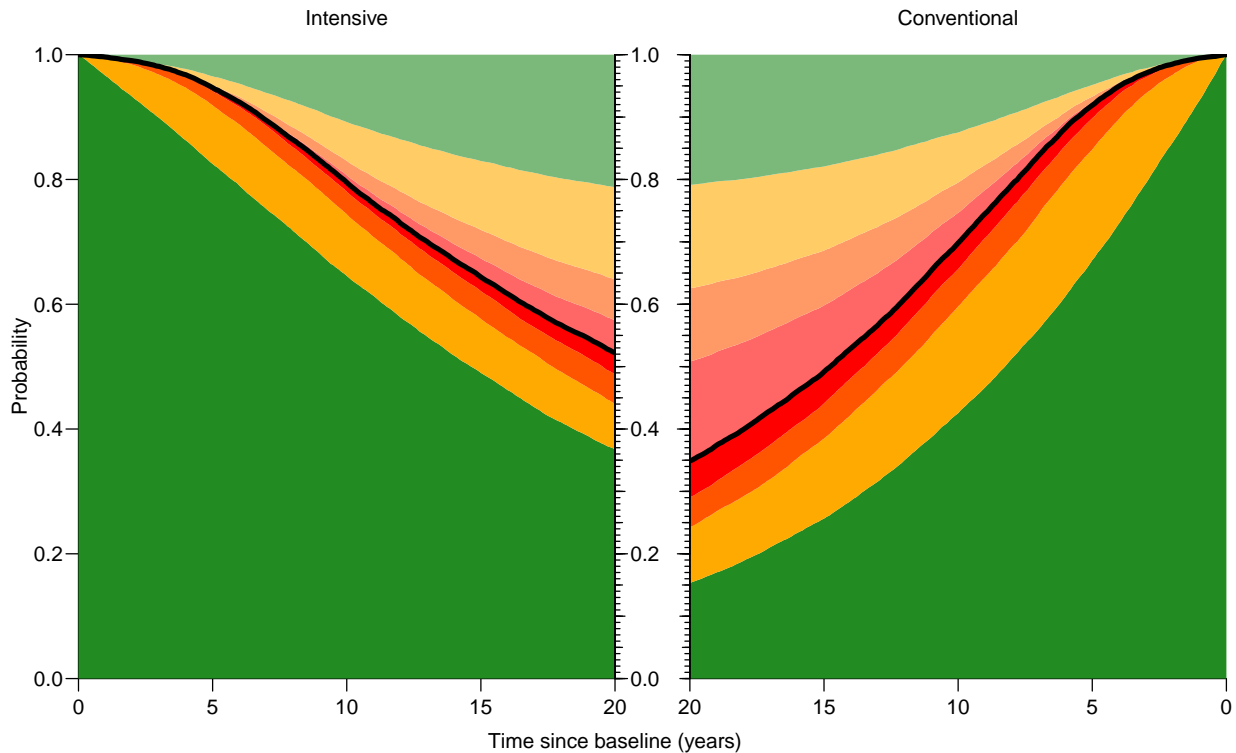


Figure 4.6: State distribution for a patient population with the same age- and sex-distribution as the Steno 2 population using models with different intervention effect across CVD states. The colors of the states correspond to the colouring in figure 4.1.

```
> Sb <- St2[NULL,]
> Sb[1,"tsb"] <- 0
> Sb[1,"per"] <- 1993
> Sb[1,"dur"] <- 5
> Sb[1,"lex.Cst"] <- "DM"
```

Once this is in place, we can simulate and compute the state distribution at different levels; note that we specify the starting age, sex and allocation inside the loop, since these are the values of the loop variables:

```
> system.time(
+ for( ig in dimnames(lp)[[3]] )
+ for( ia in dimnames(lp)[[1]] )
+ for( is in dimnames(lp)[[2]] )
+ {
+   Sb[1,"age"] <- as.numeric(ia)
+   Sb[1,"sex"] <- is
+   Sb[1,"allocation"] <- ig
+   stp <- simLexis( Tr = TM1,
+                   init = Sb,
+                   N = 5000,
+                   time.pts = seq(0,21,0.1) )
+   nst <- nState( stp, at=seq(0,20,0.1), from=0, time.scale="tsb" )
+   pst <- pState( nst, perm=prm )
+   lp[[ia,is,ig]] <- pst
+   stp <- simLexis( Tr = TM1i,
+                   init = Sb,
+                   N = 5000,
+                   time.pts = seq(0,21,0.1) )
+   nst <- nState( stp, at=seq(0,20,0.1), from=0, time.scale="tsb" )
+   pst <- pState( nst, perm=prm )
```

```
+ lpi[[ia,is,ig]] <- pst
+   } )
  user system elapsed
874.591  0.029 874.463
> save( lp, lpi, file="../data/lp.Rda" )

> load( file="../data/lp.Rda" )
```

The object `lp` is now a list with 3 dimensions (intervention, age, sex), where each element is a `pState` object, so that we can refer easily to results for any combination of age, sex and allocation.

4.4.1 State distribution

Using the `lp` and `lpi` objects we can now compare the state distribution between persons allocated to intervention and conventional, separately for all ten combinations of age (45, 50, 55, 60, 65) and sex (M, F).

Basically it is the same graph as for the Steno2 population

```
> pl10 <- function(lp){
+ par( mfrow=c(5,4), mar=c(0,0,0,3), oma=c(4,4,4,0), las=1 )
+ for( ia in dimnames(lp)[["age"]] )
+ for( is in dimnames(lp)[["sex"]][2:1] )
+ for( ig in dimnames(lp)[["grp"]] )
+   {
+ oax <- ( ig==dimnames(lp)[["grp"]][1] )
+ ptmp <- lp[[ia,is,ig]]
+ plot( ptmp, col=clr[prm], xaxt="n", yaxt="n",
+       xlim=if( oax ) c(0,20) else c(20,0) )
+ if( ia=="45" ) mtext( ig, side=3, line=1, at=10, outer=FALSE )
+ if( is=="M" & ig=="Intensive" )
+   {
+ axis( side=4-2*oax, at=1:5/5 )
+ # mtext( ia, side=2, line=3, outer=FALSE )
+ }
+ if( is=="F" & ig=="Conventional" ) mtext(ia,side=4,at=0.5,line=0.5,las=1)
+ axis( side=2, labels=NA )
+ if( ia=="65" )
+   {
+ axis( side=1 )
+ axis( side=4, at=0, labels="0.0" )
+ if( ia=="65" & is=="M" & oax ) axis( side=2, at=0, labels="0.0" )
+ }
+ lines( as.numeric(rownames(ptmp)), ptmp[,4], col="black", lwd=4 )
+ # text( 1, 0.05, paste( is, ", ", ia, "\n", ig, esp="" ),
+ #       adj=c(1-oax,0), font=2, col="white", cex=1.5 )
+ axis( side=2+2*oax, at=1:5*20 /100, if(oax) labels=NA )
+ axis( side=2+2*oax, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=2+2*oax, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=2+2*oax, at=0:100 /100, tcl=-0.2, labels=NA )
+   }
+ #mtext( "Men" , side=3, at=0.25, outer=TRUE, line=2 )
+ #mtext( "Women", side=3, at=0.75, outer=TRUE, line=2 )
+ #mtext( "Age at entry", side=3, at=-0.02, outer=TRUE, line=1, adj=0 )
+ mtext( "Time since entry (years)", side=1, outer=TRUE, line=2.5 )
+ mtext( "Probability" , side=2, outer=TRUE, line=2.5, las=0 )
+ mtext( "Men" , side=3, at=0.25, outer=TRUE, line=2.5, las=0 )
+ mtext( "Women", side=3, at=0.75, outer=TRUE, line=2.5, las=0 )
+ mtext( "Age" , side=3, at=1.00, outer=TRUE, line=2.5, las=0, adj=1 )
+ }
> pl10(lp)

> pl10(lpi)
```


4.5 Cumulative risks

From the `pState` objects we can derive the fraction of patients at say 10 and 20 years after baseline that are alive, alive without CVD, who ever had CVD (live or dead), which for the Steno 2 population will illustrate the general effect of the intervention, and for the specific age and sex groups how the intervention may be expected to work in other settings.

There are two main outcomes of which we would like to know: death and CVD; in terms of figure 4.1 “Death” is the probability of being in any of the right hand boxes; “CVD” is the probability of being in any of the lower 6 boxes (regardless of being dead or alive). We can make a table for the Steno population, and a table for the 10 combinations of age and sex we defined.

First the calculations for the Steno2 population — essentially just sums/differences between probabilities in different states:

```
> cr2 <- NArray( list( type = c("Death", "CVD"),
+                       when = rownames(pI),
+                       grp = c("Intensive", "Conventional", "Int-Conv") ) )
> cr2["Death",, "Intensive" ] <- 1-pI[,4]
> cr2["Death",, "Conventional"] <- 1-pC[,4]
> cr2["CVD" ,, "Intensive" ] <- pI[,7]-pI[,1]
> cr2["CVD" ,, "Conventional"] <- pC[,7]-pC[,1]
> cr2[,, "Int-Conv"] <- cr2[,, "Intensive"] - cr2[,, "Conventional"]
> round( ftable(cr2[,c("10", "20"),]) * 100, 1 )
      grp Intensive Conventional Int-Conv
type  when
Death 10          21.5          30.1         -8.6
      20          49.4          63.6        -14.2
CVD   10          27.5          43.4        -15.8
      20          44.7          62.8        -18.1
```

...and then the same for the 10 combinations of age and sex:

```
> cras <- NArray( list( age = seq(45,65,5),
+                       sex = c("M", "F"),
+                       type = c("Death", "CVD"),
+                       when = dimnames(lp[[1]])[["when"]],
+                       grp = c("Intensive", "Conventional", "Int-Conv") ) )
> for( ia in dimnames(cras)[["age"]] )
+ for( is in dimnames(cras)[["sex"]] )
+ for( ig in dimnames(cras)[["grp"]][-3] )
+ {
+   cras[ia,is,"Death",,ig] <- 1-lp[[ia,is,ig]][,4]
+   cras[ia,is,"CVD" ,,ig] <- lp[[ia,is,ig]][,7]-
+   lp[[ia,is,ig]][,1]
+ }
> cras[,,,,3] <- cras[,,,,2] - cras[,,,,1]
> round( ftable( cras[,,,paste(c(10,20)),],
+               col.vars=c(3,5),
+               row.vars=c(4,2,1) ) * 100, 1 )
      type      Death      CVD
      grp  Intensive Conventional Int-Conv Intensive Conventional Int-Conv
when sex age
10  M  45          6.9          10.6          3.7          20.9          34.0          13.2
      50          11.5          17.5          6.0          24.1          38.9          14.8
      55          18.5          28.2          9.7          26.8          44.7          17.9
      60          30.4          44.9          14.5          32.9          50.0          17.0
      65          46.8          61.2          14.4          37.0          54.5          17.4
      F  45          4.8          6.5          1.7          16.2          26.0          9.8
      50          7.9          11.6          3.7          18.5          32.9          14.4
      55          13.0          18.9          5.9          23.8          37.4          13.6
      60          21.3          30.2          8.9          26.3          42.0          15.7
      65          32.8          47.3          14.5          30.1          47.9          17.9
```

20	M	45	21.7	32.6	11.0	39.2	57.7	18.5
		50	33.8	48.9	15.1	43.2	62.4	19.2
		55	49.8	68.0	18.1	45.1	65.5	20.4
		60	69.2	84.6	15.4	50.6	66.8	16.2
		65	84.3	93.5	9.2	50.9	67.0	16.1
F	45	14.8	20.8	6.0	31.8	47.0	15.2	
	50	24.3	35.3	11.0	36.4	55.1	18.7	
	55	37.5	52.0	14.5	41.2	59.2	17.9	
	60	55.3	70.5	15.2	43.5	61.5	18.0	
	65	70.2	86.1	15.9	46.0	63.7	17.7	

The above table gives the cumulative risks of death and CVD respectively in percent, computed at 10 and 20 years after baseline, but we could of course also show the cumulative risks as curves; first for the Steno 2 population:

```
> tt <- as.numeric( dimnames(cr2)[[2]] )
> par( mfrow=c(1,2), mar=c(2,3,1,1), oma=c(2,0,0,0), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> for( tp in c("Death","CVD") )
+   {
+ matplot( tt, cr2[tp,,1]*100,
+         type="l", lty=1, lwd=4, col="black",
+         ylim=c(0,100), xlab="", yaxs="i", ylab=paste("Cumulative risk of", tp) )
+ matlines( tt, cr2[tp,,2]*100,
+          type="l", lty="11", lwd=4, lend="butt", col="black" )
+   }
> mtext( "Time since baseline (years)", side=1, line=0, outer=TRUE )

> par( mfrow=c(2,2), mar=c(2,3,1,1), oma=c(2,0,0,0), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> for( sx in c("M","F") )
+ for( tp in c("Death","CVD") )
+   {
+ matplot( tt, t( rbind( cras[,sx,tp,,"Intensive"] ) ) *100,
+         type="l", lty=1, lwd=1:5, col="black",
+         ylim=c(0,100), xlab="", yaxs="i", ylab=paste("Cumulative risk of",tp) )
+ matlines( tt, t( rbind( cras[,sx,tp,,"Conventional"] ) ) *100,
+          type="l", lty=c("AA","55","43","32","22"), lend="butt", lwd=1:5, col="black" )
+   text( 0, 90, sx, font=2, cex=1.5, adj=0 )
+   }
> mtext( "Time since baseline (years)", side=1, line=0, outer=TRUE )

> par( mfrow=c(2,2), mar=c(2,1,1,2), oma=c(1,2,0,0), mgp=c(3,1,0)/1.6, bty="n", las=1 )
> for( sx in c("M","F") )
+ for( tp in c("Death","CVD") )
+   {
+ matplot( tt, t( rbind( cras[,sx,tp,,"Intensive"] ) ) *100,
+         type="l", lty=1, lwd=1:5, col="black",
+         xlim=c(0, 20), yaxs="i", xlab="",
+         ylim=c(0,100), yaxs="i", ylab="" )
+ matlines( tt, t( rbind( cras[,sx,tp,,"Conventional"] ) ) *100,
+          type="l", lty=c("AA","55","43","32","22"), lend="butt", lwd=1:5, col="black" )
+ axis( side=4, at=0:5*20, labels=if(tp=="Death") NA else 0:5*20 )
+ axis( side=4, at=0:10*10, tcl=-0.4, labels=NA )
+ axis( side=4, at=0:20*5, tcl=-0.3, labels=NA )
+ axis( side=4, at=0:100, tcl=-0.2, labels=NA )
+   }
> mtext( "Time since baseline (years)", side=1, line=0, outer=TRUE )
> mtext( c("Women","Cumulative risk (%)","Men"), at=1:3/4, side=2, line=1, outer=TRUE, las=0 )
> mtext( c("Death","CVD"), at=c(1,3)/4, side=3, line=-1, outer=TRUE, las=0 )
```

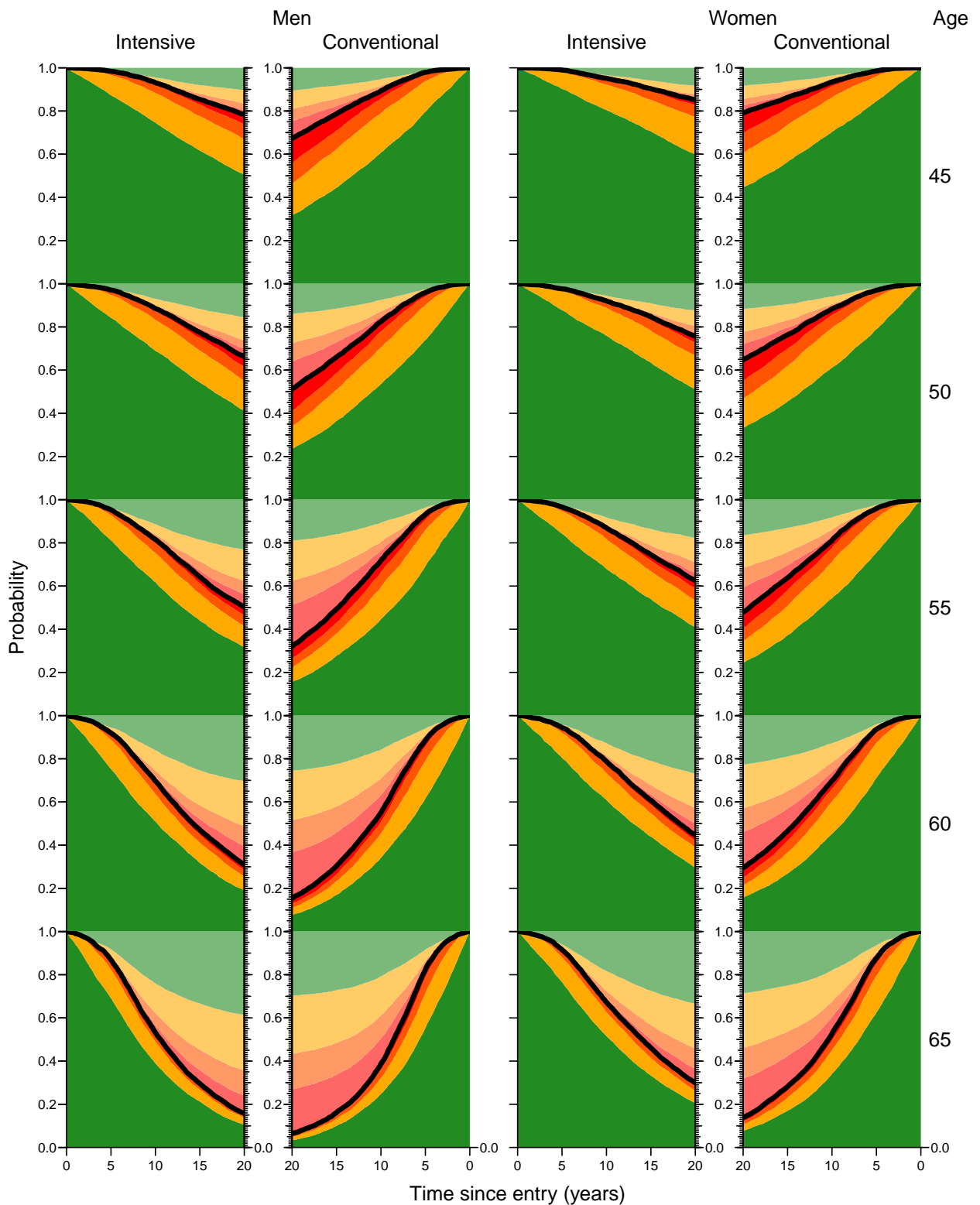


Figure 4.7: Probabilities of being in different states by age, sex and intervention group, based on models with the same intervention effect across CVD strata. State colouring is the same as in figure 4.1.

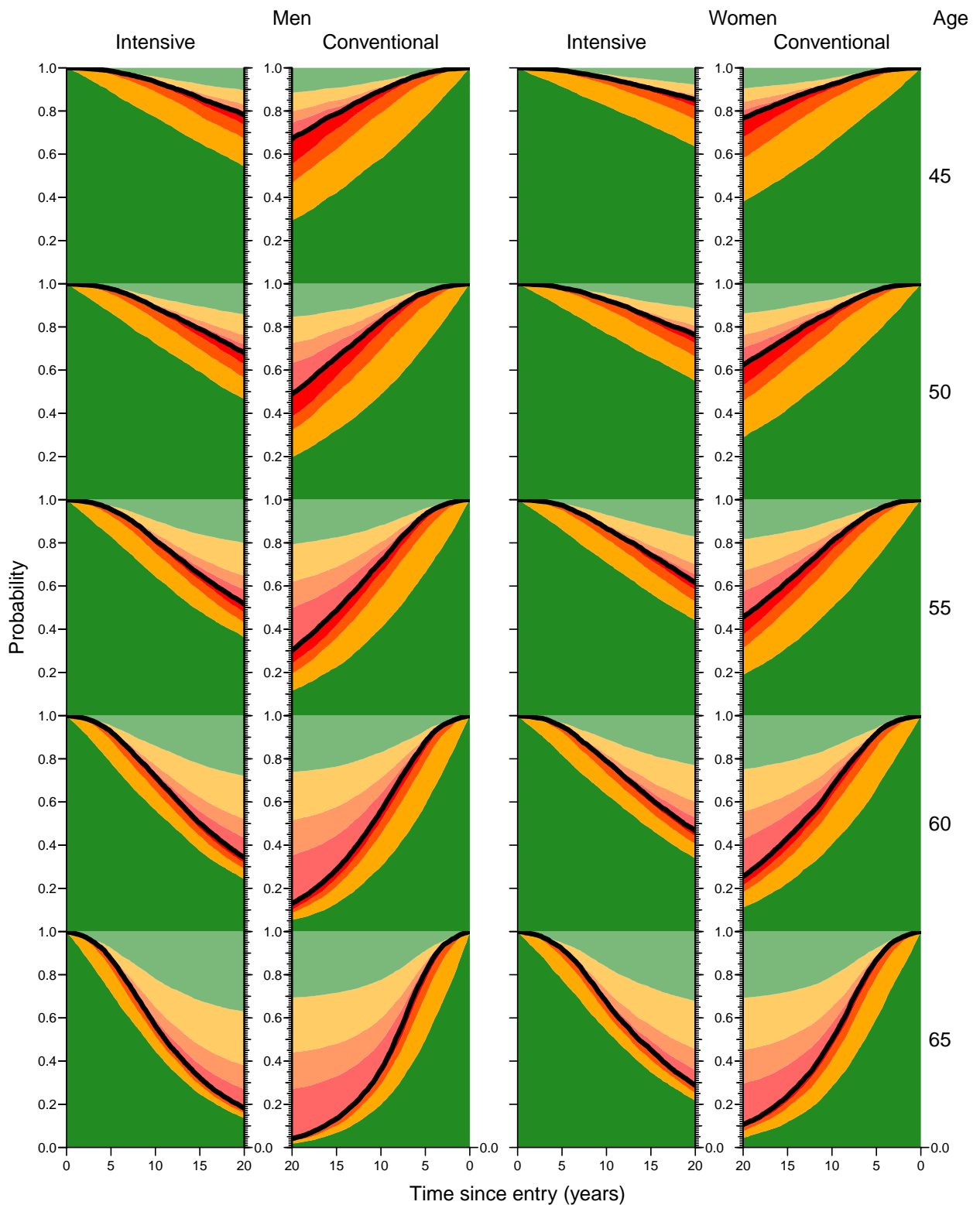


Figure 4.8: Probabilities of being in different states by age, sex and intervention group, based on models with the differential intervention effect across CVD strata. State colouring is the same as in figure 4.1.

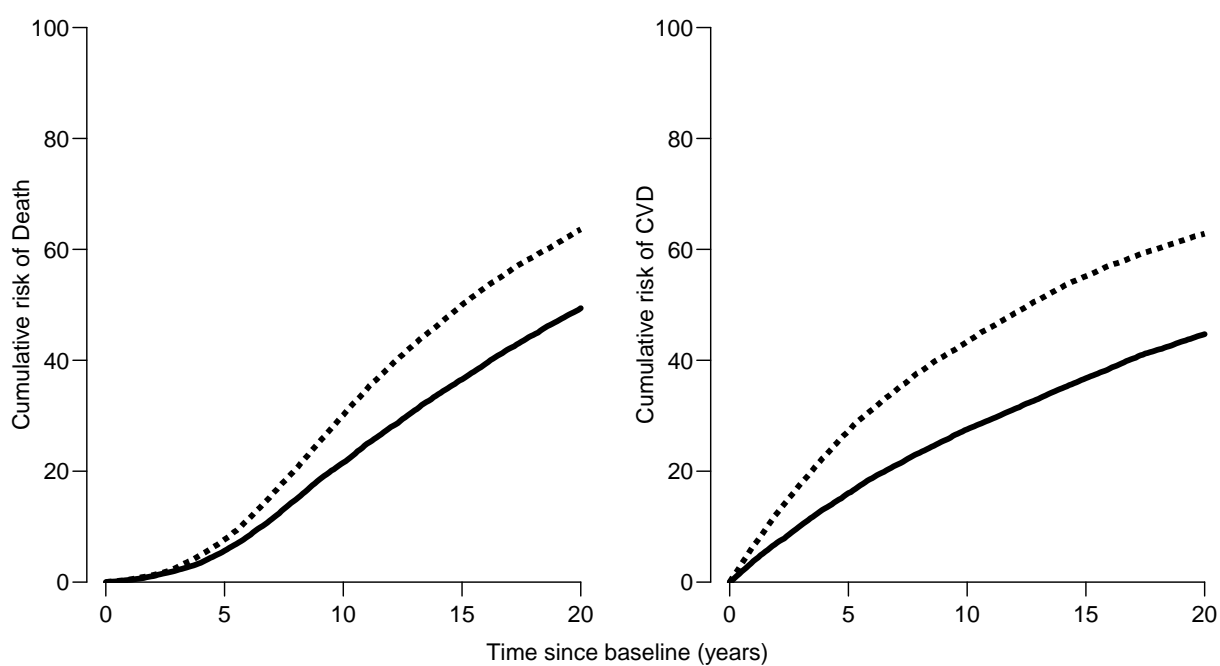


Figure 4.9: *Cumulative risk of death and CVD in the Steno 2 study, as estimated from the multistate model. The full lines is the intensive group; the broken the conventional group.*

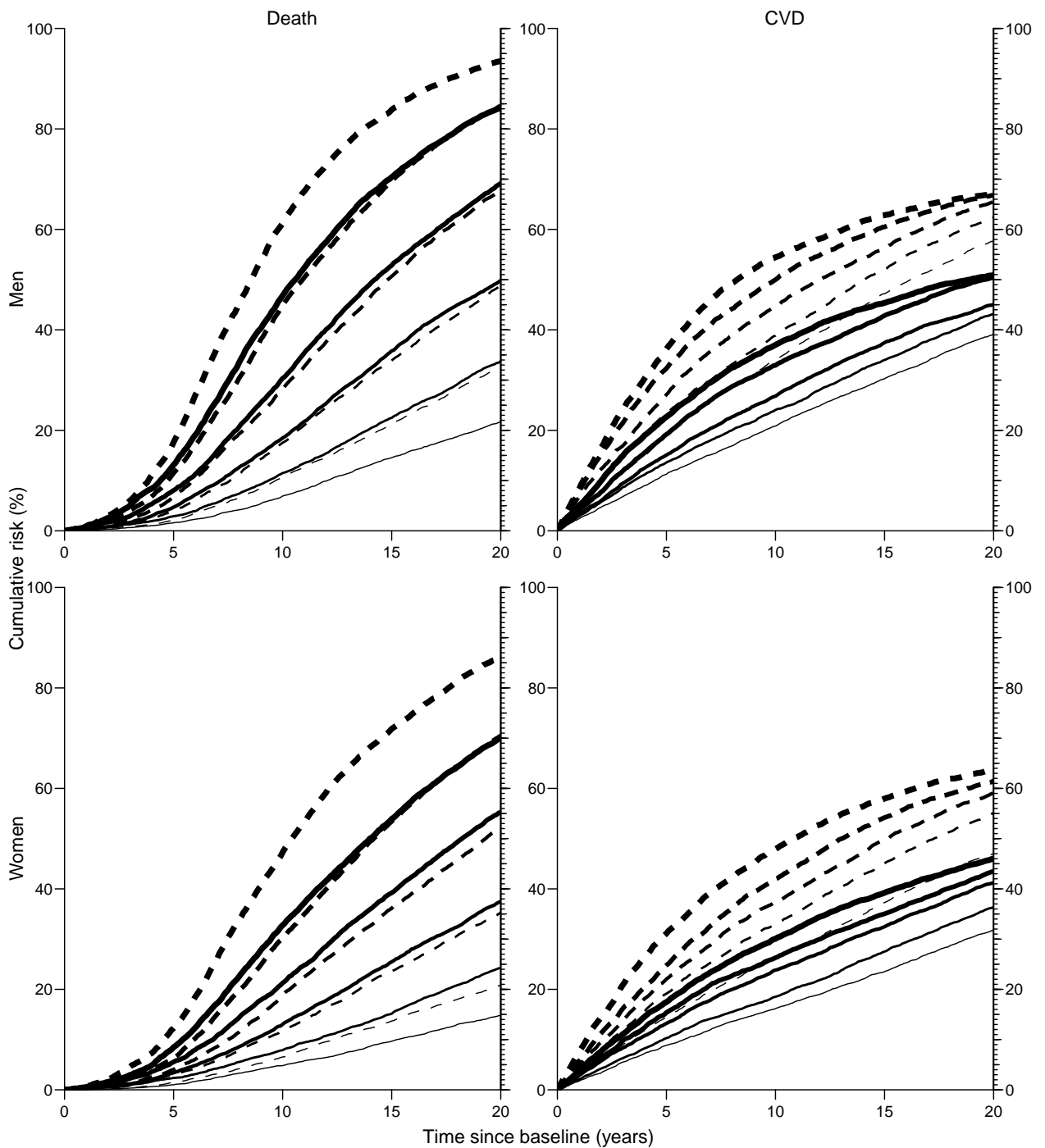


Figure 4.10: Cumulative risk of death (left) and CVD (right) in the Steno 2 study, as estimated from the multistate model. Top panels are men, bottom panels women; in order of thickness the lines refer to ages 45, 50, 55, 60 and 65 years at baseline. The full lines are for the intensive group; the broken for the conventional group.

4.6 Years lived with and without CVD

Here we need (for the sake of completeness) both the estimated state-distributions for the Steno2-study (in the file `pSt2.Rda`) and for selected combinations of age and sex (in the file `lp.Rda`):

```
> load( file="../data/pSt2.Rda" )
> load( file="../data/lp.Rda" )
```

We can now compute the sojourn time in each state during the first 20 years after randomization. This is really just a transformation of the matrix of state occupancy probabilities (the `pState` object), namely the integral of the probabilities over time. We devise a small function that does this job; note that the function returns a matrix of the same dimension as the `pState` object; it contains the time spent in each state up to a given time:

```
> cum.area <-
+ function( pp )
+ {
+   if( !inherits(pp,"pState") ) warning( "Not a pState object" )
+   il <- diff( as.numeric(rownames(pp)) )
+   st <- pp * 0
+   pp <- cbind(0,pp) # makes the code simpler
+   for( i in 2:ncol(pp) )
+     st[-1,i-1] <- cumsum( il * ( pp[-1,i ]-diff(pp[,i ])/2 -
+                                 ( pp[-1,i-1]-diff(pp[,i-1])/2 ) ) )
+   class( st ) <- "matrix"
+   st
+ }
> cum.area
function( pp )
{
if( !inherits(pp,"pState") ) warning( "Not a pState object" )
il <- diff( as.numeric(rownames(pp)) )
st <- pp * 0
pp <- cbind(0,pp) # makes the code simpler
for( i in 2:ncol(pp) )
  st[-1,i-1] <- cumsum( il * ( pp[-1,i ]-diff(pp[,i ])/2 -
                              ( pp[-1,i-1]-diff(pp[,i-1])/2 ) ) )

class( st ) <- "matrix"
st
}
> yI <- cum.area( pI )
> yC <- cum.area( pC )
> lc <- lapply( lp, cum.area )
> lci <- lapply( lpi, cum.area )
> dim( lc ) <- dim( lci ) <- dim( lp )
> dimnames( lc ) <- dimnames( lci ) <- dimnames( lp )
```

With these calculations fixed we can show the years lived with and without CVD in the two groups during the first 20 years of follow-up, and of course also the difference between the groups. First the calculation for the Steno2 population where we get the number of years lived in total, free of CVD, with 1 CVD event (which may not be of much relevance) and with any number of CVD events:

```
> y2 <- NArray( c( list( state = c("Alive","No CVD","1st CVD","Any CVD"),
+                               grp = c(dimnames(lc)[[3]],"Int-Ctr") ) ) )
> y2["No CVD",-3] <- c( yI["20","DM"],
+                       yC["20","DM"] )
> y2["1st CVD",-3] <- c( yI["20","1st CVD"],
+                       yI["20","1st CVD"] )
```

```

> y2["Any CVD",-3] <- c( yI["20","1st CVD"]+yI["20","2nd CVD"]+yI["20","3+ CVD"],
+                      yC["20","1st CVD"]+yC["20","2nd CVD"]+yC["20","3+ CVD"] )
> y2["Alive,] <- y2["No CVD,] + y2["Any CVD,]
> y2[, "Int-Ctr"] <- y2[, "Intensive"] - y2[, "Conventional"]
> round( ftable( y2 ), 1 )

```

	grp Intensive	Conventional	Int-Ctr
state			
Alive	15.6	14.1	1.5
No CVD	12.6	10.0	2.6
1st CVD	2.2	2.2	0.0
Any CVD	3.0	4.1	-1.1

Thus we see that the total years gained during the 20 first years of follow-up is 1.6, but 2.6 CVD-free years so the intensive group has actually *fewer* years lived with CVD.

However, these numbers merely tend to illustrate the effects on the specific Steno2 distribution, and they are not generalizable to populations with other age-and sex-distributions. Therefore, we also show results for specific age-groups for each sex.

```

> yl <- NArray( c( dimnames(lc)[-3],
+                 list( state = c("Alive","No CVD","1st CVD","Any CVD"),
+                       grp = c(dimnames(lc)[[3]], "Int-Ctr") ) ) )
> for( ia in dimnames(lc)[[1]] )
+ for( is in dimnames(lc)[[2]] )
+ for( ig in dimnames(lc)[[3]] )
+ {
+   yl[ia,is,"No CVD" ,ig] <- lc[[ia,is,ig]]["20","DM"]
+   yl[ia,is,"1st CVD",ig] <- lc[[ia,is,ig]]["20","1st CVD"]
+   yl[ia,is,"Any CVD",ig] <- lc[[ia,is,ig]]["20","1st CVD"] +
+     lc[[ia,is,ig]]["20","2nd CVD"] +
+     lc[[ia,is,ig]]["20","3+ CVD"]
+ }
> yl[,,"Alive,] <- yl[,,"No CVD,] + yl[,,"Any CVD,]
> yl[,,"Int-Ctr"] <- yl[,,"Intensive"] - yl[,,"Conventional"]
> round( ftable( yl[,,-3,], row.vars=c(3,1) ), 1 )

```

state	age	sex			M		
		grp Intensive	Conventional	Int-Ctr	Intensive	Conventional	Int-Ctr
Alive	45	18.9	18.4	0.5	18.3	17.5	0.8
	50	18.1	17.2	0.9	17.4	16.0	1.3
	55	17.0	15.8	1.2	15.8	14.1	1.8
	60	15.4	13.8	1.6	13.8	11.7	2.1
	65	13.5	11.3	2.3	11.4	9.5	1.9
No CVD	45	16.1	14.2	1.9	14.9	12.5	2.4
	50	15.1	12.6	2.6	13.9	11.2	2.7
	55	13.8	11.4	2.4	12.6	9.8	2.8
	60	12.5	9.9	2.5	10.8	8.2	2.6
	65	11.0	8.2	2.8	9.1	6.9	2.2
Any CVD	45	2.8	4.3	-1.4	3.4	5.0	-1.6
	50	3.0	4.7	-1.7	3.5	4.8	-1.4
	55	3.2	4.4	-1.2	3.2	4.2	-1.0
	60	2.9	3.8	-0.9	2.9	3.5	-0.5
	65	2.5	3.1	-0.6	2.3	2.6	-0.3

Thus we see that the extra years gained by the intensive during the first 20 years is between 0.5 and 2.8; largest among men and increasing by age at entry to the study. The years lived without any CVD is about 2.5 years, largely the same in all age-classes and both for men and women. This means that the years alive with CVD is largest in the conventional group about 1.5 years larger for persons entering at age 45 decreasing to about 0.5 years for persons entering in age 65.

Thus, the major treatment results seem to be about 2 more years of life, and 2.5 years longer without CVD during the first 20 years of follow-up.

Chapter 5

Microvascular events

The microvascular outcomes are measured at 6 different visits, and changes of status are therefore inherently *interval censored*. We will impute exact transition times using a random uniformly distributed time between the two visits; and we shall look into the effect of the imputation by assessing the difference between results from different imputations.

5.1 Useful functions for interval censoring

To this end we have devised functions to perform some of the tasks:

```
> source( "usefuns.R" )
```

The function `rpart` generates a vector with `a` as the first element, `b` as the last and `n` uniformly distributed points between them. This is intended for generating the transition times through (possibly more than one) intermediate step, when only the status at visit times `a` and `b` are known:

```
> rpart
function(n,a,b) a+(b-a)*c(0,sort(runif(n)),1)
> rpart( 2, 3, 7 )
[1] 3.00000 4.30165 5.76567 7.00000
> rpart( 2, 2, 7 )
[1] 2.000000 6.646254 6.937107 7.000000
```

The function `lx.gen` generates a data frame with the skeleton of a `Lexis` object “by hand”, imputing all intermediate states and transition times:

```
> lx.gen
function( st, dt, id, incr=TRUE )
{
# Generate Lexis records for one person in states st at dates dt.
#
# st is a vector of states (possibly with missing) - rows of rt
# dt is a vector dates of visits - rows of rd
# id is a number, used for labeling (as lex.id) - rownames of these
# incr indicates if only increasing states should be allowed - meaning
# that all decreases in states are overridden by the previous larger,
# effectively redefining state to "highest state achieved thus far".
# res.df is the resulting dataframe (Lexis object) with follow-up of
# ONE person
res.df <- NULL
# only states and dates known
```

```

dt <- dt[!is.na(st)]
st <- st[!is.na(st)]
# number of points with known status
np <- length( st )
# only ever output anything substantial if more than one obs
if( np > 1 )
{
# only assume increasing states
if( incr ) for( i in 2:np ) st[i] <- max( st[i-0:1] )
# loop over states seen
for( i in 2:np )
{
  if( st[i]==st[i-1] )
  {
    lex.Cst <- lex.Xst <- st[i]
    lex.dur <- dt[i] - dt[i-1]
    per <- dt[i-1]
    res.df <- rbind( res.df,
                    data.frame( per=per, lex.dur, lex.Cst, lex.Xst, lex.id=id ) )
  }
  if( st[i]!=st[i-1] )
  {
    # no. of transitions seen
    nt <- abs(st[i]-st[i-1])
    tm <- rpart( nt, dt[i-1], dt[i] )
    lex.Cst <- st[i-1]:st[i]
    lex.Xst <- c(lex.Cst[-1],rev(lex.Cst)[1])
    lex.dur <- diff( tm )
    per <- tm[-length(tm)]
    res.df <- rbind( res.df,
                    data.frame( per=per, lex.dur, lex.Cst, lex.Xst, lex.id=id ) )
  }
}
}
} else res.df <- data.frame( per = dt, # The case of only one visit
                             lex.dur = 1/12, # risk is set to 1 month
                             lex.Cst = st,
                             lex.Xst = st,
                             lex.id = id )

res.df
}

```

The function `make.dlex` merges a “raw” object of transition times (generated by `lx.gen`) with dates of death and other relevant dates and other variables from the baseline visit in the data frame `mic`.

```

> make.dlex
function( rt, rd, incr=TRUE )
{
lx.new <- NULL
for( i in 1:nrow(rt) )
  lx.new <- rbind( lx.new,
                  lx.gen( rt[i,],
                          rd[i,],
                          id = as.numeric(rownames(rt)[i]),
                          incr = incr ) )
# Get the exit time and the death status
dd <- mic[,c("ID","doDth","doEnd")]
dd <- dd[match(unique(dd$ID),dd$ID),]
# The last record in the Lexis object of each person
ll <- merge( subset( lx.new, per==ave( per, lex.id, FUN=max ) ), dd,
            by.x="lex.id", by.y="ID", all.x=TRUE )
# Change this record to represent follow-up from last visit to death in last known state
ll$lex.Cst <- ll$lex.Xst
ll$per <- ll$per+ll$lex.dur
ll$lex.Xst <- ifelse( is.na(ll$doDth),

```

```

      ll$lex.Cst,
      paste("Dead(",ll$lex.Cst,")",sep="") )
ll$lex.dur <- pmin( ll$doDth, ll$doEnd, na.rm=TRUE ) - ll$per
ll <- subset( ll, lex.dur>0 )
lx.new <- rbind( lx.new, ll[,names(lx.new)] )
lx.new <- merge( lx.new,
               mic[mic$visit==1,c("ID","sex","doBth","doVis","doBlind","doEnd","doDth","allocation",
               by.x ="lex.id", by.y="ID", all.x=TRUE )
lx.new <- transform( lx.new,
                    age = per-doBth,
                    tfe = per-doVis )
attr( lx.new, "class" ) <- c("Lexis","data.frame")
attr( lx.new, "time.scales" ) <- c("per","age","tfe")
attr( lx.new, "time.since" ) <- c("","","")
ol <- with( lx.new, order(lex.id,per) )
Relevel( lx.new[ol,] )
}

```

With these functions in place we can now analyze occurrence of the three types of microvascular events.

5.2 Retinopathy

The analysis will be done for the maximal score of left and right eye, that is maximum of `eds_l` and `eds_r`.

5.2.1 Data overview

First we provide an overview of the actually occurring retinopathy patterns in the data:

```

> library( Epi )
> library( survival )
> clear()
> load( file="../data/mic.Rda" )
> names( mic )
 [1] "ID"          "allocation" "sex"         "doBth"      "doVis"      "doESRD"
 [7] "doEye"      "doPhC"      "doBlind"    "doEnd"      "doDth"      "visit"
[13] "aer_level"  "med_aer"    "eds_l"      "eds_r"      "autoprog"   "periprog"
> with( mic, ftable(visit,!is.na(eds_r),!is.na(eds_l),useNA="ifany",col.vars=c(1,3)) )
      visit      1      2      3      4      5      6
      FALSE TRUE FALSE TRUE FALSE TRUE FALSE TRUE FALSE TRUE FALSE TRUE
FALSE      0   3   92   1   0   1   0   1   14   1   62   0
TRUE       2 155   4   59   1 147   1 126   0   78   0   0
> with( mic, table( eds_l, eds_r ) )
      eds_r
eds_l  No retinopathy Minimal non-proliferative
No retinopathy          248                    35
Minimal non-proliferative 39                    96
Moderate nonproliferative  6                    14
Pre-proliferative         0                     0
Proliferative             0                     0
Photocoagulation          1                     3
      eds_r
eds_l  Moderate nonproliferative Pre-proliferative Proliferative
No retinopathy                   3                    0                    0
Minimal non-proliferative         8                    1                    3
Moderate nonproliferative        28                    2                    1
Pre-proliferative                 1                    7                    2
Proliferative                    2                    2                   11

```

```

Photocoagulation          eds_r          4          1          5
eds_l                    Photocoagulation
No retinopathy           1
Minimal non-proliferative 0
Moderate nonproliferative 5
Pre-proliferative        1
Proliferative            5
Photocoagulation         30
> with( mic, plot( jitter(as.integer(eds_r),2),
+               jitter(as.integer(eds_l),2), pch=16, cex=0.8 ) )

```

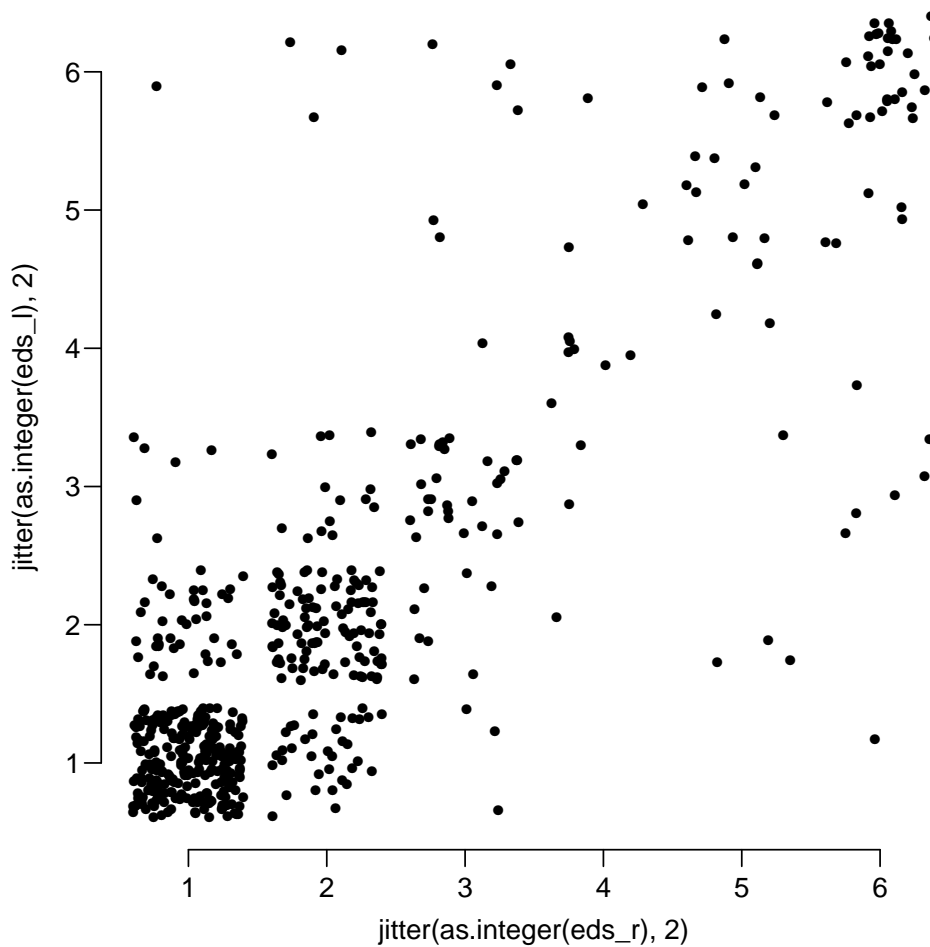


Figure 5.1: *Relationship between left and right edes-score, jittered*

For the sake of consistency we form a variable which is the maximum of the two sides.

```

> mic <- transform( mic, eds = pmax(as.integer(eds_r),
+                               as.integer(eds_l),na.rm=TRUE) )
> with( mic, table( eds ) )
eds
 1  2  3  4  5  6
259 170 59 11 22 59

```

We then generate the two 160×6 matrices of states (**rt**, maximal retinopathy score) and dates (**rd**):

```
> rd <- with( mic, tapply( doVis, list(ID,visit), mean ) )
> rt <- with( mic, tapply( eds, list(ID,visit), mean ) )
> cbind( rt, rd )[1:10,]
  1  2  3  4  5  6      1      2      3      4      5      6
1  1 NA  1  1  1 NA 1993.357 1995.158 1997.135 2001.097 2006.712 2014.802
2  2 NA  2  6  6 NA 1993.357 1995.377 1997.781 2001.814 2006.868 2014.589
3  2 NA  2  4 NA NA 1993.362 1995.180 1997.151 2001.102      NA      NA
4  1  1  1  1 NA NA 1993.362 1995.120 1997.110 2001.066      NA      NA
5  1 NA  1 NA NA NA 1993.162 1995.002 1997.707      NA      NA      NA
6  1 NA  3  3  6 NA 1993.201 1995.008 1997.036 2001.025 2006.736 2015.029
7  1  2  3  6 NA NA 1993.203 1995.043 1997.077 2001.058      NA      NA
8  2  2  2  2 NA 1993.209 1995.829 1997.729 2001.655 2006.903      NA
9  2  1 NA NA NA NA 1993.209 1995.021      NA      NA      NA      NA
10 1  2  2  6 NA NA 1993.075 1995.024 1997.020 2001.023 2006.682      NA

> table( rt[,1] )
  1  2  3  4  5  6
108 27 10  5  6  4
```

This gives the opportunity to generate the actually observed patterns of status:

```
> rr <- gsub("NA","-",apply( rt, 1, paste, collapse="" ))
> tt <- table( rr )
> rn <- gsub("-", "7",rownames( tt ))
> addmargins( tt[order(rn)] )
rr
11111- 11112- 1111-- 11121- 11122- 11123- 11126- 1112-- 11131- 111--- 11221- 11222-
  4      3      4      1      1      1      1      2      1      1      1      1
11223- 1122-- 1123-- 11266- 112--- 113--- 11-1-- 11---- 12222- 12223- 12224- 12226-
  1      1      1      1      2      1      2      1      1      1      1      2
1223-- 1226-- 122--- 1236-- 1322-- 133--- 13---- 1-111- 1-112- 1-11-- 1-122- 1-126-
  1      1      2      1      2      2      1      14     2      6      3      2
1-12-- 1-132- 1-136- 1-166- 1-1--- 1-212- 1-222- 1-22-- 1-232- 1-233- 1-23-- 1-26--
  2      2      1      1      2      1      4      3      1      1      2      1
1-2--- 1-323- 1-332- 1-336- 1-33-- 1-66-- 1--5-- 1----- 2112-- 21222- 2122-- 21----
  3      1      1      1      2      1      1      4      1      1      1      1
22112- 22222- 2222-- 222-6- 222--- 223--- 233--- 2-21-- 2-222- 2-225- 2-232- 2-236-
  1      2      1      1      1      1      1      1      1      1      1      2
2-23-- 2-24-- 2-266- 2-26-- 2-2--- 2-366- 2-3--- 2-6-6- 31322- 33---- 3-22-- 3-322-
  1      1      2      1      1      1      1      1      1      1      1      1
3-33-- 3-355- 3-466- 3-46-- 3-566- 3-66-- 43566- 45666- 4-466- 4-46-- 4-666- 555---
  1      1      1      1      1      1      1      1      1      1      1      2
5-55-- 5-65-- 5-66-- 6-66-- 6-6---      Sum
  2      1      1      2      2      160
```

5.2.2 Generating transitions

We again load the functions to generate the Lexis objects of transitions between the states:

```
> source( "usefuns.R" )
```

We now simulate exact transition times to generate data sets and inspect the difference between them:

```
> ret1 <- make.dlex( rt, rd, incr=TRUE )
> ret2 <- make.dlex( rt, rd, incr=FALSE )
> summary( ret1 )
Transitions:
  To
From  1  2  3  4  5  6 Dead(1) Dead(2) Dead(3) Dead(4) Dead(5) Dead(6) Records:
  1  159 70  0  0  0  0      21      0      0      0      0      0      250
  2   0 156 52  0  0  0      0      27      0      0      0      0      235
```

3	0	0	78	30	0	0	0	0	20	0	0	0	128
4	0	0	0	8	33	0	0	0	0	1	0	0	42
5	0	0	0	0	17	32	0	0	0	0	5	0	54
6	0	0	0	0	0	73	0	0	0	0	0	19	92
Sum	159	226	130	38	50	105	21	27	20	1	5	19	801

Transitions:

	To												
From	Events:	Risk time:	Persons:										
1	91	805.02	108										
2	79	702.75	97										
3	50	377.03	62										
4	34	68.01	35										
5	37	106.81	39										
6	19	357.99	36										
Sum	310	2417.60	160										

> summary(ret2)

Transitions:

	To													
From	1	2	3	4	5	6	Dead(1)	Dead(2)	Dead(3)	Dead(4)	Dead(5)	Dead(6)	Records:	
1	175	76	0	0	0	0	23	0	0	0	0	0	274	
2	11	165	54	0	0	0	0	29	0	0	0	0	259	
3	0	13	54	31	0	0	0	0	16	0	0	0	114	
4	0	0	1	7	33	0	0	0	0	1	0	0	42	
5	0	0	0	0	18	32	0	0	0	0	6	0	56	
6	0	0	0	0	1	72	0	0	0	0	0	18	91	
Sum	186	254	109	38	52	104	23	29	16	1	6	18	836	

Transitions:

	To													
From	Events:	Risk time:	Persons:											
1	99	878.97	115											
2	94	744.95	100											
3	60	258.27	63											
4	35	67.48	35											
5	38	110.31	39											
6	19	357.61	36											
Sum	345	2417.60	160											

Note that what differs between the data sets is the transitions and the relative distribution of the risk time between states; the total risk time is fixed for each person. The two datasets differ in transitions because *improvement* (decreasing state number) is allowed in the second.

5.2.3 Reversible retinopathy

As there are 40 observed transitions to *better* status we shall use the extended definition of transition that allows this too, except that transition from “Photocoagulation” to “Proliferative” is not allowed.

Now we set the seed explicitly so that results are reproducible

```
> set.seed( 4667571 )
> ret <- make.dlex( rt, rd, incr=FALSE )
> summary( ret )
```

Transitions:

	To													
From	1	2	3	4	5	6	Dead(1)	Dead(2)	Dead(3)	Dead(4)	Dead(5)	Dead(6)	Records:	
1	175	76	0	0	0	0	23	0	0	0	0	0	274	
2	11	165	54	0	0	0	0	29	0	0	0	0	259	
3	0	13	54	31	0	0	0	0	16	0	0	0	114	
4	0	0	1	7	33	0	0	0	0	1	0	0	42	

5	0	0	0	0	18	32	0	0	0	0	6	0	56
6	0	0	0	0	1	72	0	0	0	0	0	18	91
Sum	186	254	109	38	52	104	23	29	16	1	6	18	836

Transitions:

From	To	Events:	Risk time:	Persons:
1		99	873.08	115
2		94	757.05	100
3		60	250.40	63
4		35	56.00	35
5		38	110.47	39
6		19	370.59	36
Sum		345	2417.60	160

> levels(ret)

```
[1] "1"      "2"      "3"      "4"      "5"      "6"      "Dead(1)" "Dead(2)"
[9] "Dead(3)" "Dead(4)" "Dead(5)" "Dead(6)"
```

> levels(mic\$eds_1)

```
[1] "No retinopathy"          "Minimal non-proliferative" "Moderate nonproliferative"
[4] "Pre-proliferative"      "Proliferative"            "Photocoagulation"
```

> newl <- c("None", "Min", "Mod", "PrePr", "Pro", "PhC")

> levels(ret\$lex.Cst) <-

+ levels(ret\$lex.Xst) <- c(newl, paste("D(", newl, ")", sep=""))

> levels(ret)

```
[1] "None"      "Min"      "Mod"      "PrePr"    "Pro"      "PhC"      "D(None)"
[8] "D(Min)"    "D(Mod)"   "D(PrePr)" "D(Pro)"   "D(PhC)"
```

There is one illegal transition in the observed data; namely from photocoagulation to proliferative:

> subset(ret, lex.Cst=="PhC" & lex.Xst=="Pro")

lex.id	per	lex.dur	lex.Cst	lex.Xst	sex	doBth	doVis	doBlind	doEnd	
598	120	1997.748	0.9464413	PhC	Pro	Male	1939.29	1993.775	NA	2005.179
	doDth	allocation	age	tfe						
598	2005.179	Conventional	58.45859	3.972621						

> subset(ret, lex.id==120)

lex.id	per	lex.dur	lex.Cst	lex.Xst	sex	doBth	doVis	doBlind	doEnd	
600	120	1993.775	0.8496191	Pro	PhC	Male	1939.29	1993.775	NA	2005.179
597	120	1994.625	3.1230024	PhC	PhC	Male	1939.29	1993.775	NA	2005.179
598	120	1997.748	0.9464413	PhC	Pro	Male	1939.29	1993.775	NA	2005.179
599	120	1998.695	3.0042773	Pro	Pro	Male	1939.29	1993.775	NA	2005.179
601	120	2001.699	3.4798084	Pro	D(Pro)	Male	1939.29	1993.775	NA	2005.179
	doDth	allocation	age	tfe						
600	2005.179	Conventional	54.48597	0.0000000						
597	2005.179	Conventional	55.33559	0.8496191						
598	2005.179	Conventional	58.45859	3.9726215						
599	2005.179	Conventional	59.40503	4.9190628						
601	2005.179	Conventional	62.40931	7.9233402						

> ret[ret\$lex.id==120 & ret\$per>1995,"lex.Cst"] <- "PhC"

> ret[ret\$lex.id==120,"lex.Xst"] <- gsub("Pro","PhC",ret[ret\$lex.id==120,"lex.Xst"])

> subset(ret, lex.id==120)

lex.id	per	lex.dur	lex.Cst	lex.Xst	sex	doBth	doVis	doBlind	doEnd	
600	120	1993.775	0.8496191	Pro	PhC	Male	1939.29	1993.775	NA	2005.179
597	120	1994.625	3.1230024	PhC	PhC	Male	1939.29	1993.775	NA	2005.179
598	120	1997.748	0.9464413	PhC	PhC	Male	1939.29	1993.775	NA	2005.179
599	120	1998.695	3.0042773	PhC	PhC	Male	1939.29	1993.775	NA	2005.179
601	120	2001.699	3.4798084	PhC	D(PhC)	Male	1939.29	1993.775	NA	2005.179
	doDth	allocation	age	tfe						
600	2005.179	Conventional	54.48597	0.0000000						
597	2005.179	Conventional	55.33559	0.8496191						
598	2005.179	Conventional	58.45859	3.9726215						
599	2005.179	Conventional	59.40503	4.9190628						
601	2005.179	Conventional	62.40931	7.9233402						

With this in place we can now show the transitions through retinopathy states in the two groups; first defining the colors to use for the different states:

```
> clr <- c("forestgreen",heat.colors(7)[5:1])
> clr <- c(clr,rgb(t(col2rgb(clr)*0.6+255*0.4),max=255))
> apos = rep(0.45,12)
> apos[c(3,6,9)] <- 0.25
> par( mfrow=c(1,2), oma=c(0,0,0,0) )
> for( lv in levels(ret$allocation) )
+   {
+ boxes( subset( ret, allocation==lv ),
+         boxpos=list(x=rep(c(20,80),each=6)+7-14*(lv=="Intensive"),
+                       y=rep(seq(90,5,,6),2)),
+         show.BE="noz", scale.R=100, pos.arr=apos,
+         col.bg=clr,
+         col.border=c(clr[1:6],rep("black",6)),
+         col.txt=rep(c("white","black"),each=6) )
+ text(10+5-10*(lv=="Intensive"),98,lv,cex=1.5,font=2,adj=0)
+ apos[9] <- apos[1]
+   }
```

From figure 5.2 it is clear that the risk time in the states “Mod”, “PrePr” and “Pro” is quite small, indicating that persons travel quite quickly through these, on to “PhC” — apparent from the transition rates.

5.2.3.1 Analysis of rates

We now analyze transition rates between the two groups; what we do is to assume different transition intensities between levels of retinopathy, and fit models where intervention effect is:

- different between levels
- changing linearly across levels — separately for up/down transitions; only relevant and sensible for mortality rates
- is constant across levels

We fit three different sets of models; one set for increasing retinopathy status (deterioration) and one for decreasing (improvement), and a set of models for mortality:

In order to get readable code we use the stacked follow-up data for analysis. We create subsets of the stacked data for analysis of up-, resp. down- transitions, and shrink the transition levels to those actually present in the subset:

```
> stret <- stack( ret )
> ( trs <- levels( stret$lex.Tr ) )
[1] "None->Min"      "None->D(None)"  "Min->None"      "Min->Mod"
[5] "Min->D(Min)"    "Mod->Min"       "Mod->PrePr"     "Mod->D(Mod)"
[9] "PrePr->Mod"     "PrePr->Pro"     "PrePr->D(PrePr)" "Pro->PhC"
[13] "Pro->D(Pro)"    "PhC->D(PhC)"

> ( uptr <- trs[c(1,4,7,10,12)] )
[1] "None->Min"  "Min->Mod"  "Mod->PrePr" "PrePr->Pro" "Pro->PhC"

> ( dntr <- trs[c(9,6,3)] )
[1] "PrePr->Mod" "Mod->Min"  "Min->None"

> ( ddtr <- trs[grep("D",trs)] )
[1] "None->D(None)"  "Min->D(Min)"  "Mod->D(Mod)"  "PrePr->D(PrePr)"
[5] "Pro->D(Pro)"    "PhC->D(PhC)"
```

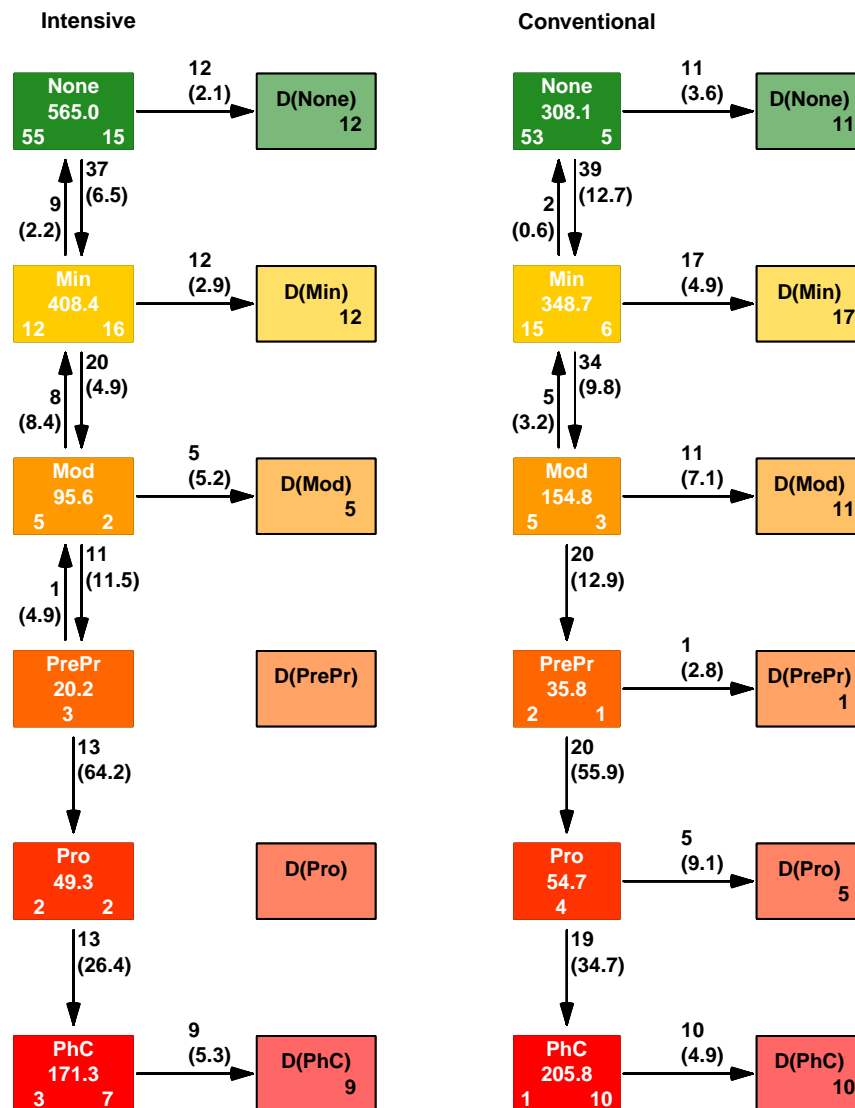



Figure 5.2: Transitions between states of retinopathy and death, allowing improvements in retinopathy status (except from Photocoagulation state). The number in the center of the boxes is the person-years, the numbers at the bottom is the number of patients starting, resp. ending their follow-up in each box, and the numbers on the arrows are the number of transitions and the overall transition rates per 100 PY.

```
> upst <- subset( stret, lex.Tr %in% uptr )
> dnst <- subset( stret, lex.Tr %in% dntr )
> ddst <- subset( stret, lex.Tr %in% ddtr )
> upst$lex.Tr <- factor( upst$lex.Tr )
> dnst$lex.Tr <- factor( dnst$lex.Tr )
```

```
> up.sep <- coxph( Surv( tfe, tfe+lex.dur, lex.Fail ) ~
+                 strata(lex.Cst) + allocation:lex.Tr,
+                 data = upst )
> up.con <- update( up.sep, . ~ . - allocation:lex.Tr + allocation )
```

```
> dn.sep <- update( up.sep, data = dnst )
> dn.con <- update( dn.sep, . ~ . - allocation:lex.Tr + allocation )
> dd.sep <- update( up.sep, data = ddst )
> dd.lin <- update( dd.sep, . ~ . - allocation:lex.Tr + allocation + allocation:as.integer(lex.Cst)
> dd.con <- update( dd.sep, . ~ . - allocation:lex.Tr + allocation )
```

For the purpose of prediction it is a bit more convenient to fit the models to the original Lexis object. We need two help-variables, namely the indicators of increase, resp. decrease in retinopathy score:

```
> ( llev <- levels(ret)[1:6] )
[1] "None" "Min" "Mod" "PrePr" "Pro" "PhC"
> ( dlev <- levels(ret)[7:12] )
[1] "D(None)" "D(Min)" "D(Mod)" "D(PrePr)" "D(Pro)" "D(PhC)"
> ret <- transform( ret, tr.up = as.integer(lex.Cst)<as.integer(lex.Xst) &
+                               lex.Xst %in% llev,
+                               tr.dn = as.integer(lex.Cst)>as.integer(lex.Xst) &
+                               lex.Xst %in% llev )
> ftable( xtabs( cbind(tr.up, tr.dn) ~ lex.Cst + lex.Xst, data=ret )
+         [1:6,1:6,], row.vars=c(3,1) )
      lex.Xst None Min Mod PrePr Pro PhC
tr.up lex.Cst
None      0  76  0  0  0  0
Min       0  0 54  0  0  0
Mod       0  0  0 31  0  0
PrePr    0  0  0  0 33  0
Pro      0  0  0  0  0 32
PhC      0  0  0  0  0  0
tr.dn lex.Cst
None      0  0  0  0  0  0
Min      11  0  0  0  0  0
Mod       0 13  0  0  0  0
PrePr    0  0  1  0  0  0
Pro      0  0  0  0  0  0
PhC      0  0  0  0  0  0
> u.sep <- coxph( Surv( tfe, tfe+lex.dur, tr.up ) ~
+               strata(lex.Cst) + lex.Cst:allocation,
+               data = subset(ret,lex.Cst %in% llev[1:5]) )
> u.com <- update( u.sep, . ~ . - lex.Cst:allocation + allocation )
> d.sep <- coxph( Surv( tfe, tfe+lex.dur, tr.dn ) ~
+               strata(lex.Cst) + lex.Cst:allocation,
+               data = subset(ret,lex.Cst %in% llev[2:6]) )
> d.com <- update( d.sep, . ~ . - lex.Cst:allocation + allocation )
> D.sep <- coxph( Surv( tfe, tfe+lex.dur, lex.Xst %in% dlev ) ~
+               strata(lex.Cst) + lex.Cst:allocation,
+               data = ret )
> D.lin <- update( D.sep, . ~ . - lex.Cst:allocation + allocation + allocation:as.integer(lex.Cst) )
> D.com <- update( D.sep, . ~ . - lex.Cst:allocation + allocation )
```

We can check that we get the same by the two approaches:

```
> round( cbind( ci.exp(u.sep), ci.exp(up.sep) ), 3 )
      exp(Est.) 2.5% 97.5% exp(Est.) 2.5% 97.5%
lex.CstNone:allocationIntensive 0.661 0.420 1.041 0.661 0.420 1.041
lex.CstMin:allocationIntensive 0.732 0.420 1.276 0.732 0.420 1.276
lex.CstMod:allocationIntensive 0.888 0.421 1.874 0.888 0.421 1.874
lex.CstPrePr:allocationIntensive 0.747 0.348 1.604 0.747 0.348 1.604
lex.CstPro:allocationIntensive 0.847 0.398 1.804 0.847 0.398 1.804
> round( cbind( ci.exp(d.sep), ci.exp(dn.sep) ), 3 )
      exp(Est.) 2.5% 97.5% exp(Est.) 2.5% 97.5%
lex.CstMin:allocationIntensive 5.178 1.109 24.168 5.178 1.109 24.168
lex.CstMod:allocationIntensive 3.085 0.980 9.714 3.085 0.980 9.714
lex.CstPrePr:allocationIntensive 29588452.321 0.000 Inf 29588452.300 0.000 Inf
```

```
> round( cbind( ci.exp(D.sep), ci.exp(dd.sep) ), 3 )
```

	exp(Est.)	2.5%	97.5%	exp(Est.)	2.5%	97.5%
lex.CstNone:allocationIntensive	0.474	0.206	1.088	0.474	0.206	1.088
lex.CstMin:allocationIntensive	0.480	0.224	1.029	0.480	0.224	1.029
lex.CstMod:allocationIntensive	0.768	0.264	2.233	0.768	0.264	2.233
lex.CstPro:allocationIntensive	0.000	0.000	Inf	0.000	0.000	Inf
lex.CstPhC:allocationIntensive	1.133	0.459	2.798	1.133	0.459	2.798

We use a Wald test to check whether the effect of intervention is the same across categories:

```
> ( CC <- t(matrix( c(1,-1,rep(0,5)), 6, 5 ) ) )
```

	[,1]	[,2]	[,3]	[,4]	[,5]	[,6]
[1,]	1	-1	0	0	0	0
[2,]	0	1	-1	0	0	0
[3,]	0	0	1	-1	0	0
[4,]	0	0	0	1	-1	0
[5,]	0	0	0	0	1	-1

```
> Wald( u.sep, ctr.mat=CC[1:4,1:5] )
```

Chisq	d.f.	P
0.5889057	4.0000000	0.9642885

```
> Wald( d.sep, ctr.mat=CC[1:2,1:3] )
```

Chisq	d.f.	P
0.2792115	2.0000000	0.8697010

```
> Wald( D.sep, ctr.mat=CC[1:4,1:5] )
```

Chisq	d.f.	P
2.6885343	4.0000000	0.6112224

```
> round( ci.exp(D.lin,pval=TRUE), 3 )
```

	exp(Est.)	2.5%	97.5%	P
allocationConventional	2.491	1.132	5.480	0.023
allocationIntensive:as.integer(lex.Cst)	1.131	0.901	1.421	0.289

We see that there is no evidence that the HR between intensive and conventional differ across the states for any of the three types of transitions; thus we assume that it is constant across retinopathy states. Thus based on the Cox-models we can report the three HRs associated with being in the intensive group:

```
> HR <- rbind( ci.exp(u.com,pval=TRUE),
+             ci.exp(d.com,pval=TRUE),
+             ci.exp(D.com,pval=TRUE) )
> rownames( HR ) <- c("Increasing ret","Decreasing ret","Death")
> HR <- cbind( 1/HR[,c(1,3,2)], HR[,c(4,1:3)] )
> colnames(HR)[c(1,5)] <- c("Int vs Conv","Conv vs Int")
> round( HR, 3 )
```

	Int vs Conv	97.5%	2.5%	P	Conv vs Int	2.5%	97.5%
Increasing ret	0.739	0.561	0.973	0.031	1.353	1.028	1.781
Decreasing ret	3.932	1.607	9.620	0.003	0.254	0.104	0.622
Death	0.572	0.370	0.883	0.012	1.748	1.132	2.701

From the Cox-models we see that there is almost a 25% smaller HR of increasing retinopathy status in the intensive group (HR 0.76; 0.58–1.01), a significantly *larger* HR of improving the retinopathy status (HR 3.4; 1.4–8.2) and a significantly lower mortality when controlling for retinopathy status (HR 0.49; 0.31–0.79), the latter pretty much confirming the overall analyses of mortality.

5.2.3.2 Parametric modeling

In order to get a more credible model for the retinopathy changes and the mortality, we fit a corresponding set of Poisson models where we also take age and sex into account. To this end we split the follow-up data in 1-month intervals:

```
> timeScales( ret )
[1] "per" "age" "tfe"
> sret <- splitLexis( ret, seq(0,24,1/12), "tfe" )
> summary( sret )
```

Transitions:

From	To	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)
None	None	10590	76	0	0	0	0	23	0	0	0	0	0
None	Min	11	9240	54	0	0	0	0	29	0	0	0	0
None	Mod	0	13	3057	31	0	0	0	0	16	0	0	0
None	PrePr	0	0	1	679	33	0	0	0	0	1	0	0
None	Pro	0	0	0	0	1256	32	0	0	0	0	5	0
None	PhC	0	0	0	0	0	4597	0	0	0	0	0	19
Sum		10601	9329	3112	710	1289	4629	23	29	16	1	5	19

```
Transitions:
To
From  Records:  Events:  Risk time:  Persons:
None  10689      99      873.08     115
Min   9334       94      757.05     100
Mod   3117       60      250.40     63
PrePr 714        35       56.00     35
Pro   1293       37      103.99     39
PhC   4616       19      377.07     36
Sum   29763     344     2417.60    160
```

In order to get the transitions right we must redefine the up and down transitions by the newly formed split records of follow-up:

```
> sret <- transform( sret, tr.up = as.integer(lex.Cst)<as.integer(lex.Xst) &
+                               lex.Xst %in% llev,
+                               tr.dn = as.integer(lex.Cst)>as.integer(lex.Xst) &
+                               lex.Xst %in% llev )
> ftable( addmargins( xtabs( cbind(up=tr.up,dn=tr.dn) ~ lex.Cst + allocation,
+                               data=sret )[1:6,,],
+                               margin=1:2 ),
+                               col.vars=3:2 )
```

lex.Cst	up			dn			
	allocation	Intensive	Conventional	Sum	Intensive	Conventional	Sum
None		37	39	76	0	0	0
Min		20	34	54	9	2	11
Mod		11	20	31	8	5	13
PrePr		13	20	33	1	0	1
Pro		13	19	32	0	0	0
PhC		0	0	0	0	0	0
Sum		94	132	226	18	7	25

We note that the number of improvements (dn) is quite thin; only 25 transitions recorded, of which a mere 7 in the conventional group.

Then we can fit models for each of the transitions, first we determine the position of the knots for the underlying splines:

```
> with( subset( sret, tr.up ), quantile( tfe+lex.dur, probs=0:3/4 ) )
      0%      25%      50%      75%
0.04287674 2.02443977 4.84442565 7.60560364
```

```
> with( subset( sret, tr.dn ), quantile( tfe+lex.dur, probs=0:3/4 ) )
      0%      25%      50%      75%
0.1170675 1.4905300 5.0865665 9.8096788
> with( subset( sret, lex.Xst %in% dlev), quantile( tfe+lex.dur, probs=0:3/4 ) )
      0%      25%      50%      75%
1.637235  7.151266 10.193018 15.863107
```

Thus we use the same set of knots for the changes in retinopathy status:

```
> ( r.kn <- c( 0, (
+   with(subset(sret,tr.up), quantile( tfe+lex.dur, probs=1:3/4 )) +
+   with(subset(sret,tr.dn), quantile( tfe+lex.dur, probs=1:3/4 ))) / 2 ) )
      25%      50%      75%
0.000000 1.757485 4.965496 8.707641
> ( d.kn <- with( subset( sret, lex.Xst %in% dlev ),
+   quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
      12.5%      37.5%      62.5%      87.5%
4.960986  8.870637 12.206708 16.810404
```

We can now fit models for increase and decrease in retinopathy score as well as mortality to the split dataset, using a smooth underlying hazard. We simplify things by using only linear interactions between times and state (non-proportionality). We make a test for interaction between state and time (removing it from the model), and a test for interaction between state and allocation (adding it to the model):

```
> # Transitions to worse retinopathy
> up.i <- glm( tr.up ~ Ns( tfe, knots=r.kn ) + lex.Cst + lex.Cst:tfe + allocation + age + sex,
+   offset = log(lex.dur),
+   family = poisson,
+   data = subset(sret,lex.Cst %in% llev[1:5]) )
> up.0 <- update( up.i, . ~ . - lex.Cst:tfe )
> up.s <- update( up.i, . ~ . + lex.Cst:allocation )
> anova( up.0, up.i, up.s, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  4  22.162 0.000186
3  4   1.213 0.875953
> # Transitions to better retinopathy status --- we leave out the
> # interaction for want of data
> dn.0 <- glm( tr.dn ~ Ns( tfe, knots=r.kn ) + lex.Cst + allocation + age + sex,
+   offset = log(lex.dur),
+   family = poisson,
+   data = subset(sret,lex.Cst %in% llev[2:6]) )
> dn.s <- update( dn.0, . ~ . + lex.Cst:allocation )
> anova( dn.0, dn.s, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  4   1.2079  0.8768
> # Transitions to death
> mt.i <- update( up.i, (lex.Xst %in% levels(lex.Xst)[7:12]) ~ .,
+   data = sret )
> mt.0 <- update( mt.i, . ~ . - lex.Cst:tfe )
> mt.s <- update( mt.i, . ~ . + lex.Cst:allocation )
> anova( mt.0, mt.i, mt.s, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  5   0.2236  0.99884
3  5   9.5370  0.08947
```

For the transitions to worse retinopathy, we see that there is an interaction between state and time, but none for mortality. For the transitions to better retinopathy status we were not able to fit a model with state by time interaction because of the rather thin data, and we saw no indication of interaction between state and allocation (also because of the rather thin data). Thus the relevant model for the deterioration is `up.i`, for improvement `dn.0`, and for mortality `mt.0`.

```

> round( ci.exp(up.i,pval=T), 3 )
              exp(Est.)  2.5% 97.5%    P
(Intercept)      0.179 0.053 0.601 0.005
Ns(tfe, knots = r.kn)1  2.965 1.536 5.725 0.001
Ns(tfe, knots = r.kn)2  1.857 0.453 7.607 0.390
Ns(tfe, knots = r.kn)3  1.780 0.843 3.758 0.130
lex.CstMin        0.726 0.388 1.356 0.315
lex.CstMod        0.793 0.339 1.854 0.592
lex.CstPrePr      1.759 0.808 3.829 0.155
lex.CstPro        1.200 0.528 2.724 0.663
allocationConventional 1.311 0.999 1.720 0.051
age               0.998 0.979 1.018 0.839
sexMale           1.112 0.819 1.510 0.498
lex.CstNone:tfe   0.809 0.715 0.915 0.001
lex.CstMin:tfe    0.856 0.758 0.967 0.012
lex.CstMod:tfe    0.912 0.794 1.048 0.193
lex.CstPrePr:tfe  1.027 0.910 1.158 0.668
lex.CstPro:tfe    1.000 1.000 1.000  NaN

> round( ci.exp(dn.0,pval=T), 3 )
              exp(Est.)  2.5% 97.5%    P
(Intercept)      0.087 0.002 3.461 0.194
Ns(tfe, knots = r.kn)1  0.417 0.123 1.416 0.161
Ns(tfe, knots = r.kn)2  0.023 0.001 0.377 0.008
Ns(tfe, knots = r.kn)3  0.446 0.146 1.364 0.157
lex.CstMod        3.780 1.667 8.571 0.001
lex.CstPrePr      0.985 0.125 7.779 0.989
lex.CstPro        0.000 0.000  Inf 0.993
lex.CstPhC        0.000 0.000  Inf 0.987
allocationConventional 0.294 0.122 0.712 0.007
age               1.008 0.950 1.070 0.787
sexMale           1.175 0.392 3.522 0.773

> round( ci.exp(mt.0,pval=T), 3 )
              exp(Est.)  2.5%    97.5%    P
(Intercept)      0.000 0.000    0.000 0.000
Ns(tfe, knots = r.kn)1  10.529 1.013    109.485 0.049
Ns(tfe, knots = r.kn)2  108.997 0.091 130456.335 0.195
Ns(tfe, knots = r.kn)3   4.689 1.070    20.555 0.040
lex.CstMin        0.880 0.500    1.547 0.657
lex.CstMod        2.203 1.133    4.287 0.020
lex.CstPrePr      0.738 0.098    5.540 0.768
lex.CstPro        2.099 0.783    5.628 0.141
lex.CstPhC        1.279 0.678    2.415 0.447
allocationConventional 1.774 1.149    2.740 0.010
age               1.125 1.085    1.167 0.000
sexMale           1.497 0.906    2.471 0.115

```

From these we extract the RRs associated with allocation:

```

> HRests <- rbind( ci.exp( up.i, subset="allo", pval=T ),
+                 ci.exp( dn.0, subset="allo", pval=T ),
+                 ci.exp( mt.0, subset="allo", pval=T ) )
> HRests <- cbind( 1/HRests[,c(1,3,2)], HRests )
> rownames( HRests ) <- c("Worse","Better","Death")
> colnames( HRests )[c(1,4)] <- c("Int vs Conv","Conv vs Int")
> round( HRests, 3 )
              Int vs Conv 97.5%  2.5% Conv vs Int  2.5% 97.5%    P
Worse         0.763 0.581 1.001    1.311 0.999 1.720 0.051
Better        3.400 1.405 8.229    0.294 0.122 0.712 0.007
Death         0.564 0.365 0.870    1.774 1.149 2.740 0.010

```

There is a 21% smaller rate of deterioration and a 3 times bigger rate of improvement in retinopathy in the intensive group; the improvement rates very uncertain, though. Controlled for retinopathy state the HR of death is 47% smaller in the intensive group.

5.2.3.3 Progression probabilities

With these models in place we can now set up a transition object; note that for retinopathy progression we use a model with a state by time interaction, and for improvement of retinopathy and mortality we use a proportional hazards model, *i.e.* a model where the intervention has the same effect in all states and in all times:

```
> Tr <- list( "None" = list( "Min" = up.i,
+                          "D(None)" = mt.0 ),
+           "Min" = list( "None" = dn.0,
+                        "Mod" = up.i,
+                        "D(Min)" = mt.0 ),
+           "Mod" = list( "Min" = dn.0,
+                        "PrePr" = up.i,
+                        "D(Mod)" = mt.0 ),
+           "PrePr" = list( "Mod" = dn.0,
+                          "Pro" = up.i,
+                          "D(PrePr)" = mt.0 ),
+           "Pro" = list( "PrePr" = dn.0,
+                        "PhC" = up.i,
+                        "D(Pro)" = mt.0 ),
+           "PhC" = list( "D(PhC)" = mt.0 ) )
> lapply(Tr,names)
$None
[1] "Min"      "D(None)"

$Min
[1] "None"    "Mod"     "D(Min)"

$Mod
[1] "Min"     "PrePr"   "D(Mod)"

$PrePr
[1] "Mod"     "Pro"     "D(PrePr)"

$Pro
[1] "PrePr"   "PhC"     "D(Pro)"

$PhC
[1] "D(PhC)"
```

In order to assess how retinopathy scores have developed in the two groups we simulate how retinopathy score and death is predicted to develop for different types of patient populations; in all case we simulate in two scenarios: Intensive and Conventional.

- A cohort that has the same baseline (joint) distribution of retinopathy scores, age and sex as the Steno 2 patients.
- A cohort where all persons have a retinopathy score of “None”, and the same age and sex distribution as the Steno 2 patients.
- Cohorts where all have retinopathy score “None”, and the same baseline age (45,50,55,60,65) and sex. This will be simulated using just one cohort with the 10 different combinations represented.

This is basically accomplished by setting up different initial cohorts; `ini.e` corresponding to the empirical distribution of score age and sex, `ini.0` with everyone having a score of 0 but the same age-sex-distribution, and finally `ini.as` with fixed values of age and sex and retinopathy score.

In order to get a Lexis object that corresponds to the baseline state we extract the earliest record for each person; the calendar time will then be the date of baseline, and age the age at baseline:

```
> ini.e <- subset( ret, per==ave(per,lex.id,FUN=min),
+               select = c("lex.id","lex.Cst",timeScales(ret),"allocation","sex") )
> ini.e <- rbind( transform( ini.e, allocation=factor(levels(allocation)[1]) ),
+               transform( ini.e, allocation=factor(levels(allocation)[2]) ) )
> str( ini.e )
Classes 'Lexis' and 'data.frame':      320 obs. of  7 variables:
 $ lex.id      : num  1 2 3 4 5 6 7 8 9 10 ...
 $ lex.Cst     : Factor w/ 12 levels "None","Min","Mod",...: 1 2 2 1 1 1 1 2 2 1 ...
 $ per        : num  1993 1993 1993 1993 1993 ...
 $ age        : num  61.1 46.5 50 48.6 57.2 ...
 $ tfe        : num  0 0 0 0 0 0 0 0 0 0 ...
 $ allocation: Factor w/ 2 levels "Intensive","Conventional": 1 1 1 1 1 1 1 1 1 1 ...
 $ sex        : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 1 2 2 2 ...
 - attr(*, "breaks")=List of 3
 ..$ per: NULL
 ..$ age: NULL
 ..$ tfe: NULL
 - attr(*, "time.scales")= chr  "per" "age" "tfe"
 - attr(*, "time.since")= chr  "" "" ""

> with( ini.e, table( allocation, lex.Cst ) )
           lex.Cst
allocation  None Min Mod PrePr Pro PhC D(None) D(Min) D(Mod) D(PrePr) D(Pro) D(PhC)
Intensive   108 27 10   5  6  4         0     0     0         0     0     0
Conventional 108 27 10   5  6  4         0     0     0         0     0     0

> ini.0 <- transform( ini.e, lex.Cst=factor( "None", levels=levels(lex.Cst) ) )
> str( ini.0 )
Classes 'Lexis' and 'data.frame':      320 obs. of  7 variables:
 $ lex.id      : num  1 2 3 4 5 6 7 8 9 10 ...
 $ lex.Cst     : Factor w/ 12 levels "None","Min","Mod",...: 1 1 1 1 1 1 1 1 1 1 ...
 $ per        : num  1993 1993 1993 1993 1993 ...
 $ age        : num  61.1 46.5 50 48.6 57.2 ...
 $ tfe        : num  0 0 0 0 0 0 0 0 0 0 ...
 $ allocation: Factor w/ 2 levels "Intensive","Conventional": 1 1 1 1 1 1 1 1 1 1 ...
 $ sex        : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 1 2 2 2 ...
 - attr(*, "breaks")=List of 3
 ..$ per: NULL
 ..$ age: NULL
 ..$ tfe: NULL
 - attr(*, "time.scales")= chr  "per" "age" "tfe"
 - attr(*, "time.since")= chr  "" "" ""

> with( ini.0, table( allocation, lex.Cst ) )
           lex.Cst
allocation  None Min Mod PrePr Pro PhC D(None) D(Min) D(Mod) D(PrePr) D(Pro) D(PhC)
Intensive   160  0  0   0  0  0         0     0     0         0     0     0
Conventional 160  0  0   0  0  0         0     0     0         0     0     0
```

With the two initiation cohorts we can now simulate the predicted outcomes according to the model for the transitions:

```
> sim.e <- simLexis( Tr=Tr, init=ini.e, time.pts=seq(0,22,0.2), N=100 )
> sim.0 <- simLexis( Tr=Tr, init=ini.0, time.pts=seq(0,22,0.2), N=100 )
> save( sim.e, sim.0, file="../data/simRet.Rda" )

> load( file="../data/simRet.Rda" )
> summary( sim.e )
```


Transitions:

From	To											
	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)
None	3670	15294	0	0	0	0	5123	0	0	0	0	0
Min	2487	4495	11013	0	0	0	0	5357	0	0	0	0
Mod	0	2658	905	5856	0	0	0	0	3788	0	0	0
PrePr	0	0	194	86	6339	0	0	0	0	237	0	0
Pro	0	0	0	0	295	6085	0	0	0	0	1159	0
PhC	0	0	0	0	0	3019	0	0	0	0	0	3866
Sum	6157	22447	12112	5942	6634	9104	5123	5357	3788	237	1159	3866

Transitions:

From	To			
	Records:	Events:	Risk time:	Persons:
None	24087	20417	174119.59	22618
Min	23352	18857	165896.96	20629
Mod	13207	12302	52634.28	12213
PrePr	6856	6770	10922.26	6772
Pro	7539	7244	19377.49	7539
PhC	6885	3866	74531.31	6885
Sum	81926	69456	497481.89	32000

> summary(sim.0)

Transitions:

From	To											
	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)
None	5110	21972	0	0	0	0	7041	0	0	0	0	0
Min	2123	5214	10445	0	0	0	0	5940	0	0	0	0
Mod	0	1750	896	4376	0	0	0	0	3508	0	0	0
PrePr	0	0	85	71	4033	0	0	0	0	187	0	0
Pro	0	0	0	0	278	2931	0	0	0	0	824	0
PhC	0	0	0	0	0	1478	0	0	0	0	0	1453
Sum	7233	28936	11426	4447	4311	4409	7041	5940	3508	187	824	1453

Transitions:

From	To			
	Records:	Events:	Risk time:	Persons:
None	34123	29013	248461.79	32000
Min	23722	18508	172455.13	21444
Mod	10530	9634	42326.30	10033
PrePr	4376	4305	6105.24	4346
Pro	4033	3755	10132.37	4033
PhC	2931	1453	26116.25	2931
Sum	79715	66668	505597.08	32000

5.2.3.4 Prediction from baseline retinopathy distribution

Here we show how a population of patients with an age- sex- and retinopathy-score distribution as the Steno 2 patients would fare through the states were they subjected to intensive, respectively conventional treatment.

```
> prm <- c(1:6,12:7)
> prm
[1] 1 2 3 4 5 6 12 11 10 9 8 7
> levels( sim.e )[prm]
[1] "None"      "Min"        "Mod"        "PrePr"      "Pro"        "PhC"        "D(PhC)"
[8] "D(Pro)"     "D(PrePr)"  "D(Mod)"     "D(Min)"     "D(None)"
> rEi <- pState( nState( subset(sim.e,allocation=="Intensive"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> rEc <- pState( nState( subset(sim.e,allocation=="Conventional"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> round( head( rEi ), 4 )
```

```

      State
when   None   Min   Mod  PrePr   Pro   PhC D(PhC) D(Pro) D(PrePr) D(Mod) D(Min)
0     0.6750 0.8438 0.9062 0.9375 0.9750 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000
0.25 0.6541 0.8471 0.9046 0.9349 0.9734 0.9996 0.9997 0.9997 0.9997 0.9998 0.9998
0.5  0.6352 0.8466 0.9034 0.9329 0.9710 0.9994 0.9994 0.9996 0.9996 0.9997 0.9998
0.75 0.6178 0.8440 0.9019 0.9304 0.9690 0.9990 0.9991 0.9992 0.9993 0.9994 0.9996
1     0.6000 0.8420 0.9001 0.9276 0.9664 0.9982 0.9983 0.9986 0.9986 0.9988 0.9991
1.25 0.5844 0.8378 0.8979 0.9238 0.9633 0.9972 0.9973 0.9976 0.9977 0.9981 0.9984
      State
when   D(None)
0           1
0.25       1
0.5         1
0.75       1
1           1
1.25       1

```

Then plot the estimated distributions over the

```

> retst <- function( Pi, Pc, rv=FALSE, leg=rep(15,12) )
+ {
+ par( mfrow=c(1,2), mar=c(0,3,0,0), oma=c(3,0,2,3-2*rv), las=1 )
+ plot( Pi, col=clr[prm], xlim=c(0,20), xlab="", # yaxt="n",
+       col.lab="transparent" )
+ lines( as.numeric(rownames(Pi)), Pi[, "PhC"], lwd=4 )
+ Pi <- cbind(0,Pi)
+ wr <- cbind( match(paste(leg),rownames(Pi)), 2:13 )
+ wl <- cbind( match(paste(leg),rownames(Pi)), 1:12 )
+ if(is.numeric(leg)) text( leg,
+                           (Pi[wl]+Pi[wr])/2,
+                           colnames(Pi)[-1],
+                           col=rep(c("white","black"),each=6) )
+ axis( side=4, at=0:5/5 )
+ axis( side=4, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Probability", side=2, line=1, las=0 )
+ mtext( "Intensive", side=3, line=1 )
+
+ plot( Pc, col=clr[prm], xlim=if(rv) c(20,0) else c(0,20),
+       xlab="", yaxt="n", col.lab="transparent" )
+ lines( as.numeric(rownames(Pc)), Pc[, "PhC"], lwd=4 )
+ axis( side=4-2*rv, at=1:5/5, if(rv) labels=NA )
+ axis( side=4-2*rv, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4-2*rv, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4-2*rv, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Conventional", side=3, line=1 )
+ mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
+ }
> retst(rEi,rEc,rv=FALSE,leg=c(6:8,5,10,12,18,19,rep(18,4)) )

```

We can also plot it reversely for the conventional group to facilitate comparison

```

> retst(rEi,rEc,rv=TRUE,leg=c(6:8,6,9,12,18,19,rep(18,4)) )

```

5.2.3.5 Prediction given absence of retinopathy

Here we show how a population of patients with an age- and sex-distribution as the Steno 2 patients all without retinopathy at baseline, would fare through the states were they subjected to intensive, respectively conventional treatment.

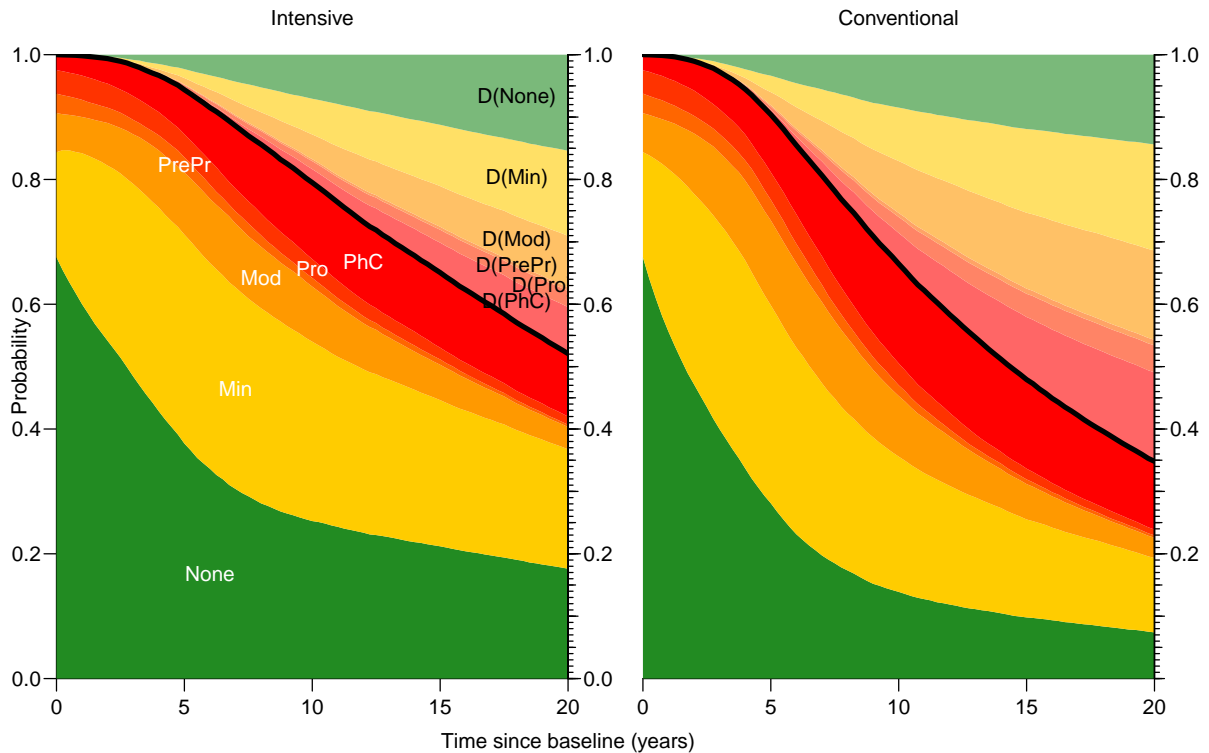


Figure 5.3: Patients with age- sex- and retinopathy score distribution as in Steno 2 study: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

```
> rZi <- pState( nState( subset(sim.0,allocation=="Intensive"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> rZc <- pState( nState( subset(sim.0,allocation=="Conventional"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> round( head( rZi ), 4 )
      State
when  None  Min  Mod  PrePr  Pro  PhC D(PhC) D(Pro) D(PrePr) D(Mod) D(Min)
0     1.0000 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000
0.25 0.9584 0.9996 0.9999 0.9999 0.9999 0.9999 0.9999 0.9999 0.9999 0.9999 0.9999
0.5   0.9219 0.9976 0.9996 0.9996 0.9996 0.9996 0.9996 0.9996 0.9996 0.9996 0.9996
0.75 0.8906 0.9956 0.9991 0.9991 0.9992 0.9992 0.9992 0.9992 0.9992 0.9992 0.9992
1     0.8602 0.9912 0.9983 0.9985 0.9986 0.9986 0.9986 0.9986 0.9986 0.9986 0.9988
1.25 0.8348 0.9885 0.9975 0.9978 0.9978 0.9979 0.9979 0.9979 0.9979 0.9979 0.9982
      State
when  D(None)
0     1
0.25 1
0.5   1
0.75 1
1     1
1.25 1
```

Then plot the estimated distributions over the

```
> retst(rZi,rZc,rv=FALSE,leg=c(6:8,6,9,12,18,19,rep(18,4)) )
```

We can also reverse the x -axis for the conventional group to facilitate comparison

```
> retst(rZi,rZc,rv=TRUE,leg=c(6:8,6,9,12,18,19,rep(18,4)) )
```

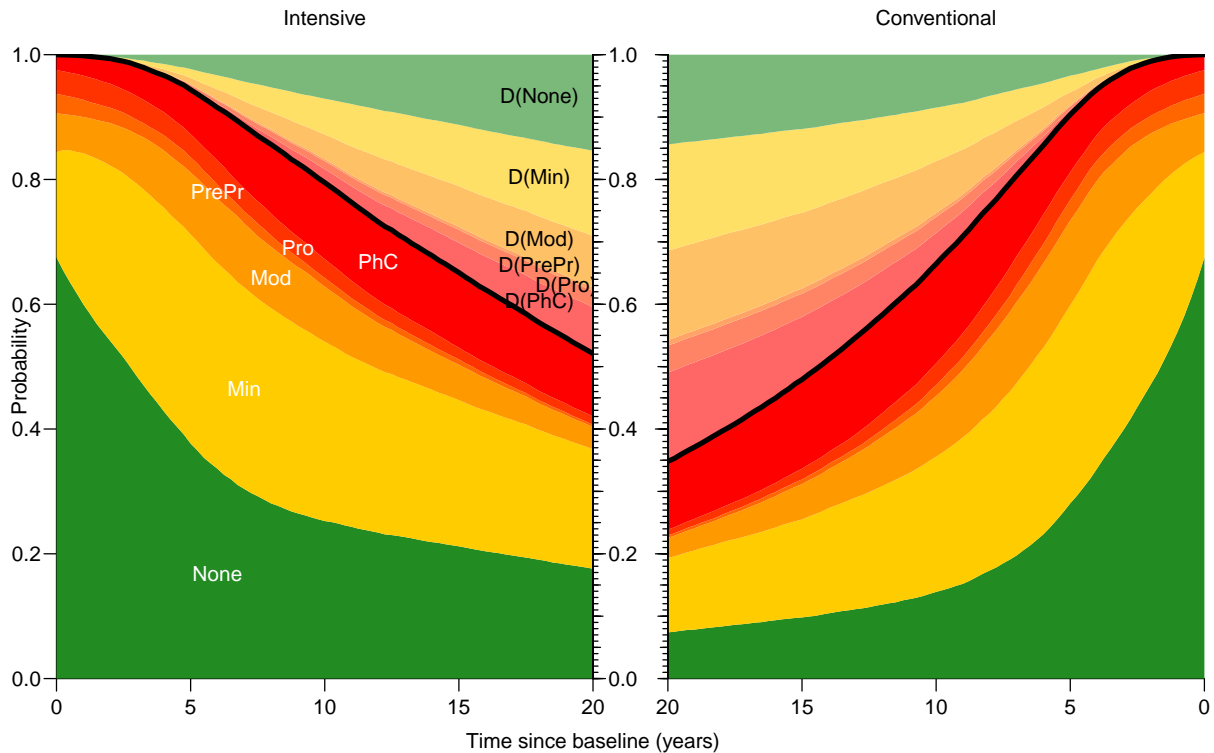


Figure 5.4: Patients with age- sex- and retinopathy score distribution as in Steno 2 study: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

This is the same figure as 5.3, except that the x-axis of the right-hand panel is reversed.

5.2.4 Non-reversible retinopathy

In figure 5.2 we see that the the number of transitions to a better state is quite limited, which means that including these very uncertain transitions in the predictions makes them quite uncertain, By this token we also make an analysis of data where we disallow improvement; that is redefining the states to “the worst state seen so far”.

We again set the random number generator seed explicitly so that results are reproducible

```
> set.seed( 4667571 )
> ret <- make.dlex( rt, rd, incr=TRUE )
> levels( ret )
[1] "1"      "2"      "3"      "4"      "5"      "6"      "Dead(1)" "Dead(2)"
[9] "Dead(3)" "Dead(4)" "Dead(5)" "Dead(6)"
> levels( mic$eds_1 )
[1] "No retinopathy"      "Minimal non-proliferative" "Moderate nonproliferative"
[4] "Pre-proliferative"  "Proliferative"           "Photocoagulation"
> new1 <- c("None", "Min", "Mod", "PrePr", "Pro", "PhC")
> levels( ret$lex.Cst ) <-
+ levels( ret$lex.Xst ) <- c( new1, paste("D(", new1, ")", sep="" ) )
> levels( ret )
[1] "None"      "Min"      "Mod"      "PrePr"    "Pro"      "PhC"      "D(None)"
[8] "D(Min)"    "D(Mod)"   "D(PrePr)" "D(Pro)"   "D(PhC)"
> summary( ret )
```

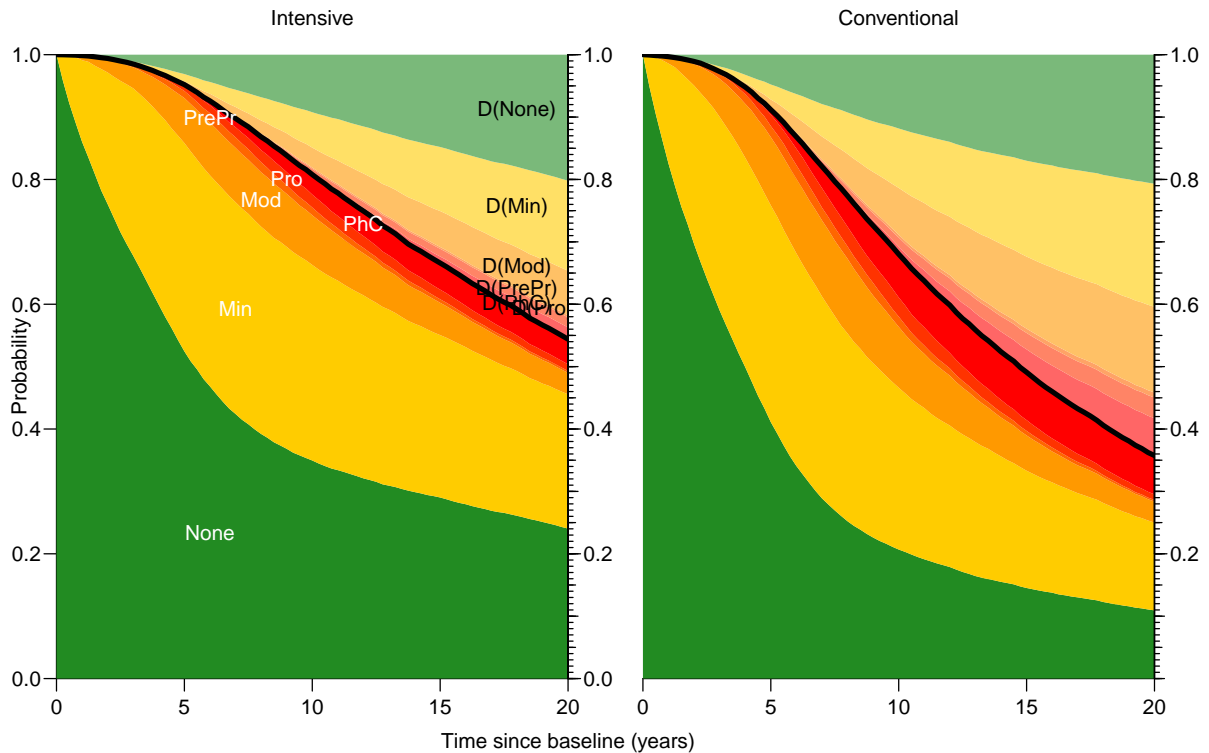


Figure 5.5: Patients all initiating with retinopathy score 0; age- and sex-distribution as in Steno 2 study population: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

Transitions:

From	To	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)	Records:
None		159	70	0	0	0	0	21	0	0	0	0	0	250
Min		0	156	52	0	0	0	0	27	0	0	0	0	235
Mod		0	0	78	30	0	0	0	0	20	0	0	0	128
PrePr		0	0	0	8	33	0	0	0	0	1	0	0	42
Pro		0	0	0	0	17	32	0	0	0	0	5	0	54
PhC		0	0	0	0	0	73	0	0	0	0	0	19	92
Sum		159	226	130	38	50	105	21	27	20	1	5	19	801

Transitions:

From	To	Events:	Risk time:	Persons:
None		91	806.73	108
Min		79	699.02	97
Mod		50	372.22	62
PrePr		34	72.65	35
Pro		37	97.61	39
PhC		19	369.36	36
Sum		310	2417.60	160

With this in place we can now show the transitions through retinopathy states in the two groups:

```
> par( mfrow=c(1,2), oma=c(0,0,0,0) )
> for( lv in levels(ret$allocation) )
+ {
+ boxes( subset( ret, allocation==lv ),
+         boxpos=list(x=rep(c(20,80),each=6)+7-14*(lv=="Intensive"),
```

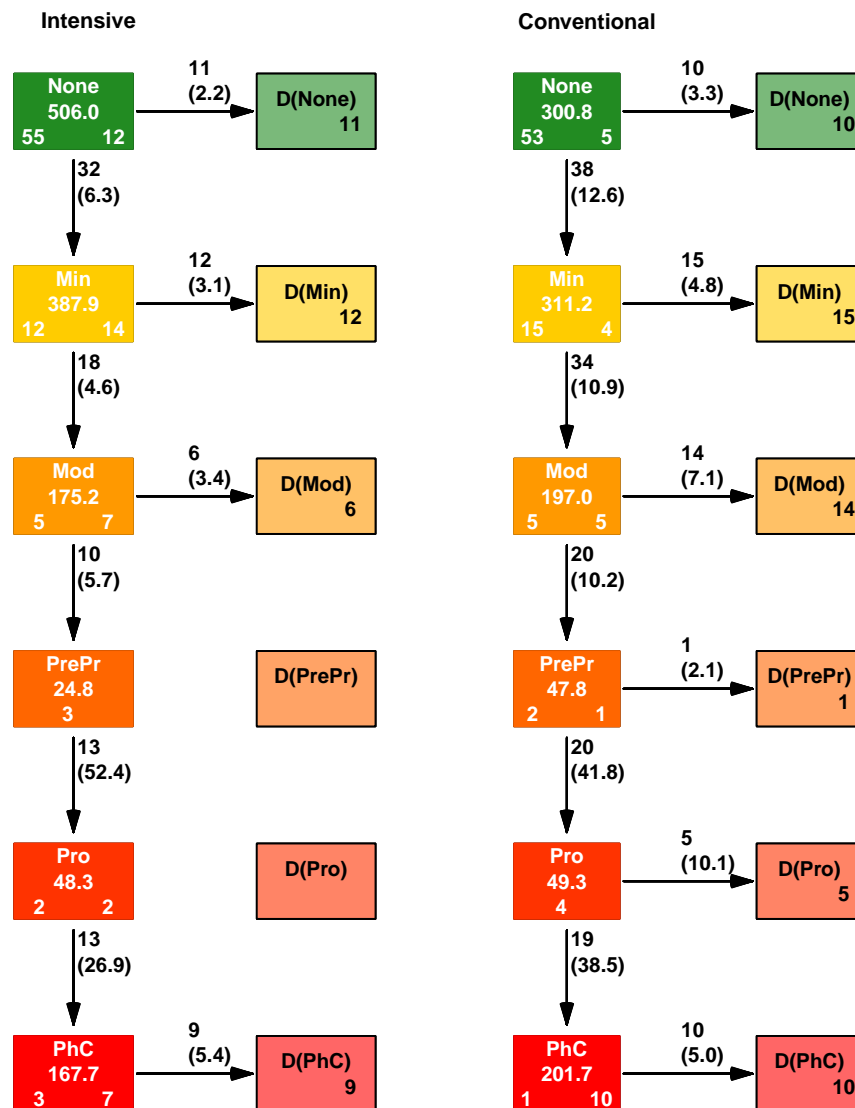



Figure 5.7: Transitions between states of retinopathy and death allowing only progression of retinopathy. The number in the center of the boxes is the person-years, the numbers at the bottom is the number of patients starting, resp. ending their follow-up in each box, and the numbers on the arrows are the number of transitions and the overall transition rates per 100 PY.

```

> u.sep <- coxph( Surv( tfe, tfe+lex.dur, tr.up ) ~
+                 strata(lex.Cst) + lex.Cst:allocation,
+                 data = subset(ret,lex.Cst %in% llev[1:5]) )
> u.com <- update( u.sep, . ~ . - lex.Cst:allocation + allocation )
> D.sep <- coxph( Surv( tfe, tfe+lex.dur, lex.Xst %in% dlev ) ~
+                 strata(lex.Cst) + lex.Cst:allocation,
+                 data = ret )
> D.lin <- update( D.sep, . ~ . - lex.Cst:allocation + allocation + allocation:as.integer(lex.Cst) )
> D.com <- update( D.sep, . ~ . - lex.Cst:allocation + allocation )

```

We can check the resulting estimates;

```
> round( ci.exp(u.sep), 3 )
                    exp(Est.)  2.5% 97.5%
lex.CstNone:allocationIntensive  0.634 0.395 1.019
lex.CstMin:allocationIntensive   0.641 0.360 1.141
lex.CstMod:allocationIntensive   0.691 0.321 1.486
lex.CstPrePr:allocationIntensive 1.098 0.477 2.527
lex.CstPro:allocationIntensive   1.060 0.516 2.179

> round( ci.exp(D.sep), 3 )
                    exp(Est.)  2.5% 97.5%
lex.CstNone:allocationIntensive  0.517 0.217 1.229
lex.CstMin:allocationIntensive   0.497 0.223 1.103
lex.CstMod:allocationIntensive   0.524 0.199 1.377
lex.CstPrePr:allocationIntensive 0.000 0.000  Inf
lex.CstPro:allocationIntensive   0.000 0.000  Inf
lex.CstPhC:allocationIntensive   1.128 0.457 2.786
```

We use a Wald test to check whether the effect of intervention is the same across categories:

```
> ( CC <- t(matrix( c(1,-1,rep(0,5)), 6, 5 ) ) )
      [,1] [,2] [,3] [,4] [,5] [,6]
[1,]    1   -1    0    0    0    0
[2,]    0    1   -1    0    0    0
[3,]    0    0    1   -1    0    0
[4,]    0    0    0    1   -1    0
[5,]    0    0    0    0    1   -1

> Wald( u.sep, ctr.mat=CC[1:4,1:5] )
      Chisq      d.f.      P
2.4888772  4.0000000  0.6466286

> Wald( D.sep, ctr.mat=CC[1:5,1:6] )
      Chisq      d.f.      P
2.2657049  5.0000000  0.8112905

> round( ci.exp(D.lin,pval=TRUE), 3 )
                    exp(Est.)  2.5% 97.5%      P
allocationConventional      2.469 1.097 5.559 0.029
allocationIntensive:as.integer(lex.Cst) 1.117 0.885 1.409 0.351
```

We see that there is no evidence that the HRs differ across the states, thus that the HR of increase between intensive and conventional is constant across retinopathy states, as is the mortality HR. Thus based on the Cox-models we can report the three HRs associated with being in the intensive group:

```
> HR <- rbind( ci.exp(u.com,pval=TRUE),
+             ci.exp(D.com,pval=TRUE) )
> rownames( HR ) <- c("Increasing ret","Death")
> HR <- cbind( 1/HR[,c(1,3,2)], HR )
> colnames(HR)[c(1,4)] <- c("Int vs Conv","Conv vs Int")
> round( HR, 3 )
      Int vs Conv 97.5% 2.5% Conv vs Int 2.5% 97.5%      P
Increasing ret   0.739 0.557 0.980      1.353 1.020 1.795 0.036
Death           0.560 0.363 0.864      1.785 1.158 2.751 0.009
```

Thus we see that there is a 26% smaller HR of increasing retinopathy status in the intensive group (HR 0.74; 0.56–0.99), and a lower mortality when controlling for retinopathy status (HR 0.50; 0.31–0.80), the latter again pretty much confirming the overall analyses of mortality.

5.2.4.1 Parametric modeling

In order to get a more credible model for the retinopathy changes and the mortality, we fit a corresponding set of Poisson models where we also take age and sex into account. To this end we split the follow-up data in 1-month intervals:

```
> timeScales( ret )
[1] "per" "age" "tfe"
> sret <- splitLexis( ret, seq(0,24,1/12), "tfe" )
> summary( sret )
```

Transitions:

From	To											
	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)
None	9782	70	0	0	0	0	21	0	0	0	0	0
Min	0	8533	52	0	0	0	0	27	0	0	0	0
Mod	0	0	4538	30	0	0	0	0	20	0	0	0
PrePr	0	0	0	879	33	0	0	0	0	1	0	0
Pro	0	0	0	0	1183	32	0	0	0	0	5	0
PhC	0	0	0	0	0	4503	0	0	0	0	0	19
Sum	9782	8603	4590	909	1216	4535	21	27	20	1	5	19

```
Transitions:
  To
From  Records:  Events:  Risk time:  Persons:
None      9873      91      806.73      108
Min       8612      79      699.02       97
Mod       4588      50      372.22       62
PrePr     913       34       72.65       35
Pro      1220       37       97.61       39
PhC      4522       19      369.36       36
Sum      29728     310     2417.60     160
```

In order to get the transitions right we must redefine the up transition indicator in the newly formed split records of follow-up:

```
> sret <- transform( sret, tr.up = as.integer(lex.Cst)<as.integer(lex.Xst) &
+                               lex.Xst %in% llev )
```

Then we can fit models for each of the transitions, first we determine the position of the knots for the underlying splines:

```
> with( subset( sret, tr.up ), quantile( tfe+lex.dur, probs=0:3/4 ) )
      0%      25%      50%      75%
0.04100879 2.05938196 5.12452553 7.70114052
> with( subset( sret, lex.Xst %in% dlev), quantile( tfe+lex.dur, probs=0:3/4 ) )
      0%      25%      50%      75%
1.637235 7.151266 10.193018 15.863107
```

Thus we use knots for the changes in retinopathy status where we have 0 as the first knot, and for mortality only the quantile knots:

```
> ( r.kn <- c( 0,
+   with(subset(sret,tr.up), quantile( tfe+lex.dur, probs=1:3/4 )) ) )
      25%      50%      75%
0.000000 2.059382 5.124526 7.701141
> ( d.kn <- with( subset( sret, lex.Xst %in% dlev ),
+   quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
      12.5%      37.5%      62.5%      87.5%
4.960986 8.870637 12.206708 16.810404
```

We can now fit models for increase and decrease in retinopathy score as well as mortality to the split dataset, using a smooth underlying hazard. We simplify things by using only linear interactions between times and state (non-proportionality). We make a test for interaction between states (removing it from the model), and a test for interaction between state and allocation effect (adding it to the model):

```
> # Transitions to worse retinopathy
> up.i <- glm( tr.up ~ Ns( tfe, knots=r.kn) + lex.Cst + lex.Cst:tfe + allocation + age + sex,
+           offset = log(lex.dur),
+           family = poisson,
+           data = subset(sret,lex.Cst %in% llev[1:5]) ) # exclude
+                                     # risk in the PhC state
> up.0 <- update( up.i, . ~ . - lex.Cst:tfe )
> up.s <- update( up.i, . ~ . + lex.Cst:allocation )
> anova( up.0, up.i, up.s, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  4  23.7004 9.172e-05
3  4   1.9143  0.7515
> # Transitions to death
> mt.i <- update( up.i, (lex.Xst %in% levels(lex.Xst)[7:12]) ~ .,
+               data = sret )
> mt.0 <- update( mt.i, . ~ . - lex.Cst:tfe )
> mt.s <- update( mt.i, . ~ . + lex.Cst:allocation )
> anova( mt.0, mt.i, mt.s, test="Chisq" )[-1,-(1:2)]
  Df Deviance Pr(>Chi)
2  5   1.0578 0.95777
3  5   9.5125 0.09029
```

For the transitions to worse retinopathy, we see that there is an interaction between state and time, but none between mortality state and allocation, while neither of these are present for mortality rates.

We then update the HR collector for the retinopathy

```
> up.z <- update( up.0, . ~ . - age - sex )
> zz <- rbind( ci.exp( up.0, subset="allo", pval=TRUE ),
+            ci.exp( up.z, subset="allo", pval=TRUE ) )
> zz[,1:3] <- 1/zz[,c(1,3,2)]
> load( file="./data/mainCI.Rda" )
> dimnames( mainCI )[[1]][7]
[1] "Retinopathy progression"
> mainCI[7,,] <- zz
> save( mainCI, file="./data/mainCI.Rda" )
```

With these models in place we can now set up a transition object.

```
> Tr <- list( "None" = list( "Min" = up.i,
+                          "D(None)" = mt.0 ),
+           "Min" = list( "Mod" = up.i,
+                         "D(Min)" = mt.0 ),
+           "Mod" = list( "PrePr" = up.i,
+                         "D(Mod)" = mt.0 ),
+           "PrePr" = list( "Pro" = up.i,
+                           "D(PrePr)" = mt.0 ),
+           "Pro" = list( "PhC" = up.i,
+                         "D(Pro)" = mt.0 ),
+           "PhC" = list( "D(PhC)" = mt.0 ) )
> lapply(Tr,names)
$None
[1] "Min" "D(None)"

$Min
[1] "Mod" "D(Min)"
```

```

$Mod
[1] "PrePr" "D(Mod)"

$PrePr
[1] "Pro"      "D(PrePr)"

$Pro
[1] "PhC"      "D(Pro)"

$PhC
[1] "D(PhC)"

```

5.2.4.2 Progression probabilities

In order to assess how retinopathy scores have developed in the two groups we simulate how retinopathy score and death is predicted to develop for different types of patient populations; in all case we simulate in two scenarios: Intensive and Conventional.

- A cohort that has the same joint baseline distribution of retinopathy scores, age and sex as the Steno 2 patients.
- A cohort where all persons have a retinopathy score of “None”, and the same age and sex distribution as the Steno 2 patients.
- Cohorts where all have retinopathy score “None”, and the same baseline age (45,50,55,60,65) and sex. This will be simulated using just one cohort with the 10 different combinations represented.

For the two first scenarios we can use the Lexis objects generated in the previous section, `ini.e` and `ini.0`, respectively.

With the two initiation cohorts we can now simulate the predicted outcomes according to the model for the transitions:

```

> sim.e <- simLexis( Tr=Tr, init=ini.e , time.pts=seq(0,22,0.1), N=100 )
> sim.0 <- simLexis( Tr=Tr, init=ini.0 , time.pts=seq(0,22,0.1), N=100 )
> save( sim.e, sim.0, file="../data/simRet1.Rda" )

```

```

> load( file="../data/simRet1.Rda" )
> summary( sim.e )

```

Transitions:

From	To												
	None	Min	Mod	PrePr	Pro	PhC	D(None)	D(Min)	D(Mod)	D(PrePr)	D(Pro)	D(PhC)	
None	3093	13908	0	0	0	0	4599	0	0	0	0	0	
Min	0	4028	10519	0	0	0	0	4761	0	0	0	0	
Mod	0	0	2030	5915	0	0	0	0	4574	0	0	0	
PrePr	0	0	0	205	6491	0	0	0	0	219	0	0	
Pro	0	0	0	0	282	6115	0	0	0	0	1294	0	
PhC	0	0	0	0	0	3037	0	0	0	0	0	3878	
Sum	3093	17936	12549	6120	6773	9152	4599	4761	4574	219	1294	3878	

Transitions:

From	To			
	Records:	Events:	Risk time:	Persons:
None	21600	18507	160280.17	21600
Min	19308	15280	151980.20	19308
Mod	12519	10489	79838.91	12519
PrePr	6915	6710	15137.05	6915
Pro	7691	7409	17986.99	7691
PhC	6915	3878	73866.54	6915
Sum	74948	62273	499089.86	32000

```
> summary( sim.0 )
Transitions:
  To
From  None   Min   Mod PrePr   Pro  PhC D(None) D(Min) D(Mod) D(PrePr) D(Pro) D(PhC)
None  4534 20525    0    0    0    0   6941    0    0    0    0    0
Min   0   4769 10091    0    0    0    0   5665    0    0    0    0
Mod   0    0  1892  4243    0    0    0    0   3956    0    0    0
PrePr 0    0    0  197 3880    0    0    0    0   166    0    0
Pro   0    0    0    0  233 2823    0    0    0    0   824    0
PhC   0    0    0    0    0  1440    0    0    0    0    0  1383
Sum   4534 25294 11983  4440 4113 4263   6941   5665   3956   166   824  1383
```

```
Transitions:
  To
From  Records:  Events: Risk time:  Persons:
None   32000    27466  237868.87   32000
Min    20525    15756  163158.34   20525
Mod    10091     8199   62758.97   10091
PrePr  4243     4046   9043.21    4243
Pro    3880     3647   8439.08    3880
PhC    2823     1383   24603.63   2823
Sum    73562    60497  505872.10  32000
```

5.2.4.3 Prediction from baseline retinopathy distribution

Here we show how a population of patients with an age- sex- and retinopathy-score distribution as the Steno 2 patients would fare through the states were they subjected to intensive, respectively conventional treatment.

```
> prm <- c(1:6,12:7)
> levels( sim.e )[prm]
[1] "None"      "Min"      "Mod"      "PrePr"    "Pro"      "PhC"      "D(PhC)"
[8] "D(Pro)"    "D(PrePr)" "D(Mod)"   "D(Min)"   "D(None)"

> Ei <- pState( nState( subset(sim.e,allocation=="Intensive"),
+                   at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> Ec <- pState( nState( subset(sim.e,allocation=="Conventional"),
+                   at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> round( head( Ei ), 4 )

      State
when  None   Min   Mod  PrePr   Pro   PhC D(PhC) D(Pro) D(PrePr) D(Mod) D(Min)
0     0.6750 0.8438 0.9062 0.9375 0.9750 1.0000 1.0000 1.0000 1.0000 1.0000 1.0000
0.25 0.6444 0.8376 0.9047 0.9354 0.9724 0.9996 0.9996 0.9997 0.9998 0.9998 0.9998
0.5  0.6198 0.8316 0.9034 0.9336 0.9707 0.9995 0.9995 0.9997 0.9998 0.9998 0.9998
0.75 0.5986 0.8244 0.9014 0.9312 0.9692 0.9992 0.9992 0.9995 0.9996 0.9996 0.9996
1     0.5782 0.8184 0.8983 0.9280 0.9664 0.9983 0.9983 0.9986 0.9986 0.9988 0.9989
1.25 0.5612 0.8121 0.8960 0.9260 0.9641 0.9979 0.9979 0.9982 0.9982 0.9984 0.9987

      State
when  D(None)
0     1
0.25 1
0.5  1
0.75 1
1     1
1.25 1
```

Then we plot the estimated state distribution over the 20 years from baseline

```
> retst(Ei, Ec, rv=FALSE, leg=17)
```

We can also plot it reversely for the conventional group to facilitate comparison

```
> retst(Ei, Ec, rv=TRUE, leg=17)
```

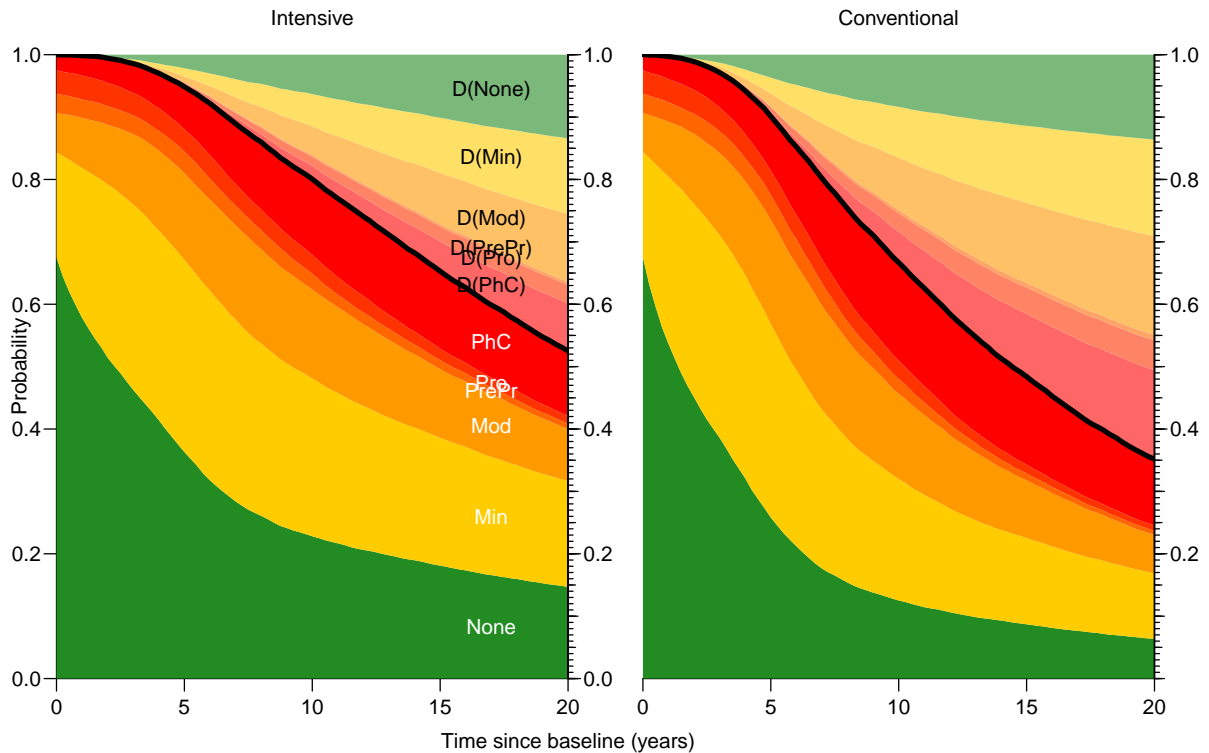


Figure 5.8: Model with only progression possible: Patients with age- sex- and retinopathy score distribution as in Steno 2 study: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which persons have died.

5.2.4.4 Prediction given absence of retinopathy

Here we show how a population of patients with an age- and sex-distribution as the Steno 2 patients all without retinopathy at baseline, would fare through the states were they subjected to intensive, respectively conventional treatment.

```
> Zi <- pState( nState( subset(sim.0,allocation=="Intensive"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> Zc <- pState( nState( subset(sim.0,allocation=="Conventional"),
+                       at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=prm )
> round( head( Zi ), 4 )
```

when	None	Min	Mod	PrePr	Pro	PhC	D(PhC)	D(Pro)	D(PrePr)	D(Mod)	D(Min)
0	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000	1.0000
0.25	0.9559	0.9984	0.9995	0.9995	0.9995	0.9995	0.9995	0.9995	0.9995	0.9995	0.9995
0.5	0.9180	0.9969	0.9991	0.9992	0.9992	0.9992	0.9992	0.9992	0.9992	0.9992	0.9992
0.75	0.8835	0.9945	0.9984	0.9986	0.9986	0.9986	0.9986	0.9986	0.9986	0.9986	0.9986
1	0.8544	0.9909	0.9977	0.9981	0.9981	0.9981	0.9981	0.9981	0.9981	0.9981	0.9981
1.25	0.8269	0.9867	0.9968	0.9974	0.9976	0.9976	0.9976	0.9976	0.9976	0.9976	0.9977

```
State
when  D(None)
0      1
0.25  1
0.5    1
0.75  1
1      1
1.25  1
```

Then plot the estimated distributions over the

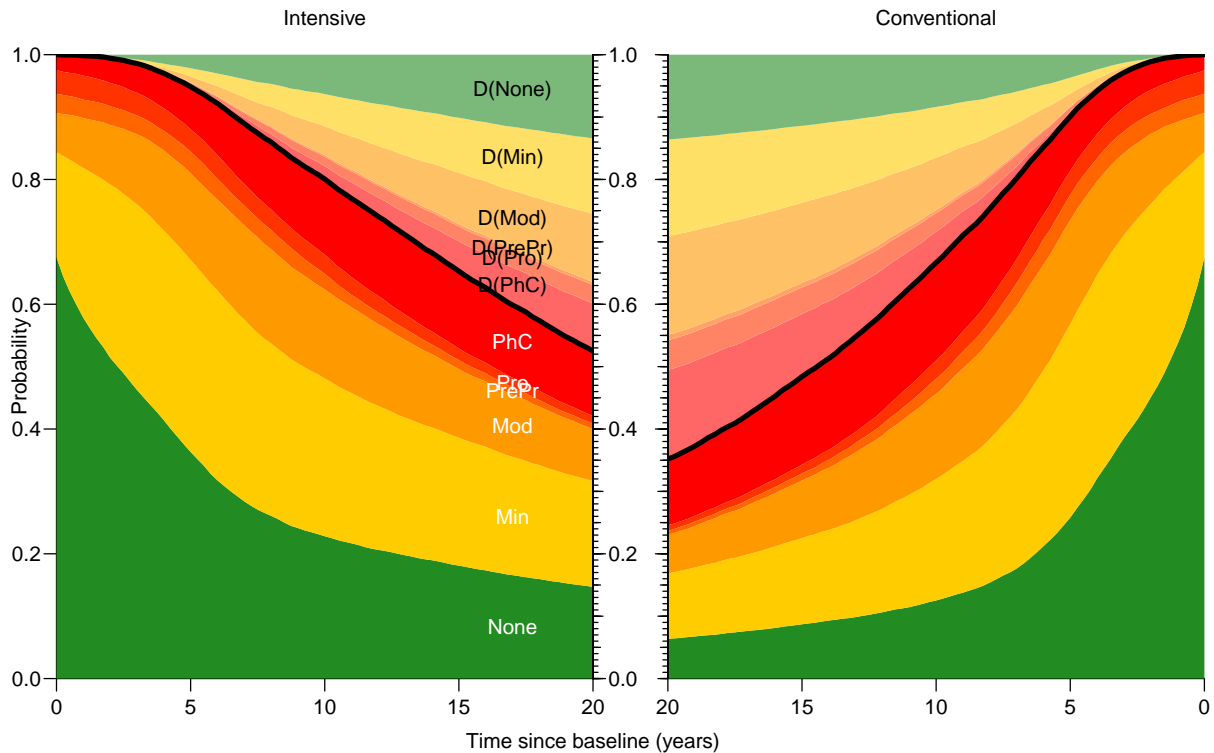


Figure 5.9: *Model with only progression possible: Patients with age- sex- and retinopathy score distribution as in Steno 2 study: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died. This is the same figure as 5.8, except that the x-axis of the right-hand panel is reversed.*

```
> retst(Zi,Zc,rv=FALSE,leg=17)
```

We can also reverse the x -axis for the conventional group to facilitate comparison

```
> retst(Zi,Zc,rv=TRUE,leg=17)
```

5.2.4.5 Predictions by age and sex

Finally we make predictions of the development of retinopathy and death for homogeneous groups of patients all starting with retinopathy score 0, and with identical age and sex. The predictions are made for 10 different combinations of age and sex. First we set up a list structure to hold each of the simulated data sets:

```
> lp <- list()
> length( lp ) <- 20
> dim( lp ) <- c(5,2,2)
> dimnames( lp ) <- list( age = seq(45,65,5),
+                         sex = levels(ini.0$sex),
+                         grp = levels(ini.0$allocation)[2:1] )
```

Then we simulate transitions for each combination of age, sex and allocation:

```
> ini <- ini.0[1,]
> system.time(
+ for( ig in dimnames(lp)[[3]] )
+ for( ia in dimnames(lp)[[1]] )
```

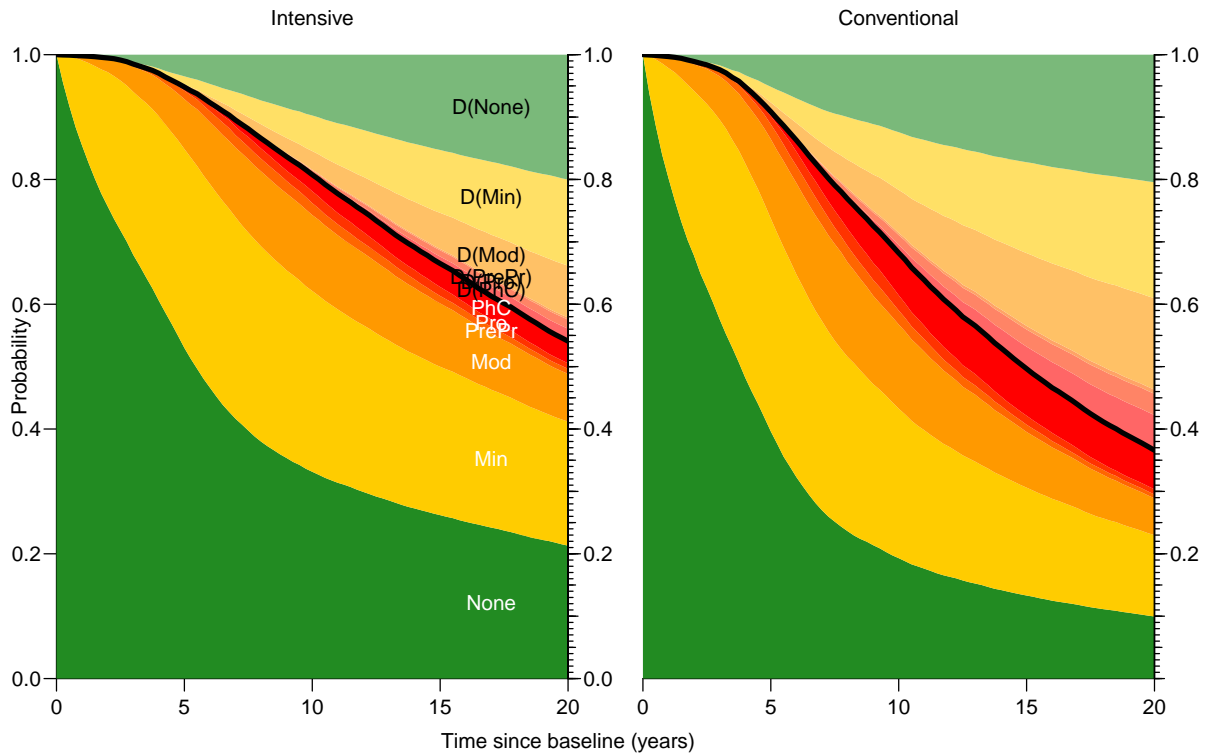


Figure 5.10: Model with only progression possible, assuming all persons initiating without retinopathy and age- and sex-distribution as in Steno 2 study population: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

```
+ for( is in dimnames(lp)[[2]] )
+ {
+ ini[1,"age"] <- as.numeric(ia)
+ ini[1,"sex"] <- is
+ ini[1,"allocation"] <- ig
+ stp <- simLexis( Tr = Tr,
+               init = ini,
+               N = 1000,
+               time.pts = seq(0,21,0.1) )
+ nst <- nState( stp, at=seq(0,20,0.1), from=0, time.scale="tfe" )
+ pst <- pState( nst, perm=prm )
+ lp[[ia,is,ig]] <- pst
+ } )
> save( lp, file="../data/ret1-lp.Rda" )

> load( file="../data/ret1-lp.Rda" )
```

The object `lp` is now a list with 3 dimensions (intervention, age, sex), where each element is a `pState` object, so that we can refer easily to results for any combination of age, sex and allocation.

Using the `lp` object we can now compare the state distribution between persons allocated to intervention and conventional, separately for all ten combinations of age (45, 50, 55, 60, 65) and sex (M, F).

Basically it is the same graph as for the CVD events in the Steno2 population:

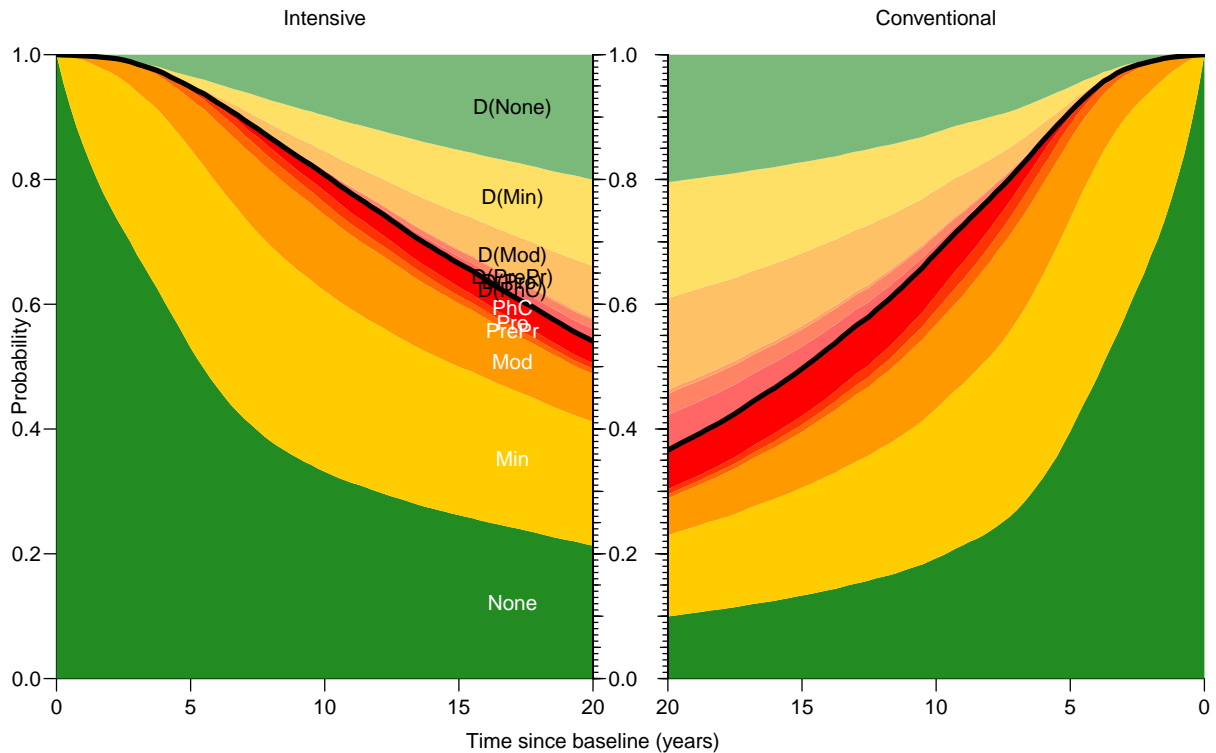


Figure 5.11: Model with only progression possible, assuming all persons initiating without retinopathy and age- and sex-distribution as in Steno 2 study population: Probability of being in each state of retinopathy as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

This is the same figure as 5.10, except that the x-axis of the right-hand panel is reversed.

```
> pl10 <- function(lp){
+ par( mfrow=c(5,4), mar=c(0,0,0,3), oma=c(4,4,4,0), las=1 )
+ for( ia in dimnames(lp)[["age"]] )
+ for( is in dimnames(lp)[["sex"]][2:1] )
+ for( ig in dimnames(lp)[["grp"]][2:1] )
+ {
+ oax <- ( ig==dimnames(lp)[["grp"]][2] )
+ ptmp <- lp[[ia,is,ig]]
+ plot( ptmp, col=clr[prm], xaxt="n", yaxt="n",
+       xlim=if( oax ) c(0,20) else c(20,0) )
+ if( ia=="45" ) mtext( ig, side=3, line=1, at=10, outer=FALSE )
+ if( is=="M" & ig=="Intensive" )
+ {
+ axis( side=4-2*oax, at=1:5/5 )
+ # mtext( ia, side=2, line=3, outer=FALSE )
+ }
+ if( is=="F" & ig=="Conventional" ) mtext(ia,side=4,at=0.5,line=0.5,las=1)
+ axis( side=2, labels=NA )
+ if( ia=="65" )
+ {
+ axis( side=1 )
+ axis( side=4, at=0, labels="0.0" )
+ if( ia=="65" & is=="M" & oax ) axis( side=2, at=0, labels="0.0" )
+ }
+ lines( as.numeric(rownames(ptmp)), ptmp[,4], col="black", lwd=4 )
+ # text( 1, 0.05, paste( is, " ", " ", ia, "\n", ig, esp="" ),
+ #       adj=c(1-oax,0), font=2, col="white", cex=1.5 )
}
```



```
+ axis( side=2+2*oax, at=1:5*20 /100, if(oax) labels=NA )
+ axis( side=2+2*oax, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=2+2*oax, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=2+2*oax, at=0:100 /100, tcl=-0.2, labels=NA )
+   }
+ #mtext( "Men" , side=3, at=0.25, outer=TRUE, line=2 )
+ #mtext( "Women", side=3, at=0.75, outer=TRUE, line=2 )
+ #mtext( "Age at entry", side=3, at=-0.02, outer=TRUE, line=1, adj=0 )
+ mtext( "Time since entry (years)", side=1, outer=TRUE, line=2.5 )
+ mtext( "Probability" , side=2, outer=TRUE, line=2.5, las=0 )
+ mtext( "Men" , side=3, at=0.25, outer=TRUE, line=2.5, las=0 )
+ mtext( "Women", side=3, at=0.75, outer=TRUE, line=2.5, las=0 )
+ mtext( "Age" , side=3, at=1.00, outer=TRUE, line=2.5, las=0, adj=1 )
+ }
> pl10(lp)
```

Finally we save the estimated state distributions:

```
> save( rEi, rEc, rZi, rZc, Ei, Ec, Zi, Zc, file="../data/pRet.Rda" )
```

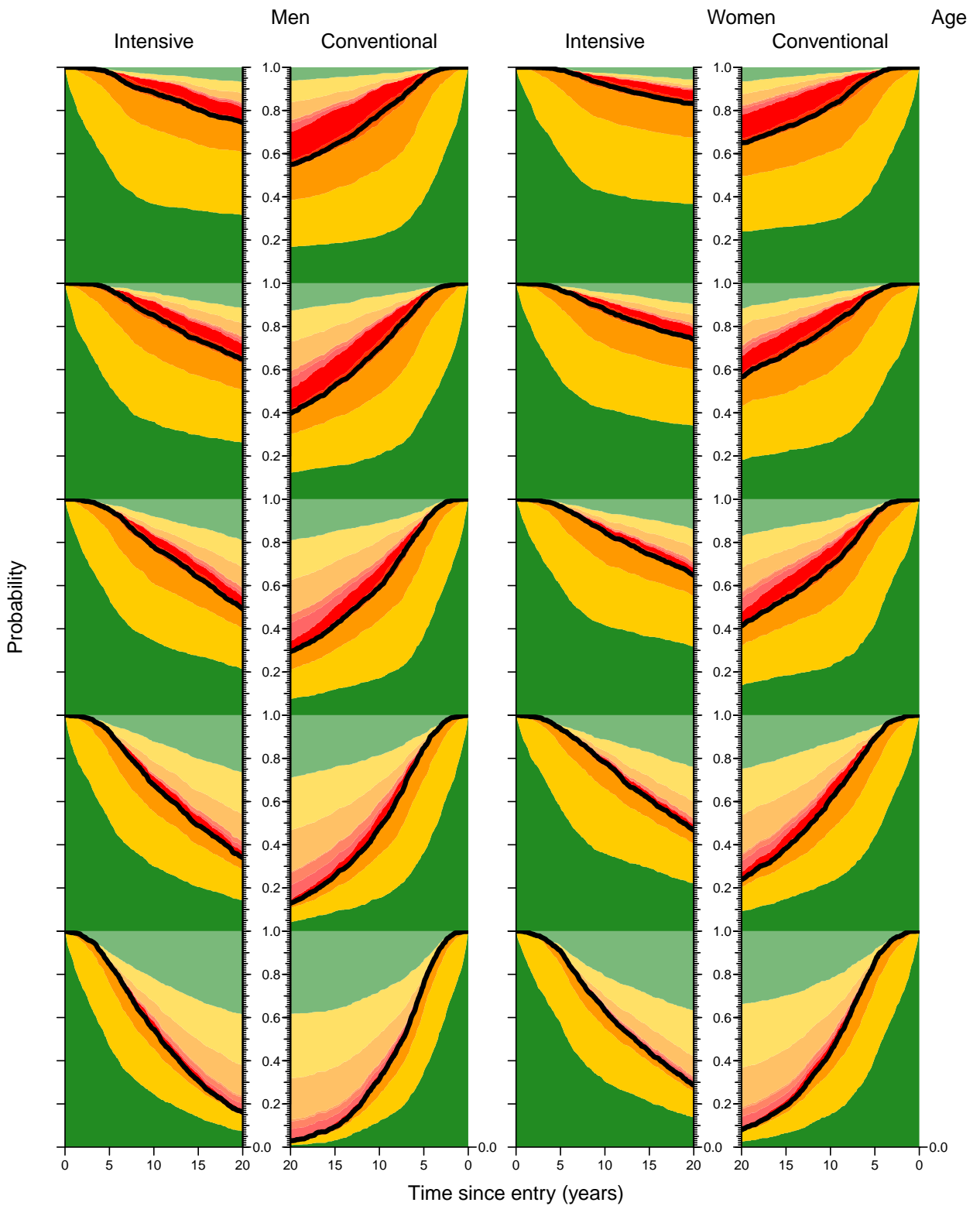


Figure 5.12: Probabilities of being in different states of retinopathy by age, sex and intervention group. State coloring is the same as in figure ??.

5.3 Neuropathy

5.3.1 Data

First we provide an overview of the actually occurring neuropathy patterns in the data, for both types of neuropathy:

```
> library( Epi )
> library( survival )
> clear()
> load( file="../data/mic.Rda" )
> with( mic, ftable(visit,autoprog,periprog,useNA="ifany",col.vars=c(1,3)) )
```

	visit	1			2			3			4			5			6		
autoprog	periprog	0	1	NA	0	1	NA	0	1	NA	0	1	NA	0	1	NA	0	1	NA
0		0	0	0	0	0	121	95	20	0	41	30	0	24	16	0	18	12	1
1		0	0	0	0	0	20	24	8	0	33	19	0	21	22	2	10	12	1
NA		0	0	160	0	0	15	0	2	0	2	1	2	4	1	3	0	0	8

```
> with( mic, ftable(addmargins(table(visit,allocation,autoprog,periprog,useNA="ifany"),
+ margin=2:4),row.vars=c(1,4)) )
```

visit	periprog	allocation Intensive				Conventional				Sum									
		autoprog	0	1	NA	Sum	0	1	NA	Sum	0	1	NA	Sum					
1	0		0	0	0	0		0	0	0	0	0	0	0	0	0	0	0	0
	1		0	0	0	0		0	0	0	0	0	0	0	0	0	0	0	0
	NA		0	0	80	80		0	0	80	80	0	0	160	160				
	Sum		0	0	80	80		0	0	80	80	0	0	160	160				
2	0		0	0	0	0		0	0	0	0	0	0	0	0	0	0	0	0
	1		0	0	0	0		0	0	0	0	0	0	0	0	0	0	0	0
	NA		61	10	7	78		60	10	8	78	121	20	15	156				
	Sum		61	10	7	78		60	10	8	78	121	20	15	156				
3	0		53	7	0	60		42	17	0	59	95	24	0	119				
	1		9	3	1	13		11	5	1	17	20	8	2	30				
	NA		0	0	0	0		0	0	0	0	0	0	0	0				
	Sum		62	10	1	73		53	22	1	76	115	32	2	149				
4	0		25	14	0	39		16	19	2	37	41	33	2	76				
	1		19	7	1	27		11	12	0	23	30	19	1	50				
	NA		0	0	1	1		0	0	1	1	0	0	2	2				
	Sum		44	21	2	67		27	31	3	61	71	52	5	128				
5	0		17	15	2	34		7	6	2	15	24	21	4	49				
	1		10	8	1	19		6	14	0	20	16	22	1	39				
	NA		0	1	1	2		0	1	2	3	0	2	3	5				
	Sum		27	24	4	55		13	21	4	38	40	45	8	93				
6	0		15	6	0	21		3	4	0	7	18	10	0	28				
	1		8	7	0	15		4	5	0	9	12	12	0	24				
	NA		0	0	4	4		1	1	4	6	1	1	8	10				
	Sum		23	13	4	40		8	10	4	22	31	23	8	62				

So there are no assessment of peripheral neuropathy progression until at the third visit. We provide an overview of the patterns of neuropathy progressions over time; visits with no assessment are shown with a “-”, no progression with 0 and progression with 1.

```
> pt <- with(mic,tapply( periprog, list(ID,visit), mean ))
> pp <- gsub("NA","-",apply( pt, 1, paste, collapse=" "))
> cbind( sort( addmargins( table( pp ) ) ) )
```

	[,1]
- - - - 0 -	1
- - 0 0 - 1	1
- - 0 1 0 1	1
- - 0 1 - 1	1
- - - 1 - -	1
- - - 1 - 0	1
- - 1 1 - 1	1

```

-- 0 - 0 -      2
-- 0 0 1 0      2
-- 0 1 1 0      2
-- 1 1 1 0      2
-- 0 1 1 1      3
-- 0 0 1 1      4
-- 0 1 0 0      4
-- 0 1 - -      5
-- 1 - - -      5
-- 0 0 0 1      6
-- 0 0 1 -      6
-- 1 1 1 -      6
-- 0 1 1 -      7
-- 1 1 1 1      7
-- - - - -      8
-- 1 1 - -      9
-- 0 0 0 0     17
-- 0 - - -     18
-- 0 0 0 -     18
-- 0 0 - -     22
Sum              160
> at <- with(mic,tapply( autoprog, list(ID,visit), mean ))
> aa <- gsub("NA","-",apply( at, 1, paste, collapse=" "))
> cbind( sort( addmargins( table( aa ) ) ) )
      [,1]
-- - 0 - -      1
-- 0 - - -      1
- 0 0 0 - 1      1
-- 0 0 1 -      1
- 0 0 - 1 -      1
- 0 0 1 - 0      1
- 0 0 1 - 1      1
- 0 0 1 1 0      1
- 0 - 1 - 0      1
- 0 1 - 0 -      1
- 0 1 0 1 0      1
- 0 1 1 0 -      1
- 0 1 1 1 0      1
-- 1 - - -      1
- 1 0 0 0 -      1
- 1 0 0 1 -      1
-- 1 0 1 -      1
- 1 0 1 0 0      1
- 1 0 1 1 0      1
- 1 0 1 1 1      1
-- 1 1 - -      1
- 1 1 0 - -      1
- 1 1 0 0 -      1
- 1 1 - - 1      1
- 1 1 1 0 -      1
- 1 1 1 0 0      1
- 1 1 1 1 1      1
-- 0 0 0 0      2
- 0 0 0 0 1      2
- 0 0 1 0 -      2
- 0 0 1 0 0      2
- 0 0 1 0 1      2
-- 0 1 - -      2
- 0 1 1 1 -      2
- 1 - - - -      2
- 1 0 - - -      2
- 1 1 1 1 -      2
- 1 1 1 - -      3
- 0 - - - -      4
-- 0 0 - -      4
- 0 0 0 1 0      4

```

```

- 0 1 - - - 4
- 0 1 0 - - 4
- 0 1 1 - - 4
- - - - - 5
- 0 0 1 1 1 5
- 0 0 0 1 - 6
- 0 0 1 1 - 7
- 0 0 0 - - 8
- 0 0 0 0 - 8
- 0 0 1 - - 8
- 0 0 0 1 1 9
- 0 0 - - - 15
- 0 0 0 0 0 15
Sum          160

```

As for the other microvascular outcomes we generate states and dates. Since there are some persons who has no assessments of neuropathy at all we cannot include these in the calculations, so they are excluded *a priori*:

```

> vd <- with( mic, tapply( doVis, list(ID,visit), mean ) )
> at <- with( mic, tapply( autoprog, list(ID,visit), mean ) )
> pt <- with( mic, tapply( periprog, list(ID,visit), mean ) )
> cbind( at, pt, vd )[1:10,]

```

	1	2	3	4	5	6	1	2	3	4	5	6	1	2	3	4	5
1	NA	0	0	0	0	0	NA	NA	0	0	0	0	1993.357	1995.158	1997.135	2001.097	2006.712
2	NA	1	1	1	1	1	NA	NA	0	1	0	1	1993.357	1995.377	1997.781	2001.814	2006.868
3	NA	0	0	1	NA	NA	NA	NA	0	1	NA	NA	1993.362	1995.180	1997.151	2001.102	NA
4	NA	NA	0	0	NA	NA	NA	NA	1	1	NA	NA	1993.362	1995.120	1997.110	2001.066	NA
5	NA	1	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	1993.162	1995.002	1997.707	NA	NA
6	NA	1	1	1	1	NA	NA	NA	0	0	1	NA	1993.201	1995.008	1997.036	2001.025	2006.736
7	NA	0	1	1	NA	NA	NA	NA	0	1	NA	NA	1993.203	1995.043	1997.077	2001.058	NA
8	NA	0	1	1	1	NA	NA	NA	0	0	0	NA	1993.209	1995.829	1997.729	2001.655	2006.903
9	NA	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	1993.209	1995.021	NA	NA	NA
10	NA	0	1	1	1	NA	NA	NA	0	0	0	NA	1993.075	1995.024	1997.020	2001.023	2006.682

```

6
1 2014.802
2 2014.589
3 NA
4 NA
5 NA
6 2015.029
7 NA
8 NA
9 NA
10 NA

> table( wh.a <- apply( at, 1, function(x) any(!is.na(x)) ) )
FALSE TRUE
5 155

> table( wh.p <- apply( pt, 1, function(x) any(!is.na(x)) ) )
FALSE TRUE
8 152

```

Since we are recording *progression* of neuropathy since baseline, we set the state to 0 at baseline:

```

> at[,1] <- 0
> pt[,1] <- 0

```

5.3.2 Autonomous and peripheral neuropathy

We then set up a proper Lexis objects using `make.dlex` separately for autonomous and peripheral neuropathy. These objects are based on simulated transition times that are uniformly distributed on the intervals in which status changes has taken place, and an assumption of the last registered state being carried over to either death of last recorded visit. Entry is taken as the first visit with a neuropathy measurement. Since the measurements are first made at either 2nd visit (auto) or 3rd visit (peri) follow-up starts at the 2nd visit for autonomous neuropathy, when including peripheral neuropathy we are essentially assuming absence of peripheral neuropathy at 2nd visit.

Since we are simulating an exact transition time we need a random generator seed in order to make the dataset reproducible:

```
> source("usefuns.R")
> set.seed(7471568)
> LxA <- make.dlex( at[wh.a,], vd[wh.a,], incr=TRUE )
> LxP <- make.dlex( pt[wh.p,], vd[wh.p,], incr=TRUE )
> levels(LxA$lex.Cst) <- levels(LxA$lex.Xst) <- c("None", "Auto", "D(None)", "D(Auto)")
> levels(LxP$lex.Cst) <- levels(LxP$lex.Xst) <- c("None", "Peri", "D(None)", "D(Peri)")
> summary( LxA )
```

Transitions:

	To							
From	None	Auto	D(None)	D(Auto)	Records:	Events:	Risk time:	Persons:
None	338	97	37	0	472	134	1319.06	155
Auto	0	229	0	52	281	52	1058.67	97
Sum	338	326	37	52	753	186	2377.73	155

```
> summary( LxP )
```

Transitions:

	To							
From	None	Peri	D(None)	D(Peri)	Records:	Events:	Risk time:	Persons:
None	264	74	53	0	391	127	1536.82	152
Peri	0	168	0	33	201	33	833.78	74
Sum	264	242	53	33	592	160	2370.59	152

5.3.3 Joint occurrence of autonomous and peripheral neuropathy

In order to set up transitions through states Auto and Peri in combination, we start out with the `LxA` object (the one with the longest risk time), and then cut this at the dates of entry to the Peri state.

```
> lxA <- Relevel( LxA, list("W"=1, "A"=2, "D"=3:4), print=F )
> lxP <- Relevel( LxP, list("W"=1, "P"=2, "D"=3:4), print=F )
> summary( lxA )
```

Transitions:

	To						
From	W	A	D	Records:	Events:	Risk time:	Persons:
W	338	97	37	472	134	1319.06	155
A	0	229	52	281	52	1058.67	97
Sum	338	326	89	753	186	2377.73	155

```
> summary( lxP )
```

Transitions:

	To						
From	W	P	D	Records:	Events:	Risk time:	Persons:
W	264	74	53	391	127	1536.82	152
P	0	168	33	201	33	833.78	74
Sum	264	242	86	592	160	2370.59	152

```
> ( noP <- setdiff( lxA$lex.id, lxP$lex.id ) )
```

```
[1] 9 66 76 134
> ( noA <- setdiff( lxP$lex.id, lxA$lex.id ) )
[1] 60
> length( bothAP <- intersect( lxP$lex.id, lxA$lex.id ) )
[1] 151
```

We note that there are 4 persons with autonomous and no peripheral assessment and one who is vice versa, and 151 patients with at least one assessment of both types of neuropathy. The joint analysis will be based on these persons.

```
> lxA <- subset( lxA, lex.id %in% bothAP )
> lxP <- subset( lxP, lex.id %in% bothAP )
```

From the object with peripheral neuropathy measurements we derive the dates of onset which we shall use to cut the follow-up for autonomous neuropathy. From the peripheral Lexis object we derive the first date of “P”:

```
> cutP <- subset( lxP, lex.Cst=="P", select=c("lex.id", "per") )
> cutP <- cutP[match(unique(cutP$lex.id), cutP$lex.id),]
> names( cutP ) [2] <- "cut"
> head( cutP )
  lex.id      cut
7       2 1999.180
12      3 1998.809
14      4 1996.584
20      5 1993.845
25      6 2003.266
29      7 1998.171
```

Then we cut the records originating in the “W” state, meaning that the new state must be “P”:

```
> cutP$new.state = "P"
> lxWP <- cutLexis( subset(lxA, lex.Cst=="W"),
+                  cut=cutP, timescale="per", precursor.states=c("W") )
> summary( subset(lxA, lex.Cst=="W") )
Transitions:
  To
From W A D Records: Events: Risk time: Persons:
W 335 96 34      465      130    1308.22      151
> summary( lxWP )
Transitions:
  To
From W P A D Records: Events: Risk time: Persons:
W 252 46 72 25      395      143    935.70      151
P 0 83 24 9      116      33    372.51      46
Sum 252 129 96 34      511      176    1308.22      151
```

However in this object, the transitions labeled from “P” to “A”, are really from “P” to “A+P”, so we fix this:

```
> lxWP <- transform( lxWP, lex.Xst = ifelse( lex.Cst=="P" & lex.Xst=="A",
+                                          "A+P", as.character(lex.Xst) ) )
> summary( lxWP )
Transitions:
  To
From A A+P D P W Records: Events: Risk time: Persons:
W 72 0 25 46 252      395      143    935.70      151
P 0 24 9 83 0      116      33    372.51      46
Sum 72 24 34 129 252      511      176    1308.22      151
```

Then we cut the records originating in the “A” state, meaning that the new state must be “A+P”; note that “A” is now taken as a precursor state too:

```
> cutP$new.state = "A+P"
> lxAP <- cutLexis( subset(lxA,lex.Cst=="A"),
+                 cut=cutP, timescale="per", precursor.states=c("W","A") )
> summary( lxAP )
Transitions:
  To
From W  A A+P D  Records:  Events: Risk time:  Persons:
  A   0 133 27 28     188      55    598.33      72
  A+P 0   0 95 23     118      23    459.54      51
  Sum 0 133 122 51     306      78   1057.87     96
```

Finally we stack the two and check that it really is an expansion of the lxA:

```
> LxAP <- Relevel( rbind( lxAP, lxWP ) )
> levels( LxAP )
[1] "W" "A" "A+P" "P" "D"
```

Ultimately we would also like to see how many died from which state, so we split the Death state:

```
> LxAP <- transform( LxAP, lex.Xst = ifelse( lex.Xst=="D",
+                                           paste("D(",as.character(lex.Cst),")",sep=""),
+                                           as.character(lex.Xst) ) )
> levels( Relevel( LxAP ) )
[1] "W" "A" "A+P" "P" "D(A)" "D(A+P)" "D(P)" "D(W)"
> LxAP <- Relevel( LxAP, c(1,2,4,3,8,5,7,6) )
> levels( LxAP )
[1] "W" "A" "P" "A+P" "D(W)" "D(A)" "D(P)" "D(A+P)"
> levels( LxAP$lex.Cst ) <-
+ levels( LxAP$lex.Xst ) <- gsub("W","Well",
+                               gsub("A","Auto",
+                               gsub("P","Peri",levels(LxAP))))
> summary( lxA )
Transitions:
  To
From W  A  D  Records:  Events: Risk time:  Persons:
  W   335 96 34     465      130    1308.22     151
  A    0 228 51     279      51    1057.87     96
  Sum 335 324 85     744      181    2366.09     151
> summary( LxAP )
Transitions:
  To
From Well Auto Peri Auto+Peri D(Well) D(Auto) D(Peri) D(Auto+Peri) Records:
  Well 252 72 46 0 25 0 0 0 395
  Auto 0 133 0 27 0 28 0 0 188
  Peri 0 0 83 24 0 0 9 0 116
  Auto+Peri 0 0 0 95 0 0 0 23 118
  Sum 252 205 129 146 25 28 9 23 817
Transitions:
  To
From Events: Risk time: Persons:
  Well 143 935.70 151
  Auto 55 598.33 72
  Peri 33 372.51 46
  Auto+Peri 23 459.54 51
  Sum 254 2366.09 151
```


5.3.4 Collected data for neuropathy

We now have the Lexis objects for separate analysis of autonomous and peripheral neuropathy. For the sake of completeness we merge in the date of diabetes diagnosis and date of baseline from the baseline dataset:

```
> load( "../data/st2.Rda" )
> stb <- st2[,c("ID","doDM","doBase")]
> names( stb )[1] <- "lex.id"
> str( stb )
'data.frame':      160 obs. of  3 variables:
 $ lex.id: num  1 2 3 4 5 6 7 8 9 10 ...
 $ doDM  : num 1991 1982 1983 1977 1986 ...
 $ doBase: num 1993 1993 1993 1993 1993 ...
> LxA <- merge( LxA , stb , all.x=TRUE )
> LxP <- merge( LxP , stb , all.x=TRUE )
> LxAP <- merge( LxAP, stb , all.x=TRUE )
```

Then we save the three Lexis objects:

```
> save( LxA, LxP, LxAP, file="../data/LxNeu.Rda" )
```

```
> clr <- c( "forestgreen", heat.colors(5)[c(2,3,1)] )
> clr <- c(clr,rgb(t(col2rgb(clr)*0.6+255*0.4),max=255))
> summary( LxAP )
```

Transitions:

From	To	Well	Auto	Peri	Auto+Peri	D(Well)	D(Auto)	D(Peri)	D(Auto+Peri)	Records:
Well		252	72	46	0	25	0	0	0	395
Auto		0	133	0	27	0	28	0	0	188
Peri		0	0	83	24	0	0	9	0	116
Auto+Peri		0	0	0	95	0	0	0	23	118
Sum		252	205	129	146	25	28	9	23	817

Transitions:

From	To	Events:	Risk time:	Persons:
Well		143	935.70	151
Auto		55	598.33	72
Peri		33	372.51	46
Auto+Peri		23	459.54	51
Sum		254	2366.09	151

```
> par( mfcol=c(1,2), oma=c(0,0,0,0) )
> for( lv in levels(LxAP$allocation) )
+ {
+ boxes( subset( LxAP, allocation==lv ),
+         boxpos=list(x=c(20,30,30,80,50,80,80,80)+7-14*(lv=="Intensive"),
+                       y=c(65,90,10,60,48,90,10,30)),
+         show.BE="noz", scale.R=100,
+         # pos.arr=c(0.45,0.3,0.3),
+         col.bg=clr,
+         col.border=c(clr[1:4],rep("black",4)),
+         col.txt=rep(c("white","black"),each=4) )
+ text(10+5-10*(lv=="Intensive"),98,lv,cex=1.5,font=2,adj=0)
+ }
```

```
> clr <- c( "forestgreen", "orange" )
> clr <- c(clr,rgb(t(col2rgb(clr)*0.6+255*0.4),max=255))
> i <- 0
> par( mfcol=c(2,2), oma=c(0,0,0,0) )
> for( lv in levels(LxA$allocation) )
+ for( Lx in list(LxA,LxP) )
```

```

+ {
+ i <- i + 1
+ boxes( subset( Lx, allocation=="lv ),
+        boxpos=list(x=rep(c(20,80),each=2)+7-14*(lv=="Intensive"),
+                    y=rep(c(75,15),2)),
+        show.BE="noz", scale.R=100,
+        pos.arr=c(0.45,0.3,0.3), col.bg=clr,
+        col.border=c(clr[1:2],rep("black",2)),
+        col.txt=rep(c("white","black"),each=2) )
+ if( i %in% c(1,3) ) text(10+5-10*(lv=="Intensive"),98,lv,cex=1.5/0.83,font=2,adj=0)
+ }

```

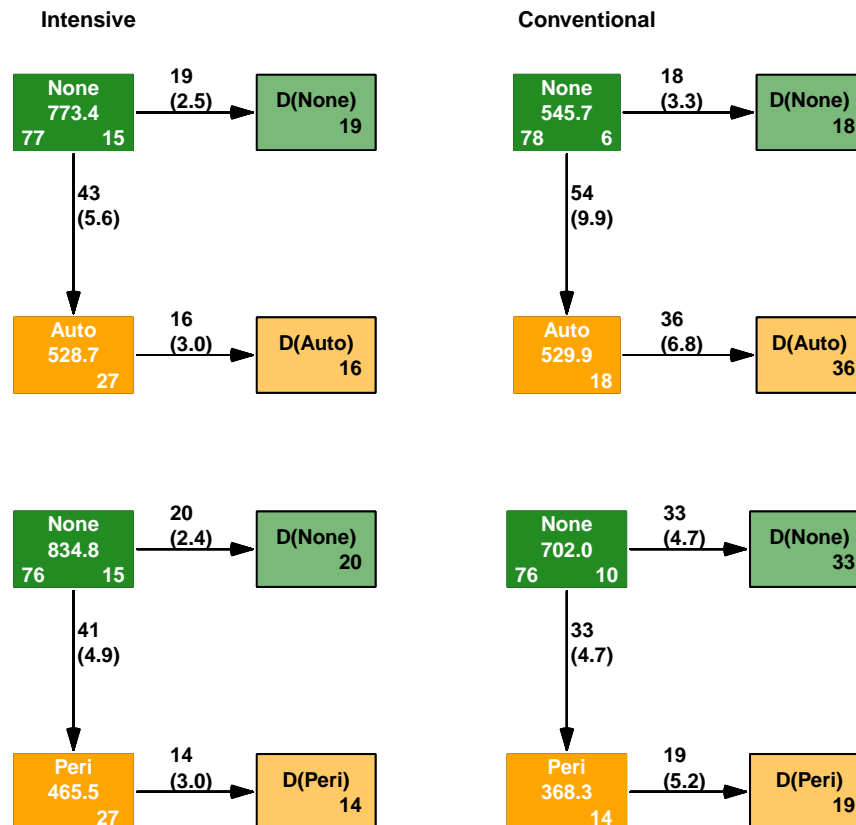


Figure 5.13: Progression of autonomous and peripheral neuropathy, respectively. The number of persons is smaller than the total number randomized, because only persons with at least one assessment of the relevant type of neuropathy are included. The number in the center of the boxes is the person-years (PY), the numbers at the bottom is the number of patients starting, resp. ending their follow-up in each box (state), and the numbers on the arrows are the number of transitions and the overall transition rates per 100 PY.

5.3.5 Autonomous neuropathy

Then we fit a simple Cox-model with allocation as the only covariate and amend it by age and sex:

```

> a1 <- coxph( Surv(tfe,tfe+lex.dur,lex.Xst=="Auto") ~ allocation, data = LxA)
> a2 <- update( a1, . ~ . + I((age-55)/10) + sex )
> round( ci.exp( a2 ), 3 )

```

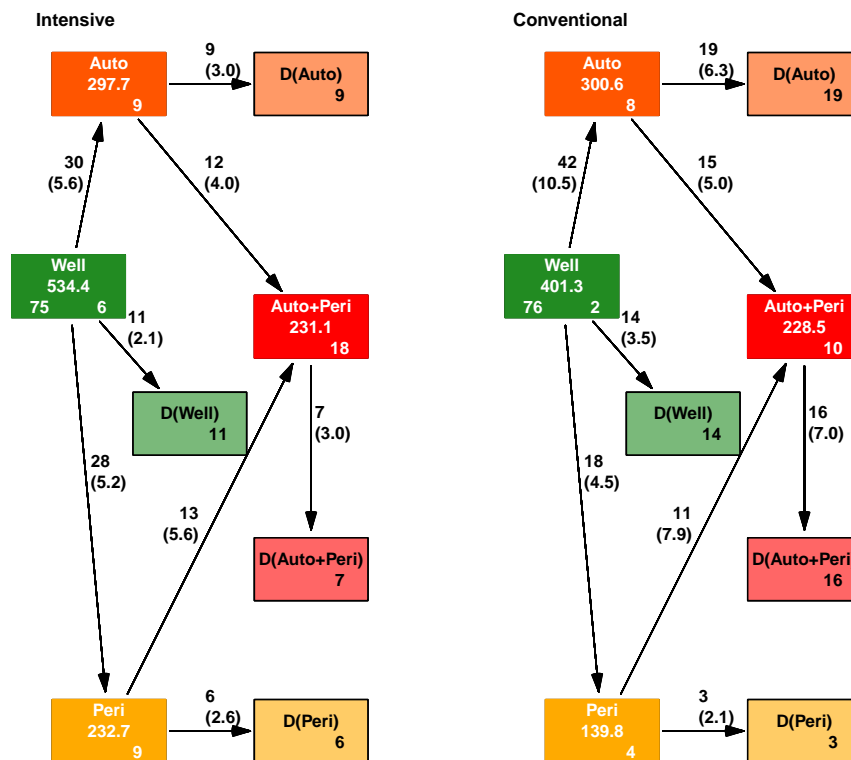


Figure 5.14: Joint progression of autonomous and peripheral neuropathy. The number of persons is smaller than the total number randomized, because only persons with at least one assessment of both types of neuropathy are included. The number in the center of the boxes is the person-years (PY), the numbers at the bottom is the number of patients starting, resp. ending their follow-up in each box (state), and the numbers on the arrows are the number of transitions and the overall transition rates per 100 PY.

```

                                exp(Est.)  2.5% 97.5%
allocationConventional          1.303 1.047 1.622
I((age - 55)/10)                 1.020 0.870 1.196
sexMale                           1.299 1.000 1.687
> HR <- rbind( ci.exp( a1, pval=TRUE ),
+             ci.exp( a2, pval=TRUE ) )
> HR <- cbind( 1/HR[,c(1,3,2)], HR )
> rownames( HR )[1:2] <- c("Unadj.", "Adj sex age")
> colnames( HR )[c(1,4)] <- c("Int vs Conv", "Conv vs Int")
> round( HR[1:2,], 3 )

                Int vs Conv 97.5%  2.5% Conv vs Int  2.5% 97.5%    P
Unadj.           0.774 0.622 0.963          1.292 1.038 1.608 0.022
Adj sex age      0.767 0.616 0.955          1.303 1.047 1.622 0.018

```

So we see that there is a 25% lower occurrence rate of autonomous neuropathy in the intervention group (HR=0.75, 95% c.i.: 0.59–0.95), $p=0.017$, hence a significant effect of intervention, but the confidence interval covers both trivial and substantial effects. The model controlled for age and sex shows a similar intervention effect (HR 0.73 (0.57,0.93)), and reveals that there is virtually no age-effect but a 40% higher HR of autonomous neuropathy among men as compared to women.

5.3.5.1 Prediction of burden

In order to predict the risk of autonomous neuropathy we fit parametric models for the transitions. So we split the follow-up in pieces of 1 month:

```
> SxA <- splitLexis( LxA, breaks=seq(0,25,1/12), time.scale="tfe" )
```

For convenience we use one set of knots for the mortalities, and one set for incidence of neuropathy:

```
> summary( LxA )
Transitions:
  To
From  None Auto D(None) D(Auto)  Records:  Events: Risk time:  Persons:
None  338  97    37      0      472      134   1319.06     155
Auto   0 229    0      52      281      52   1058.67     97
Sum   338 326    37     52      753     186  2377.73    155
> summary( SxA )
Transitions:
  To
From  None  Auto D(None) D(Auto)  Records:  Events: Risk time:  Persons:
None 16091  97    37      0     16225    134   1319.06     155
Auto   0 12932  0      52     12984    52   1058.67     97
Sum  16091 13029  37     52     29209    186  2377.73    155
> ( d.kn <- with( subset( SxA, lex.Xst %in% c("D(None)", "D(Auto)") ),
+               quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
  12.5%   37.5%   62.5%   87.5%
5.426420 9.119781 12.479124 16.870637
> ( n.kn <- with( subset( LxA, lex.Cst=="None" & lex.Xst=="Auto" ),
+               quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
  12.5%   37.5%   62.5%   87.5%
1.294970 3.442148 6.201048 10.236395
```

We fit models for mortality including age and sex and test for interaction between allocation and time since baseline:

```
> m1 <- glm( (lex.Xst %in% c("D(None)", "D(Auto)")) ~
+           Ns( tfe, knots=d.kn ) + sex + I((age-tfe)/10) + allocation + lex.Cst,
+           offset=log(lex.dur),
+           family=poisson,
+           data = SxA )
> mi <- update( m1, . ~ . + allocation:lex.Cst )
> mn <- update( m1, . ~ . + allocation:tfe )
> anova( mn, m1, mi, test="Chisq" )
Analysis of Deviance Table

Model 1: (lex.Xst %in% c("D(None)", "D(Auto)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst + allocation:tfe
Model 2: (lex.Xst %in% c("D(None)", "D(Auto)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst
Model 3: (lex.Xst %in% c("D(None)", "D(Auto)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst + allocation:lex.Cst
  Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1      29200      1130.7
2      29201      1130.7 -1 -0.006715  0.9347
3      29200      1130.4  1  0.277018  0.5987
> round( ci.exp( m1, pval=T ), 3 )
              exp(Est.)  2.5%  97.5%    P
(Intercept)          0.000 0.000  0.000 0.000
Ns(tfe, knots = d.kn)1  1.512 0.692  3.305 0.300
Ns(tfe, knots = d.kn)2  7.023 3.238 15.232 0.000
Ns(tfe, knots = d.kn)3  1.332 0.760  2.335 0.317
sexMale              1.384 0.833  2.300 0.209
I((age - tfe)/10)    2.803 1.991  3.948 0.000
allocationConventional 2.076 1.339  3.220 0.001
lex.CstAuto          0.969 0.619  1.517 0.890
```

We see that there is no sign of interaction between allocation and state (None/Auto) nor time since baseline — both p-values are over 0.7. Moreover there is no difference in mortality between those with or without autonomous neuropathy.

The occurrence of autonomous neuropathy is modelled likewise (this is essentially the same model as the Cox-model above, except that we model the baseline intensity parametrically):

```
> n1 <- glm( (lex.Xst == "Auto") ~
+           Ns( tfe, knots=d.kn) + sex + I(age-tfe) + allocation,
+           offset=log(lex.dur),
+           family=poisson,
+           data = subset(SxA, lex.Cst=="None" ) )
> ni <- update( n1, . ~ . + allocation:tfe )
> n2 <- update( ni, . ~ . + allocation:I(tfe^2) )
> aov <- anova( n1, ni, n2, test="Chisq")[-1,3:5]
> rownames( aov ) <- c("allo x time","allo x t-sq")
> round( aov, 3 )
      Df Deviance Pr(>Chi)
allo x time 1    0.964  0.326
allo x t-sq 2    5.728  0.057
> round( ci.exp( ni, pval=T ), 3 )
              exp(Est.)  2.5% 97.5%    P
(Intercept)           0.011 0.002 0.064 0.000
Ns(tfe, knots = d.kn)1 0.934 0.222 3.929 0.925
Ns(tfe, knots = d.kn)2 0.166 0.035 0.795 0.025
Ns(tfe, knots = d.kn)3 0.054 0.006 0.520 0.011
sexMale                1.509 0.935 2.436 0.092
I(age - tfe)           1.022 0.994 1.051 0.128
allocationConventional 2.344 1.135 4.841 0.021
allocationIntensive:tfe 1.056 0.946 1.180 0.332
allocationConventional:tfe 1.000 1.000 1.000 NaN
```

We then update the HR collector for autonomic neuropathy

```
> nz <- update( n1, . ~ . - I(age-tfe) - sex )
> zz <- rbind( ci.exp( nz, subset="allo", pval=TRUE ),
+            ci.exp( n1, subset="allo", pval=TRUE ) )
> zz[,1:3] <- 1/zz[,c(1,3,2)]
> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][8]
[1] "Autonomic neuropathy"
> mainCI[8,,] <- zz
> round( ftable(mainCI,row.vars=1), 3 )
      model      raw      age/sex
      what Estimate 2.5% 97.5% P Estimate 2.5% 97.5% P
outcome
All cause mortality 0.551 0.364 0.835 0.005 0.526 0.346 0.800 0.003
CVD mortality       0.379 0.191 0.754 0.006 0.353 0.177 0.705 0.003
non-CVD mortality  0.703 0.413 1.197 0.195 0.686 0.401 1.172 0.168
Death or 1st CVD   NA    NA    NA    NA    NA    NA    NA    NA
Death | CVD state  0.835 0.536 1.301 0.425 0.874 0.564 1.355 0.548
CVD event | CVD state 0.552 0.394 0.772 0.001 0.575 0.412 0.803 0.001
Retinopathy progression 0.668 0.507 0.881 0.004 0.673 0.511 0.887 0.005
Autonomic neuropathy 0.594 0.397 0.890 0.011 0.573 0.382 0.859 0.007
Peripheral neuropathy 1.120 0.707 1.774 0.630 1.101 0.694 1.747 0.683
Macroalbuminuria   0.516 0.316 0.842 0.008 0.495 0.302 0.811 0.005
> save( mainCI, file="../data/mainCI.Rda" )
```

There is a non-significant interaction with time since baseline, with decreasing allocation effect by time as seen in figure 5.15, and the curvature term is borderline significant. We can show how the estimated linear and quadratic effects look:

```

> tpt <- seq(2,20,0.5)
> CM <- cbind(-1,tpt,-tpt,tpt^2,-(tpt^2))
> RRc <- ci.exp( n1, subset="allo", ctr.mat=CM[,1,drop=F] )
> RRl <- ci.exp( ni, subset="allo", ctr.mat=CM[,1:3] )
> RRq <- ci.exp( n2, subset="allo", ctr.mat=CM )
> matplot( tpt, cbind(RRc,RRq,RRl),
+         type="l", lty=1, lwd=c(3,1,1),
+         log="y", ylim=c(0.1,5),
+         col=rep(c(gray(0.5),"blue"),c(6,3)),
+         xlab="Time since baseline (years)",
+         ylab="HR of autonomous neuropathy: Int. vs Conv." )
> axis( side=1, at=2:20, tcl=-0.4, labels=NA )
> abline( h=1, col=gray(0.7) )

```

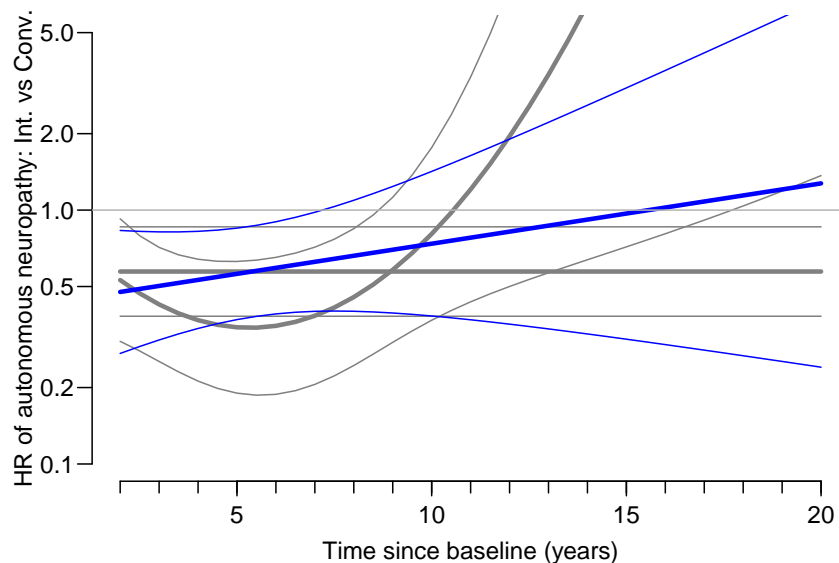


Figure 5.15: Hazard ratio of autonomous neuropathy progression between intensive and conventional arms by time since baseline. Three different models: Constant (proportional hazards, gray), linear (blue) and quadratic (gray) interactions with time.

The linear interaction model is not better than the proportional hazards model ($P=0.327$), and although the curvature term is borderline significant ($P=0.058$), the shape of it as shown in figure 5.15 is not credible and so we shall stick to the proportional hazards model.

With these models fitted we shall show the probability of having autonomous neuropathy in the two groups, so we set up a transition object as well as a baseline object from which to simulate.

```

> trA <- list( "None" = list( "Auto" = n1,
+                           "D(None)" = m1 ),
+            "Auto" = list( "D(Auto)" = m1 ) )
> lapply( trA, names )
$None
[1] "Auto" "D(None)"

$Auto
[1] "D(Auto)"

```

Since autonomous neuropathy is only measured as from the 2nd visit, we set the starting time to 2 years, but we use as initiating state for each person the status at the first

measurement of autonomous neuropathy (which for a few persons is at a later visit than the 2nd). We use two copies of this dataset (that is the distribution of age, sex and neuropathy) one where we assume intensive treatment and one where we assume conventional:

```
> # selecting columns with "[" loses the attributes so this is not applicable:
> # iniA <- LxA[match(unique(LxA$lex.id),LxA$lex.id),c("sex","allocation","lex.Cst","tfe","age","per")]
> iniA <- subset( LxA, per==ave(per,lex.id,FUN=min),
+               select=c("sex","allocation","lex.Cst","tfe","age","per") )
> iniA$tfe = 0
> iniA <- rbind(iniA,iniA)
> iniA$allocation <- factor( rep( levels(iniA$allocation),
+                               each=nrow(iniA)/2 ) )
> str( iniA )
Classes 'Lexis' and 'data.frame':      310 obs. of  6 variables:
 $ sex      : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 1 2 2 ...
 $ allocation: Factor w/ 2 levels "Conventional",...: 2 2 2 2 2 2 2 2 ...
 $ lex.Cst   : Factor w/ 4 levels "None","Auto",...: 1 1 1 1 1 1 1 1 ...
 $ tfe      : num  0 0 0 0 0 0 0 0 ...
 $ age      : num  61.1 46.5 50 48.6 57.2 ...
 $ per      : num  1993 1993 1993 1993 1993 ...
 - attr(*, "breaks")=List of 3
 ..$ per: NULL
 ..$ age: NULL
 ..$ tfe: NULL
 - attr(*, "time.scales")= chr  "per" "age" "tfe"
 - attr(*, "time.since")= chr  "" "" ""
> with(iniA,table(allocation,lex.Cst))
      lex.Cst
allocation  None Auto D(None) D(Auto)
Conventional 155  0      0      0
Intensive    155  0      0      0
```

Then we can simulate the neuropathy development and death occurrence in a hypothetical cohort with the sex- and age-distribution as the entire steno 2 cohort.

```
> system.time( sA <- simLexis( Tr=trA, init=iniA, N=100, time.pts=seq(0,22,0.2) ) )
  user system elapsed
72.799  0.548  73.336
> save( sA, file="../data/sA.Rda" )

> load( file="../data/sA.Rda" )
> summary( sA )
Transitions:
  To
From  None  Auto D(None) D(Auto)  Records:  Events: Risk time:  Persons:
None 4267 19355  7378      0    31000    26733    270061.4    31000
Auto  0  8254      0  11101    19355    11101    220557.0    19355
Sum  4267 27609  7378  11101    50355    37834    490618.5    31000
```

In order to show how persons fare with respect to development of autonomous neuropathy and death we compute the state distribution by time since entry, starting at 2 years:

```
> prm <- c(1,2,4,3)
> pAi <- pState( nState( subset(sA,allocation=="Intensive"),
+                       at=seq(0,20,0.1), from=0, time.scale="tfe" ), prm )
> pAc <- pState( nState( subset(sA,allocation=="Conventional"),
+                       at=seq(0,20,0.1), from=0, time.scale="tfe" ), prm )
```

We can then plot the state distribution from 2 years, assuming the initial state distribution as the observed in the entire steno2 population:

```

> neust <-
+ function( Pi, Pc, rv=FALSE, leg=rep(15,4), xl=c(0,20) )
+ {
+ par( mfrow=c(1,2), mar=c(0,3,0,0), oma=c(3,0,2,3-2*rv), las=1 )
+ plot( Pi, col=clr[prm], xlim=xl, xlab="", # yaxt="n",
+       col.lab="transparent" )
+ lines( as.numeric(rownames(Pi)), Pi[,2], lwd=4 )
+ Pi <- cbind(0,Pi)
+ wr <- cbind( match(paste(leg),rownames(Pi)), 2:5 )
+ wl <- cbind( match(paste(leg),rownames(Pi)), 1:4 )
+ if(is.numeric(leg)) text( leg,
+                           (Pi[wl]+Pi[wr])/2,
+                           colnames(Pi)[-1],
+                           col=rep(c("white","black"),each=2) )
+ axis( side=2, at=1:10/10, labels=NA )
+ axis( side=4, at=0:5/5 )
+ axis( side=4, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Probability", side=2, line=2, las=0 )
+ mtext( "Intensive", side=3, line=1 )
+
+ plot( Pc, col=clr[prm], xlim=if(rv) xl[2:1] else xl,
+       xlab="", yaxt="n", col.lab="transparent" )
+ lines( as.numeric(rownames(Pc)), Pc[,2], lwd=4 )
+ axis( side=2+2*rv, at=1:10/10, labels=NA )
+ axis( side=4-2*rv, at=1:5/5, if(rv) labels=NA )
+ axis( side=4-2*rv, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4-2*rv, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4-2*rv, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Conventional", side=3, line=1 )
+ mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
+ }
> neust( pAi, pAc, rv=FALSE, xl=c(0,20) )

> neust( pAi, pAc, rv=TRUE, xl=c(0,20) )

```

5.3.6 Peripheral neuropathy

Then we fit a simple Cox-model for the progression of peripheral neuropathy with allocation as the only covariate and amend it by age and sex:

```

> summary( LxP )
Transitions:
  To
From  None Peri D(None) D(Peri)  Records:  Events: Risk time:  Persons:
None  264  74    53     0     391     127    1536.82    152
Peri   0 168     0     33     201     33     833.78     74
Sum   264 242     53    33     592     160    2370.59    152
> p1 <- coxph( Surv(tfe,tfe+lex.dur,lex.Xst=="Peri") ~ allocation, data = LxP)
> p2 <- update( p1, . ~ . + I((age-55)/10) + sex )
> round( ci.exp( p2, pval=TRUE ), 3 )

```

	exp(Est.)	2.5%	97.5%	P
allocationConventional	0.941	0.729	1.214	0.640
I((age - 55)/10)	0.795	0.663	0.954	0.014
sexMale	0.937	0.707	1.241	0.650

```

> HR <- rbind( ci.exp( p1, pval=TRUE ),
+             ci.exp( p2, pval=TRUE ) )
> round( HR[3:4,], 3 )

```

	exp(Est.)	2.5%	97.5%	P
I((age - 55)/10)	0.795	0.663	0.954	0.014
sexMale	0.937	0.707	1.241	0.650

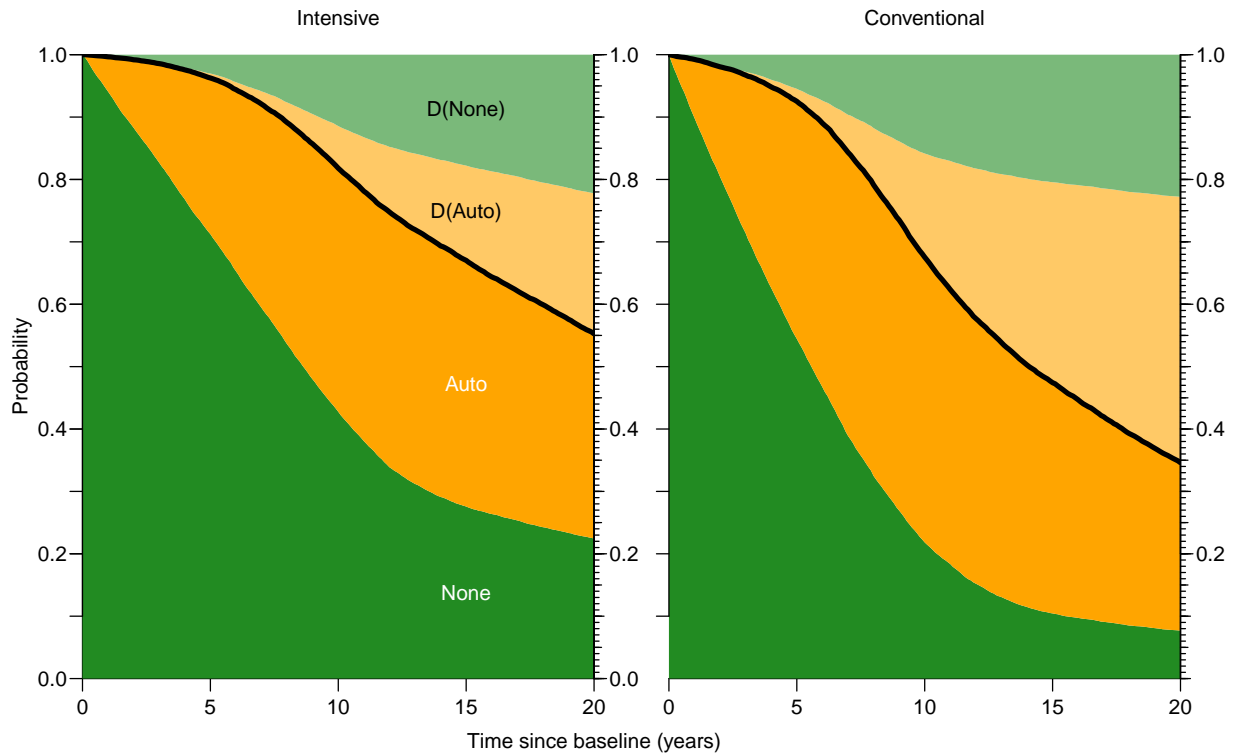


Figure 5.16: Probabilities of autonomous neuropathy, using the baseline distribution of age, sex and autonomous neuropathy for the entire Steno 2 cohort.

```
> HR <- cbind( 1/HR[,c(1,3,2)], HR )
> rownames( HR )[1:2] <- c("Unadj.", "Adj sex age")
> colnames( HR )[c(1,4)] <- c("Int vs Conv", "Conv vs Int")
> round( HR[1:2,], 3 )
```

	Int vs Conv	97.5%	2.5%	Conv vs Int	2.5%	97.5%	P
Unadj.	1.063	0.824	1.372	0.941	0.729	1.214	0.639
Adj sex age	1.063	0.824	1.371	0.941	0.729	1.214	0.640

So we see that there is no intervention effect on peripheral neuropathy. The model controlled for age and sex shows the same intervention effect (HR 1.02 (0.79,1.33)), but reveals that there is a significantly decreasing effect of age, HR=0.79 per 10 years of age (95% c.i. 0.65–0.95, p=0.010).

5.3.6.1 Prediction of burden

In order to predict the risk of peripheral neuropathy we fit parametric models for the transitions. So we split the follow-up in pieces of 1 month:

```
> SxP <- splitLexis( LxP, breaks=seq(0,25,1/12), time.scale="tfe" )
```

For convenience we use one set of knots for the mortalities, and one set for incidence of neuropathy:

```
> summary( LxP )
```

Transitions:

From	To	None	Peri	D(None)	D(Peri)	Records:	Events:	Risk time:	Persons:
None	None	264	74	53	0	391	127	1536.82	152
None	Peri	0	168	0	33	201	33	833.78	74
Peri	None	264	242	53	33	592	160	2370.59	152
Peri	Peri	0	168	0	33	201	33	833.78	74
Sum		264	242	53	33	592	160	2370.59	152

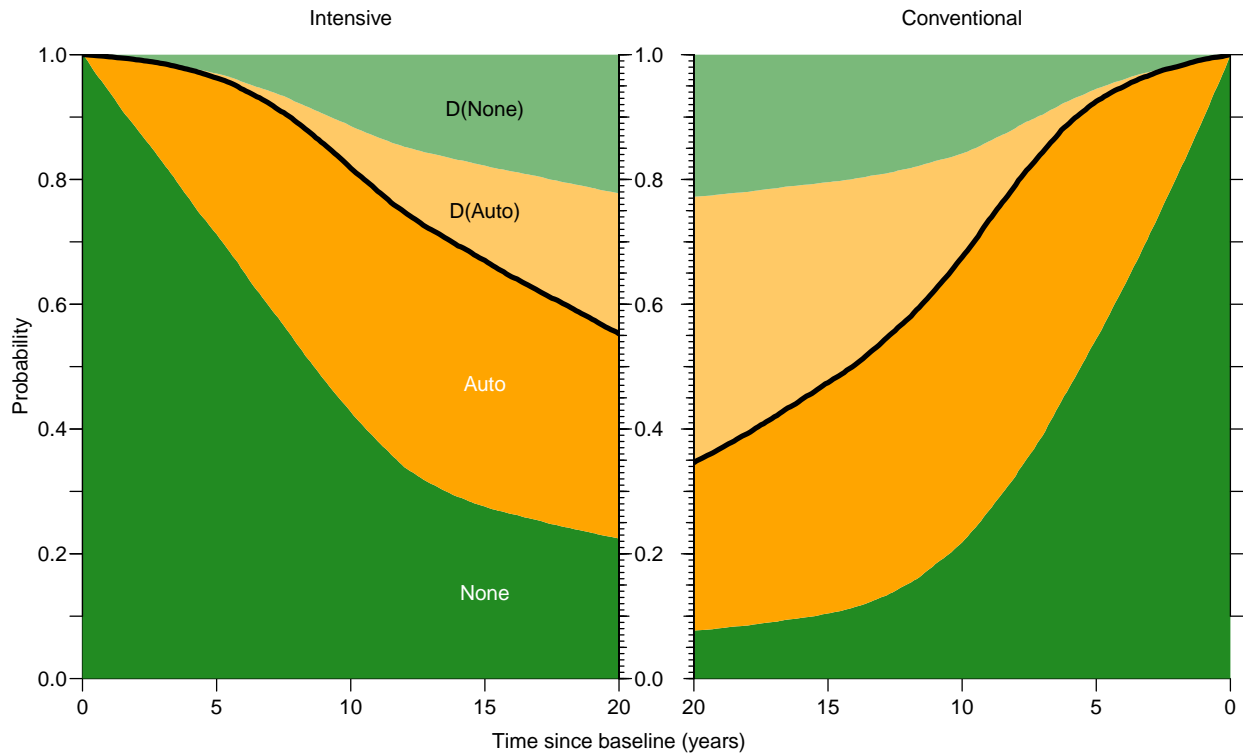


Figure 5.17: Probabilities of autonomous neuropathy, using the baseline distribution of age, sex and autonomous neuropathy for the entire Steno 2 cohort. This is the same plot as 5.16, except that the plot for the conventional group is mirrored.

```
> summary( SxP )
Transitions:
  To
From  None  Peri D(None) D(Peri)  Records:  Events: Risk time:  Persons:
None 18627   74    53      0    18754    127    1536.82    152
Peri   0 10177    0     33    10210    33     833.78     74
Sum  18627 10251   53    33    28964   160    2370.59    152

> ( d.kn <- with( subset( SxP, lex.Xst %in% c("D(None)","D(Peri)") ),
+               quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
      12.5%   37.5%   62.5%   87.5%
6.061602  9.202601 12.645106 16.907598

> ( n.kn <- with( subset( LxP, lex.Cst=="None" & lex.Xst=="Peri" ),
+               quantile( tfe+lex.dur, probs=(1:4-0.5)/4 ) ) )
      12.5%   37.5%   62.5%   87.5%
1.091433  3.637061  6.096798 10.651029
```

We fit models for mortality including age and sex and test for interaction between allocation and time since baseline:

```
> m1 <- glm( (lex.Xst %in% c("D(None)","D(Peri)")) ~
+           Ns( tfe, knots=d.kn ) + sex + I((age-tfe)/10) + allocation + lex.Cst,
+           offset=log(lex.dur),
+           family=poisson,
+           data = SxP )
> mi <- update( m1, . ~ . + allocation:lex.Cst )
> mn <- update( m1, . ~ . + allocation:tfe )
> anova( mn, m1, mi, test="Chisq")
```

Analysis of Deviance Table

```

Model 1: (lex.Xst %in% c("D(None)", "D(Peri)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst + allocation:tfe
Model 2: (lex.Xst %in% c("D(None)", "D(Peri)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst
Model 3: (lex.Xst %in% c("D(None)", "D(Peri)")) ~ Ns(tfe, knots = d.kn) +
sex + I((age - tfe)/10) + allocation + lex.Cst + allocation:lex.Cst
Resid. Df Resid. Dev Df Deviance Pr(>Chi)
1      28955      1091.5
2      28956      1091.5 -1 -0.020599  0.8859
3      28955      1091.3  1  0.237259  0.6262

```

```
> round( ci.exp( m1, pval=T ), 3 )
```

	exp(Est.)	2.5%	97.5%	P
(Intercept)	0.000	0.000	0.000	0.000
Ns(tfe, knots = d.kn)1	1.334	0.601	2.962	0.479
Ns(tfe, knots = d.kn)2	6.810	3.233	14.348	0.000
Ns(tfe, knots = d.kn)3	1.183	0.670	2.086	0.563
sexMale	1.385	0.828	2.315	0.214
I((age - tfe)/10)	2.712	1.910	3.852	0.000
allocationConventional	2.057	1.326	3.191	0.001
lex.CstPeri	0.916	0.582	1.441	0.704

We see that there is no sign of interaction between allocation and state (None/Peri) nor time since baseline — both p-values are larger than 0.5. Moreover, there is no difference in mortality between those with and without progression of peripheral neuropathy (HR 0.87 (0.55;1.38), P=0.557).

The occurrence of peripheral neuropathy is modelled likewise (this is essentially the same model as the Cox-model above, except that we model the baseline intensity parametrically):

```

> n1 <- glm( (lex.Xst == "Peri") ~
+           Ns( tfe, knots=d.kn) + sex + I(age-tfe) + allocation,
+           offset=log(lex.dur),
+           family=poisson,
+           data = subset(SxP, lex.Cst=="None" ) )
> ni <- update( n1, . ~ . + allocation:tfe )
> n2 <- update( ni, . ~ . + allocation:I(tfe^2) )
> aov <- anova( n1, ni, n2, test="Chisq")[-1,3:5]
> rownames( aov ) <- c("allo x time", "allo x t-sq")
> round( aov, 3 )

```

	Df	Deviance	Pr(>Chi)
allo x time	1	1.255	0.263
allo x t-sq	2	1.654	0.437

```
> round( ci.exp( n1, pval=T ), 3 )
```

	exp(Est.)	2.5%	97.5%	P
(Intercept)	0.168	0.027	1.054	0.057
Ns(tfe, knots = d.kn)1	0.289	0.073	1.153	0.079
Ns(tfe, knots = d.kn)2	0.341	0.151	0.771	0.010
Ns(tfe, knots = d.kn)3	0.333	0.133	0.833	0.019
sexMale	0.950	0.569	1.589	0.846
I(age - tfe)	0.983	0.951	1.016	0.301
allocationConventional	0.908	0.572	1.442	0.683

There are no significant interactions with time since baseline, so a proportional hazards model seems to be reasonable to apply in prediction.

We then update the HR collector for peripheral neuropathy

```

> nz <- update( n1, . ~ . - I(age-tfe) - sex )
> zz <- rbind( ci.exp( nz, subset="allo", pval=TRUE ),
+             ci.exp( n1, subset="allo", pval=TRUE ) )
> zz[,1:3] <- 1/zz[,c(1,3,2)]
> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][9]
[1] "Peripheral neuropathy"
> mainCI[9,,] <- zz
> round( ftable(mainCI,row.vars=1), 3 )

```

	model	raw			age/sex			P	
outcome	what	Estimate	2.5%	97.5%	P	Estimate	2.5%	97.5%	P
All cause mortality		0.551	0.364	0.835	0.005	0.526	0.346	0.800	0.003
CVD mortality		0.379	0.191	0.754	0.006	0.353	0.177	0.705	0.003
non-CVD mortality		0.703	0.413	1.197	0.195	0.686	0.401	1.172	0.168
Death or 1st CVD		NA	NA	NA	NA	NA	NA	NA	NA
Death CVD state		0.835	0.536	1.301	0.425	0.874	0.564	1.355	0.548
CVD event CVD state		0.552	0.394	0.772	0.001	0.575	0.412	0.803	0.001
Retinopathy progression		0.668	0.507	0.881	0.004	0.673	0.511	0.887	0.005
Autonomic neuropathy		0.594	0.397	0.890	0.011	0.573	0.382	0.859	0.007
Peripheral neuropathy		1.120	0.707	1.774	0.630	1.101	0.694	1.747	0.683
Macroalbuminuria		0.516	0.316	0.842	0.008	0.495	0.302	0.811	0.005

```

> save( mainCI, file="../data/mainCI.Rda" )

```

With these models fitted we shall show the probability of having peripheral neuropathy in the two groups, so we set up a transition object as well as a a baseline object from which to simulate.

```

> trP <- list( "None" = list( "Peri" = n1,
+                             "D(None)" = m1 ),
+             "Peri" = list( "D(Peri)" = m1 ) )
> lapply( trP, names )
$None
[1] "Peri" "D(None)"

$Peri
[1] "D(Peri)"

```

Since peripheral neuropathy is only measured as from the 3rd visit, we set the starting time to 4 years, but we use as initiating state for each person the status at the first measurement of peripheral neuropathy (which for a few persons is at a later visit than the 2nd). We use two copies of this dataset (that is the distribution of age, sex and neuropathy) one where we assume intensive treatment and one where we assume conventional:

```

> # selecting columns with "[" loses the attributes so this is not applicable:
> # iniP <- LxP[match(unique(LxP$lex.id),LxP$lex.id),c("sex","allocation","lex.Cst","tfe","age","per")]
> iniP <- subset( LxP, per==ave(per,lex.id,FUN=min),
+               select=c("sex","allocation","lex.Cst","tfe","age","per") )
> iniP$tfe = 0
> iniP <- rbind(iniP,iniP)
> iniP$allocation <- factor( rep( levels(iniP$allocation),
+                               each=nrow(iniP)/2 ) )
> str( iniP )
Classes 'Lexis' and 'data.frame': 304 obs. of 6 variables:
 $ sex      : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 1 2 2 2 ...
 $ allocation: Factor w/ 2 levels "Conventional",...: 2 2 2 2 2 2 2 2 2 2 ...
 $ lex.Cst  : Factor w/ 4 levels "None","Peri",...: 1 1 1 1 1 1 1 1 1 1 ...
 $ tfe      : num 0 0 0 0 0 0 0 0 0 0 ...
 $ age      : num 61.1 46.5 50 48.6 57.2 ...
 $ per      : num 1993 1993 1993 1993 1993 ...
 - attr(*, "breaks")=List of 3
 ..$ per: NULL

```

```

..$ age: NULL
..$ tfe: NULL
- attr(*, "time.scales")= chr "per" "age" "tfe"
- attr(*, "time.since")= chr "" "" ""
> with(iniP,table(allocation,lex.Cst))
      lex.Cst
allocation  None Peri D(None) D(Peri)
Conventional 152  0      0      0
Intensive    152  0      0      0

```

Then we can simulate the neuropathy development and death occurrence in a hypothetical cohort with the sex-, age- and neuropathy-distribution as the entire steno 2 cohort.

```

> system.time( sP <- simLexis( Tr=trP, init=iniP, N=100, time.pts=seq(0,22,0.2) ) )
  user system elapsed
62.831  0.336  63.157
> save( sP, file="../data/sP.Rda" )

```

```

> load( file="../data/sP.Rda" )
> summary( sP )
Transitions:
  To
From  None  Peri D(None) D(Peri)  Records:  Events: Risk time:  Persons:
None 4898 14945  10557      0    30400    25502  314147.6    30400
Peri  0    7737      0    7208    14945     7208  175101.0    14945
Sum  4898 22682  10557    7208    45345    32710  489248.6    30400

```

In order to show how persons fare with respect to development of autonomous neuropathy and death we compute the state distribution by time since entry, starting at 2 years:

```

> prm <- c(1,2,4,3)
> pPi <- pState( nState( subset(sP,allocation=="Intensive"),
+                       at=seq(0,20,0.1), from=0, time.scale="tfe" ), prm )
> pPc <- pState( nState( subset(sP,allocation=="Conventional"),
+                       at=seq(0,20,0.1), from=0, time.scale="tfe" ), prm )

```

We can then plot the state distribution from 2 years, assuming the initial state distribution as the observed in the entire steno2 population:

```

> neust( pPi, pPc, rv=FALSE, xl=c(0,20) )

> neust( pPi, pPc, rv=TRUE, xl=c(0,20) )

```

From figure 5.19 we see that the intensive group have a 20-year survival probability of 56.6% out of which $33.7 / 56.6 = 59.5\%$ have peripheral neuropathy, whereas the conventional group have a 20-year survival probability of 35.2% out of which $12.2 / 35.2 = 57.9\%$ have peripheral neuropathy.

So largely the same fraction of the survivors at 20 years in the two groups.

Finally, we save the state-distributions for autonomous and peripheral neuropathy

```

> save( pAi, pAc, pPi, pPc, file="../data/pNeu.Rda" )

```

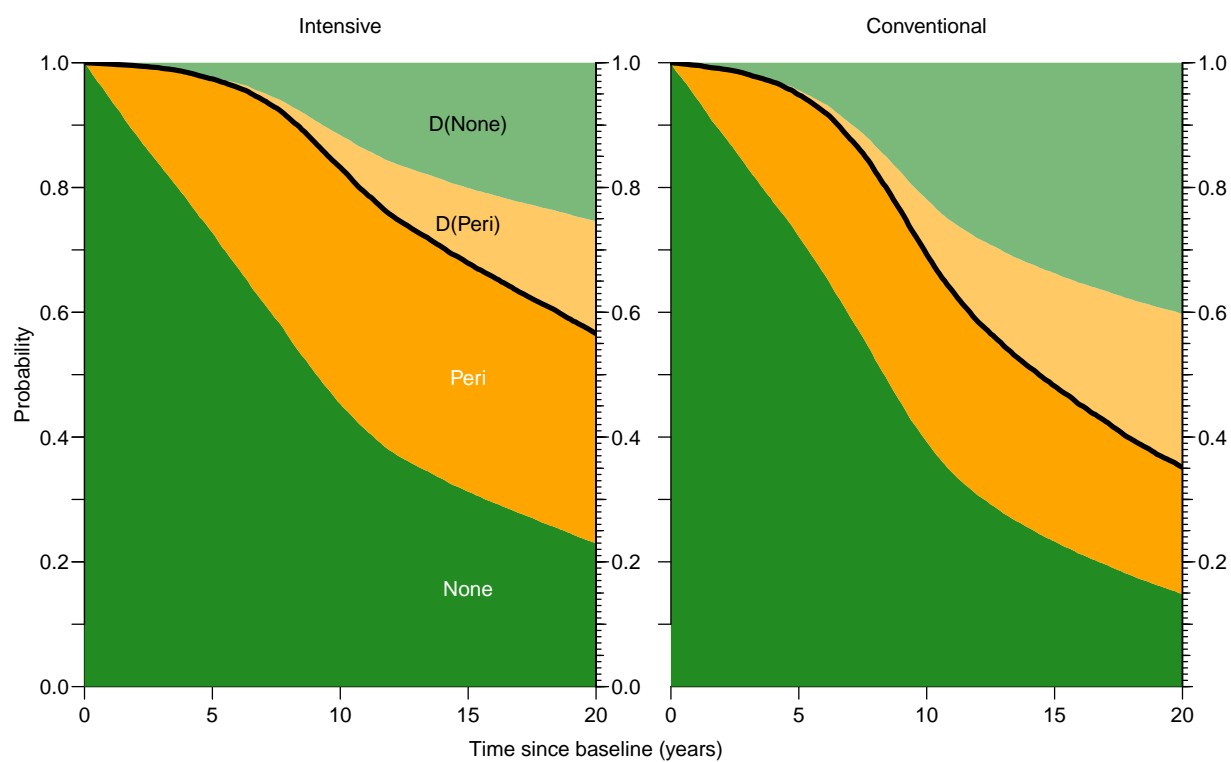


Figure 5.18: Probabilities of peripheral neuropathy, using the baseline distribution of age, sex and peripheral neuropathy for the entire Steno 2 cohort. Note that the timescale starts at 4 years after baseline.

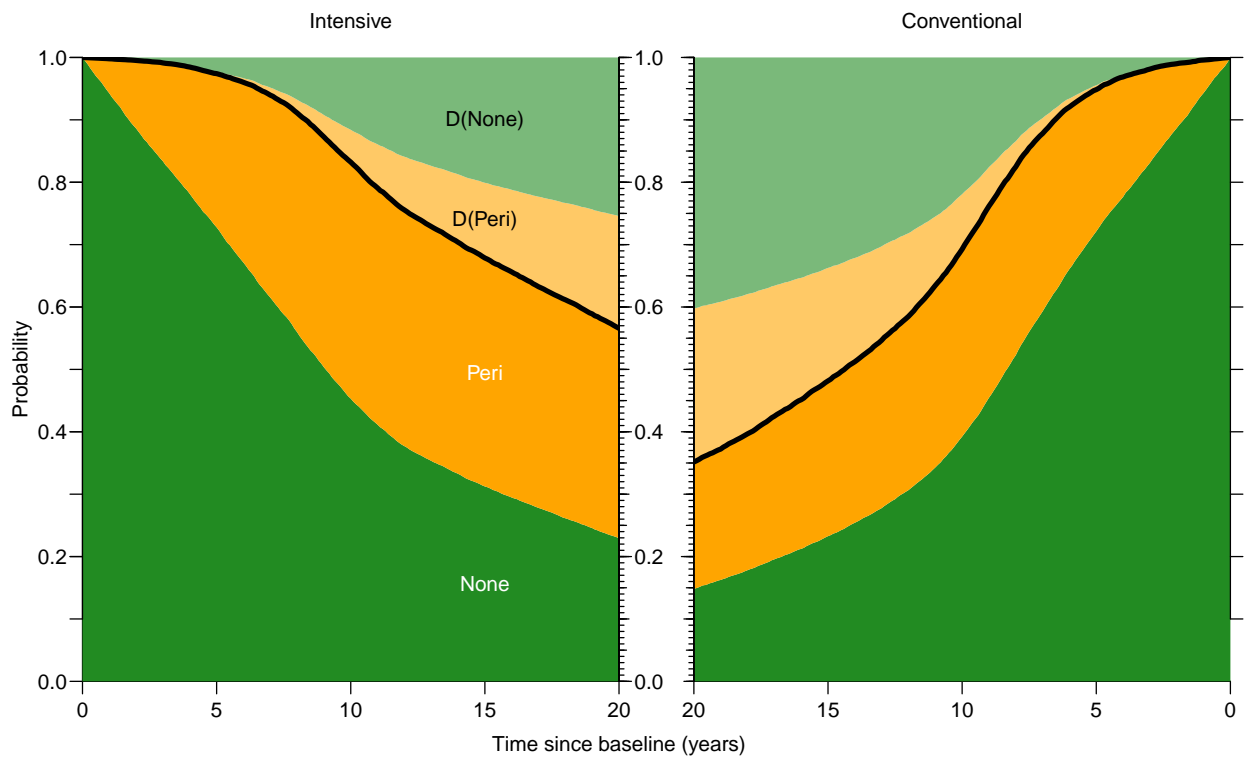


Figure 5.19: Probabilities of peripheral neuropathy, using the baseline distribution of age, sex and peripheral neuropathy for the entire Steno 2 cohort. Note that the timescale starts at 4 years after baseline. This is the same plot as 5.18, except that the plot for the conventional group is mirrored.

5.4 Albuminuria

5.4.1 Albumin measurements

First we read the data:

```
> library( Epi )
> library( survival )
> clear()
> load( file="../data/mic.Rda" )
> names( mic )
 [1] "ID"          "allocation" "sex"          "doBth"        "doVis"        "doESRD"
 [7] "doEye"       "doPhC"      "doBlind"     "doEnd"        "doDth"        "visit"
[13] "aer_level"  "med_aer"    "eds_l"       "eds_r"        "autoprog"     "periprog"
```

We will need the time since first visit in order to get the variation between persons at baseline as a parameter:

```
> fv <- subset( mic, visit==1, select=c("ID","doVis") )
> names(fv)[2] <- "doBase"
> mic <- transform( merge( mic, fv, all=TRUE ),
+                   tfBase = doVis - doBase )
```

We then fit a model for the log of the median albumin measurement, allowing separate slopes in the two groups:

```
> library( lme4 )
> m1 <- lmer( log(med_aer) ~ allocation + allocation:tfBase + (tfBase|ID), data=mic )
> summary( m1 )
```

```
Linear mixed model fit by REML ['lmerMod']
Formula: log(med_aer) ~ allocation + allocation:tfBase + (tfBase | ID)
Data: mic
```

```
REML criterion at convergence: 2204
```

```
Scaled residuals:
```

```
   Min       1Q   Median       3Q      Max
-3.4592 -0.5665  0.0109  0.5468  3.2020
```

```
Random effects:
```

```
Groups   Name             Variance Std.Dev. Corr
ID       (Intercept)  0.338363 0.5817
         tfBase       0.008483 0.0921  0.27
Residual                0.766796 0.8757
```

```
Number of obs: 723, groups: ID, 160
```

```
Fixed effects:
```

```
              Estimate Std. Error t value
(Intercept)      4.14191    0.09189   45.07
allocationConventional  0.21263    0.13157    1.62
allocationIntensive:tfBase  0.02347    0.01419    1.65
allocationConventional:tfBase 0.05305    0.01634    3.25
```

```
Correlation of Fixed Effects:
```

```
      (Intr) allctC allI:B
allctnCnvnt -0.698
allctnInt:B -0.169  0.118
allctnCnv:B  0.000 -0.177  0.000
```

We can make a formal test of equality of the two groups:

```
> round( Wald( m1, subset="allo", ctr.mat=rbind(c(1,0,0),c(0,1,-1)) ), 3 )
Chisq d.f.      P
5.664 2.000 0.059
```


so in this model with a linear trend in $\log(\text{AER})$ there is a borderline significant difference between the groups:

```
> round( cbind( ci.exp( m1 ), 1/ci.exp( m1 ) ), 3 )
              exp(Est.)  2.5% 97.5% exp(Est.)  2.5% 97.5%
(Intercept)      62.923 52.552 75.341    0.016 0.019 0.013
allocationConventional  1.237 0.956 1.601    0.808 1.046 0.625
allocationIntensive:tfBase  1.024 0.996 1.053    0.977 1.004 0.950
allocationConventional:tfBase  1.054 1.021 1.089    0.948 0.979 0.918
```

This means that the *ratio* of the median albumin levels between intensive and conventional at baseline is 0.81, and since the random effect for ID refer to the baseline the standard deviation of this (0.582) corresponds to the median difference in log-albumin levels between two randomly chosen persons. Specifically, it must be multiplied by $\sqrt{2}z_{0.75}$ and subsequently exponentiated to give the median ratio between two randomly chosen persons:

```
> str( VarCorr( m1 ) )
List of 1
 $ ID: num [1:2, 1:2] 0.33836 0.01471 0.01471 0.00848
   ..- attr(*, "dimnames")=List of 2
   .. ..$ : chr [1:2] "(Intercept)" "tfBase"
   .. ..$ : chr [1:2] "(Intercept)" "tfBase"
   ..- attr(*, "stddev")= Named num [1:2] 0.5817 0.0921
   .. ..- attr(*, "names")= chr [1:2] "(Intercept)" "tfBase"
   ..- attr(*, "correlation")= num [1:2, 1:2] 1 0.275 0.275 1
   .. ..- attr(*, "dimnames")=List of 2
   .. .. ..$ : chr [1:2] "(Intercept)" "tfBase"
   .. .. ..$ : chr [1:2] "(Intercept)" "tfBase"
   - attr(*, "sc")= num 0.876
   - attr(*, "useSc")= logi TRUE
   - attr(*, "class")= chr "VarCorr.merMod"
> ( btw <- exp( sqrt(2)*qnorm(0.75)*attr(VarCorr(m1)$ID,"stddev")[1] ) )
(Intercept)
 1.741693
```

The number $\backslash\text{Sexpr}[]\{(\text{round}(\text{btw},2))$ describes the between-person variation in the entire study population at baseline, so the baseline difference in albumin levels is quite small relative to the population variation — which of course is what should be expected in a clinical trial.

The estimated ratio of slopes in the two groups are:

```
> rt <- ci.exp( m1, subset="tfB", ctr.mat=rbind(c(-1,1)), pval=T )
> round( cbind( rt, 1/rt[, -4, drop=F] ), 3 )
              exp(Est.)  2.5% 97.5%    P exp(Est.)  2.5% 97.5%
[1,]          1.03 0.987 1.075 0.172    0.971 1.013 0.931
```

As before the between-person-variation in slopes can be extracted:

```
> exp( sqrt(2) * qnorm(0.75) * attr(VarCorr(m1)$ID,"stddev")[2] )
tfBase
1.091831
```

so the median ratio in slopes between two randomly chosen persons is 1.09; substantially larger than the estimated ratio of the mean slopes.

Thus we conclude that there is little difference in the levels and in the rate of change in albumin levels between the intensive and conventional arm. If we want to assess the ratio of the AER levels between intensive and conventional groups at different times after baseline, we compute these by `ci.exp`; and we compute the between person variation by pre-post multiplying the contrast matrix:

```

> dim( VarCorr(m1) )
NULL
> sqrt( diag(VarCorr(m1)$ID) )
(Intercept)      tfBase
 0.58168974  0.09210464
> t.pt <- 0:20
> CM <- cbind( -1, t.pt, -t.pt )
> MC <- -CM[,-2]
> vv <- sqrt(diag(MC %*% VarCorr(m1)$ID %*% t(MC)))
> vv <- exp( sqrt(2) * qnorm(0.75) * vv )
> zz <- ci.exp( m1, subset="allocation", ctr.mat=CM, pval=T )
> matplot( t.pt, zz[,-4], log="y", ylim=c(0.15,1.5),
+         type="l", col="black", lty=1, lwd=c(3,1,1), las=1,
+         xlab="Time since baseline (years)",
+         ylab="AER ratio, Intensive vs Conventional" )
> axis( side=2, at=c(0.15,2:15/10), labels=NA )
> polygon( c(rev(t.pt),t.pt),
+         c(rep(1,length(t.pt)),1/vv),
+         col=rgb(180,180,50,max=255), border="transparent" )
> abline( h=1, lty=3 )

```

From figure 5.20 we see that there is a borderline significant improvement in the mean AER-rate, but also that this is small compared to the between person variation in AER rates. The AER-ratio between the intensive and the conventional group changes from 0.81 at baseline to 0.45 at 20 years.

5.4.1.1 0-intercept models

It might be more realistic, since we have a randomized study, to assume a priori that the ratio of AER between groups is 1 at baseline, and then allow for a curved relationship in the ratio:

```

> m1 <- lmer( log(med_aer) ~ allocation:tfBase +
+           (tfBase|ID), data=mic )
> m2 <- lmer( log(med_aer) ~ allocation:tfBase + allocation:I(tfBase^2) +
+           (tfBase|ID), data=mic )
> summary( m1 )
Linear mixed model fit by REML ['lmerMod']
Formula: log(med_aer) ~ allocation:tfBase + (tfBase | ID)
Data: mic

REML criterion at convergence: 2204.4

Scaled residuals:
   Min       1Q   Median       3Q      Max
-3.4576 -0.5560  0.0277  0.5477  3.2420

Random effects:
 Groups   Name                Variance Std.Dev. Corr
 ID      (Intercept)  0.347706 0.58967
         tfBase      0.008664 0.09308  0.25
Residual                0.764292 0.87424
Number of obs: 723, groups: ID, 160

Fixed effects:
              Estimate Std. Error t value
(Intercept)      4.24577    0.06618   64.16
allocationIntensive:tfBase  0.02053    0.01418    1.45
allocationConventional:tfBase 0.05789    0.01616   3.58

Correlation of Fixed Effects:
              (Intr) allI:B
allctnInt:B -0.125
allctnCnv:B -0.178  0.022

```

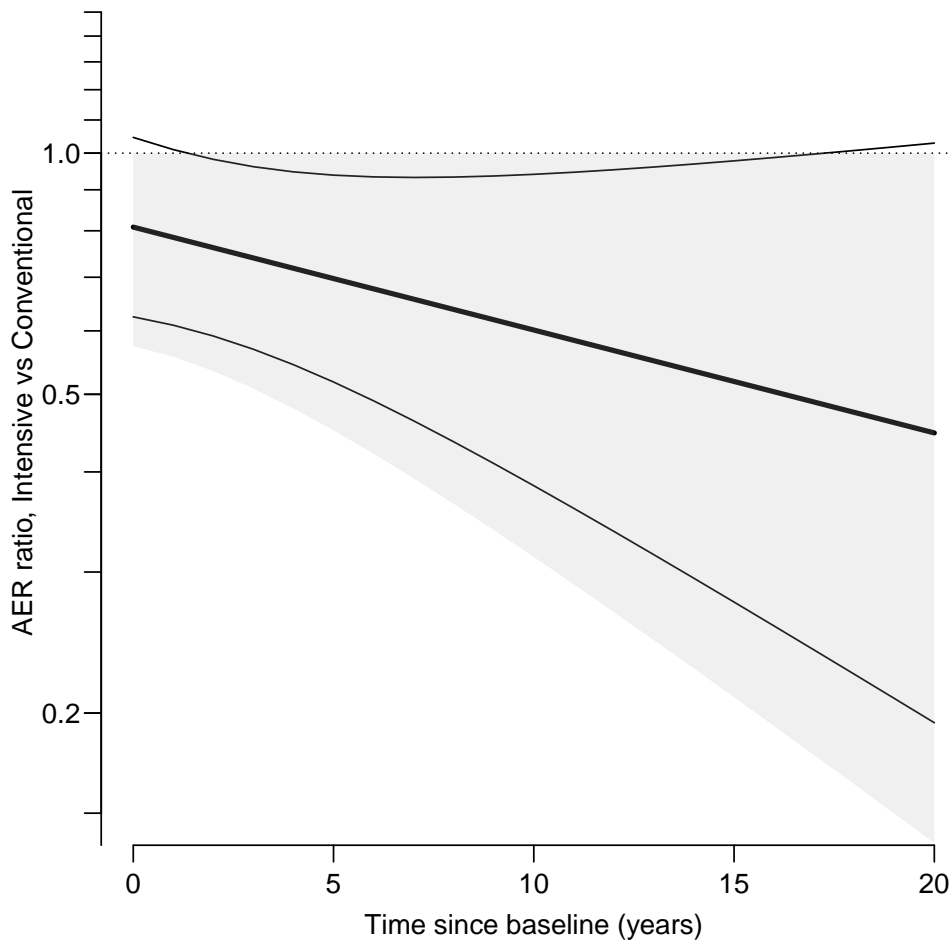


Figure 5.20: *Estimated mean AER-ratio between intensive and conventional by time since baseline. The gray area represents the between-person-variation: The height of the area is the median ratio of AER between two randomly selected persons from the same allocation group.*

```
> summary( mq )
```

```
Linear mixed model fit by REML ['lmerMod']
```

```
Formula: log(med_aer) ~ allocation:tfBase + allocation:I(tfBase^2) + (tfBase | ID)
```

```
Data: mic
```

```
REML criterion at convergence: 2206.4
```

```
Scaled residuals:
```

```
   Min      1Q  Median      3Q      Max
-3.5544 -0.5387 -0.0281  0.5169  3.2626
```

```
Random effects:
```

```
Groups   Name      Variance Std.Dev. Corr
ID       (Intercept) 0.337027 0.58054
         tfBase     0.009721 0.09859 0.25
```

```
Residual          0.730010 0.85441
```

```
Number of obs: 723, groups: ID, 160
```

```
Fixed effects:
```

```
(Intercept)                Estimate Std. Error t value
                        4.343379    0.071214    60.99
```

```

allocationIntensive:tfBase      -0.071465    0.024769   -2.89
allocationConventional:tfBase    0.049159    0.027528    1.79
allocationIntensive:I(tfBase^2)  0.005491    0.001190    4.61
allocationConventional:I(tfBase^2) 0.000420    0.001577    0.27

```

```

Correlation of Fixed Effects:
      (Intr) allI:B allC:B aI:I(B
allctnInt:B -0.320
allctnCnv:B -0.336  0.107
allI:I(B^2)  0.311 -0.807 -0.105
allC:I(B^2)  0.292 -0.093 -0.799  0.091

```

In the model with linear trend in log-AER the annual changes and their difference are:

```

> ( CC <- rbind( Int = c(1,0),
+               Conv = c(0,1),
+               "I-C" = c(1,-1) ) )
      [,1] [,2]
Int      1    0
Conv     0    1
I-C      1   -1
> ci.exp( ml, subset="tfB" )
      exp(Est.)      2.5%      97.5%
allocationIntensive:tfBase  1.020743 0.9927635 1.049510
allocationConventional:tfBase 1.059602 1.0265692 1.093698
> aer.l <- ci.exp( ml, subset="tfB", ctr.mat=CC, pval=TRUE )
> cbind( round( (aer.l[,-4]-1)*100, 1 ),
+        round( aer.l, 3 ) )
      exp(Est.) 2.5% 97.5% exp(Est.) 2.5% 97.5% P
Int      2.1 -0.7  5.0   1.021 0.993 1.050 0.148
Conv     6.0  2.7  9.4   1.060 1.027 1.094 0.000
I-C     -3.7 -7.6  0.4   0.963 0.924 1.004 0.079

```

Thus we see that there is a borderline significantly smaller decrease in the intervention group — 4% less per year,

A formal test of equality of the two groups in the liner 0-intercept model is:

```

> round( Wald( ml, subset="allocation", ctr.mat=rbind(c(-1,1)) ), 3 )
Chisq d.f. P
3.089 1.000 0.079

```

— so there is some (weak) evidence for a difference between the groups. We can compute the estimated ratio of means in the two groups as for the intercept model:

```

> CM <- cbind( t.pt, -t.pt )
> round( ll <- ci.exp( ml, subset="allocation", ctr.mat=CM, pval=T ), 3 )
      exp(Est.) 2.5% 97.5% P
[1,] 1.000 1.000 1.000 NaN
[2,] 0.963 0.924 1.004 0.079
[3,] 0.928 0.854 1.009 0.079
[4,] 0.894 0.789 1.013 0.079
[5,] 0.861 0.729 1.017 0.079
[6,] 0.830 0.674 1.022 0.079
[7,] 0.799 0.622 1.026 0.079
[8,] 0.770 0.575 1.031 0.079
[9,] 0.742 0.531 1.035 0.079
[10,] 0.714 0.491 1.040 0.079
[11,] 0.688 0.454 1.044 0.079
[12,] 0.663 0.419 1.048 0.079
[13,] 0.639 0.387 1.053 0.079
[14,] 0.615 0.358 1.058 0.079
[15,] 0.593 0.331 1.062 0.079
[16,] 0.571 0.306 1.067 0.079

```

```
[17,] 0.550 0.282 1.071 0.079
[18,] 0.530 0.261 1.076 0.079
[19,] 0.510 0.241 1.081 0.079
[20,] 0.492 0.223 1.085 0.079
[21,] 0.474 0.206 1.090 0.079
```

We can make an approximate Wald test for the quadratic extension (ignoring the random part of the model)

```
> round( ci.lin( mq, subset="2" ), 3 )
              Estimate StdErr      z      P    2.5% 97.5%
allocationIntensive:I(tfBase^2)    0.005  0.001  4.613 0.00  0.003 0.008
allocationConventional:I(tfBase^2)  0.000  0.002  0.266 0.79 -0.003 0.004
> round( Wald( mq, subset="2", ctr.mat=diag(2) ), 3 )
      Chisq  d.f.      P
21.303  2.000  0.000
```

A formal test of equality of the two groups in the quadratic model is:

```
> round( Wald( mq, subset="allocation", ctr.mat=rbind(c(-1,1,0,0),c(0,0,-1,1)) ), 3 )
      Chisq  d.f.      P
11.880  2.000  0.003
```

We can then compute the ratio of averages between the two groups in the quadratic model and plot this together with the ratio from the 0-intercept linear model:

```
> ( sqrt( diag( vc <- VarCorr(mq)$ID ) ) )
(Intercept)      tfBase
  0.5805400    0.0985947
> qv <- exp( sqrt(2) * qnorm(0.75) * sqrt(diag(MC %%% vc %%% t(MC))) ) )
> ci.exp( mq )
              exp(Est.)      2.5%      97.5%
(Intercept)  76.9671487  66.9403723  88.4958027
allocationIntensive:tfBase  0.9310284  0.8869107  0.9773407
allocationConventional:tfBase  1.0503877  0.9952176  1.1086160
allocationIntensive:I(tfBase^2)  1.0055059  1.0031628  1.0078543
allocationConventional:I(tfBase^2)  1.0004201  0.9973317  1.0035180
> CM <- cbind( t.pt, -t.pt, t.pt^2, -(t.pt^2) )
> qq <- ci.exp( mq, subset="allocation", ctr.mat=CM, pval=T )
> matplot( t.pt, cbind( qq[, -4], ll[, -4] ), log="y", ylim=c(0.15,1.5),
+         type="l", col=rep(c(gray(0.6),"black"),each=3), lty=1, lwd=c(4,1,1), las=1,
+         xlab="Time since baseline (years)",
+         ylab="AER ratio, Intensive vs Conventional" )
> axis( side=2, at=c(0.15,2:15/10), labels=NA )
> polygon( c(rev(t.pt),t.pt),
+         c(rep(1,length(t.pt)),1/qv),
+         col=rgb(180,180,180,50,max=255), border="transparent" )
> abline( h=1, lty=3 )
```

From figure 5.21 it is clear that the intervention effect is borderline significant; formally there is a significant difference between the groups ($p=0.003$), and the average ratio of AER levels between intensive and conventional allocation is about 0.4.

5.4.2 Normo-, micro- and macro-albuminuria

We load the micro-vascular data; that is records from each follow-up visit, as well as the functions needed to generate

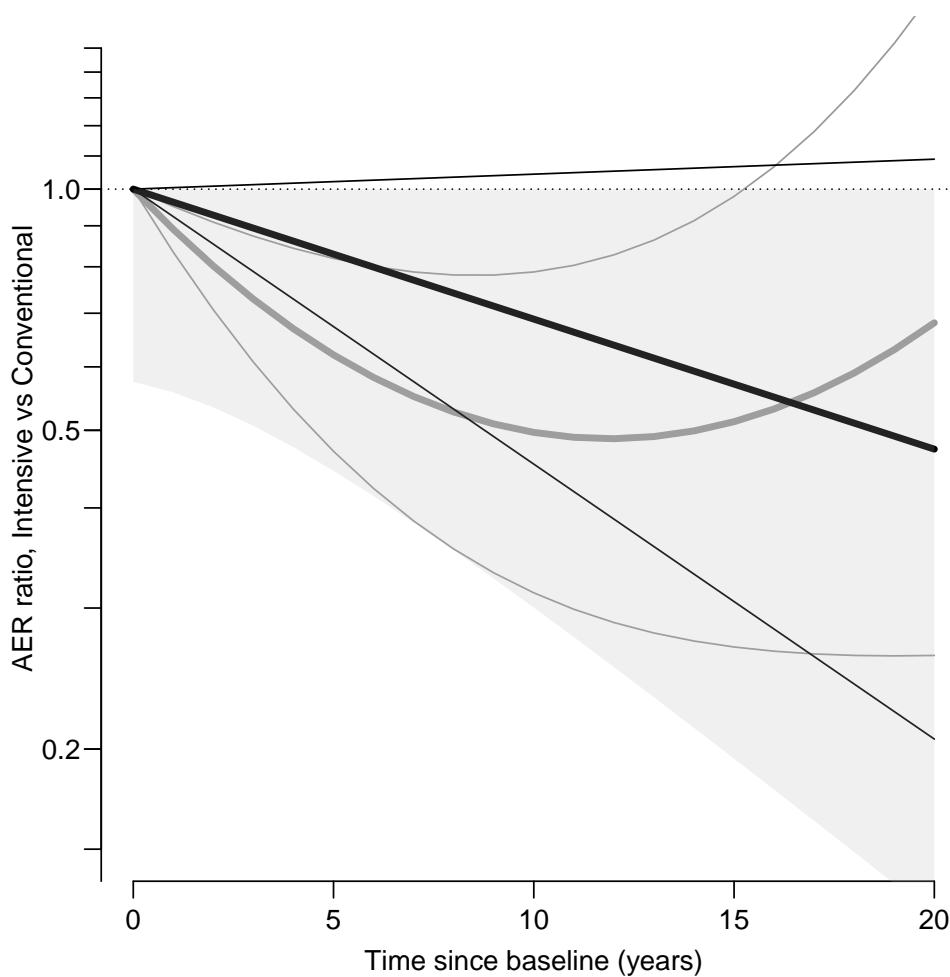


Figure 5.21: *Estimated mean AER-ratio between intensive and conventional by time since baseline by two different models; one with a linear trend and one with a quadratic trend, both assuming identical mean AER levels at baseline (black lines). Thick lines are the mean AER ratio; thin lines the 95% confidence intervals. The gray area represents the between-person-variation (from the quadratic model): The height of the area is the median ratio of AER between two randomly selected persons from the same allocation group.*

```
> library( Epi )
> clear()
> load( file="../data/mic.Rda" )
> source( "usefuns.R" )
```

We generate tables of states (`aer_level`) and dates of ascertainment of these, and generates a Lexis object with imputed follow-up times:

```
> levels( mic$aer_level )
[1] "Normoalbuminuria" "Microalbuminuria" "Macroalbuminuria"
> rt <- with( mic, tapply( as.integer(aer_level), list(ID,visit), mean ) )
> rd <- with( mic, tapply( doVis, list(ID,visit), mean ) )
> set.seed( 699817648 )
> alb <- make.dlex( rt, rd, incr=FALSE )
```

Since everyone is micro-albuminuric at start, we could either restrict transitions to be only increasing (only increase to macro), or we could allow transitions both ways. We have

chosen the latter to capture the possible beneficial effect of the intensive treatment. However we are formally making a mistake by assuming that transitions only occur in *one* direction between visits, which may not always be the case in reality.

```
> levels( alb )
[1] "1"      "2"      "3"      "Dead(1)" "Dead(2)" "Dead(3)"
> levels( alb$lex.Cst ) <-
+ levels( alb$lex.Xst ) <- c("Normo", "Micro", "Macro", "D(Norm)", "D(Mic)", "D(Mac)")
> summary( alb )
```

Transitions:

From	To	Normo	Micro	Macro	D(Norm)	D(Mic)	D(Mac)	Records:	Events:	Risk time:	Persons:
Normo		154	36	0	21	0	0	211	57	572.90	69
Micro		75	336	70	0	40	0	521	185	1419.29	160
Macro		0	23	103	0	0	32	158	55	425.66	64
Sum		229	395	173	21	40	32	890	297	2417.85	160

Here we plot the states, and the transitions between them:

```
> clr <- c("forestgreen", heat.colors(5)[c(3,1)])
> clr <- c(clr, rgb(t(col2rgb(clr)*0.6+255*0.4), max=255))
> parr <- rep(0.6, 7)
> parr[c(2,5,7)] <- 0.25
> par( mfrow=c(1,2), oma=c(0,0,0,0) )
> for( lv in levels(alb$allocation) )
+ {
+ boxes( subset(alb, allocation==lv),
+       boxpos=list(x=rep(c(20,80), each=3)+6-12*(lv=="Intensive"),
+                     y=rep(seq(85,5,,3), 2)),
+       show.BE="noz", scale.R=100,
+       col.bg=clr, pos.arr=parr,
+       col.border=c(clr[1:3], rep("black", 3)),
+       col.txt=rep(c("white", "black"), each=3) )
+ text(10+5-10*(lv=="Intensive"), 98, lv, cex=1.5, font=2, adj=0)
+ }
```

The analysis of rates can be performed by a single Cox model simultaneously for all transitions. However, since we here (as opposed to what is the case for the retinopathy and neuropathy data) have multiple possible *exits* from some states, the analysis must be performed on the *stacked* version of the dataset:

```
> stalb <- stack( alb )
> str( stalb )
```

Classes 'stacked.Lexis' and 'data.frame': 2301 obs. of 16 variables:

```
$ lex.id      : num  1 1 1 1 2 2 6 6 6 7 ...
$ per        : num  1997 1997 2006 2007 1995 ...
$ lex.dur    : num  0.435 1.497 0.575 0.175 0.45 ...
$ lex.Cst    : Factor w/ 6 levels "Normo","Micro",...: 1 1 1 1 1 1 1 1 1 1 ...
$ lex.Xst    : Factor w/ 6 levels "Normo","Micro",...: 1 2 1 2 1 2 1 1 2 1 ...
$ lex.Tr     : Factor w/ 7 levels "Normo->Micro",...: 1 1 1 1 1 1 1 1 1 1 ...
$ lex.Fail   : logi  FALSE TRUE FALSE TRUE FALSE TRUE ...
$ sex       : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 2 2 2 1 ...
$ doBth     : num  1932 1932 1932 1932 1947 ...
$ doVis     : num  1993 1993 1993 1993 1993 ...
$ doBlind   : num  NA NA NA NA NA NA NA NA NA NA ...
$ doEnd     : num  2015 2015 2015 2015 2015 ...
$ doDth     : num  NA NA NA NA NA ...
$ allocation: Factor w/ 2 levels "Intensive","Conventional": 1 1 1 1 1 1 2 2 2 ...
$ age      : num  64.4 64.8 73.8 74.4 48.1 ...
$ tfe     : num  3.34 3.78 12.78 13.36 1.57 ...
- attr(*, "time.scales")= chr "per" "age" "tfe"
> with( stalb, ftable(table(lex.Tr, allocation, lex.Fail), col.vars=2:3) )
```

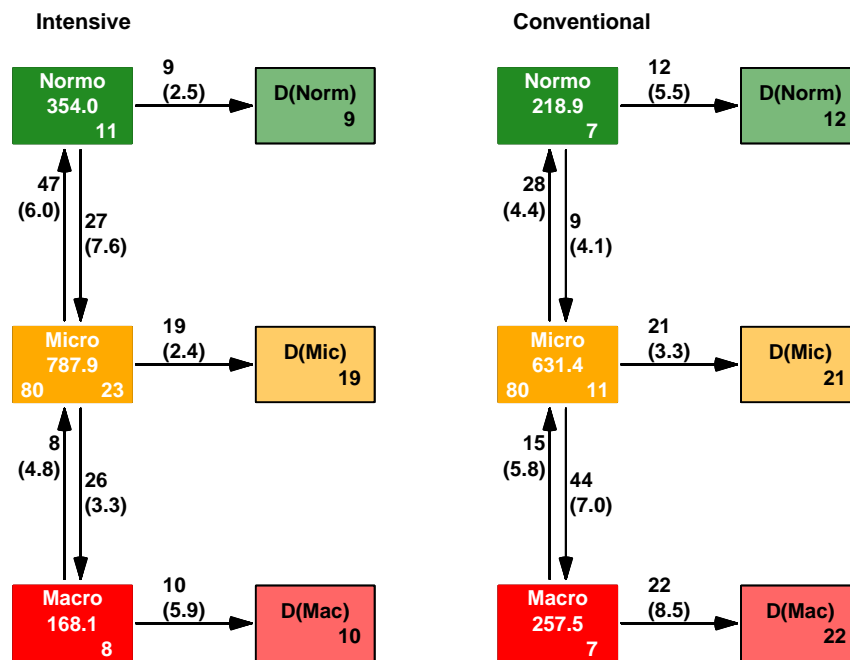


Figure 5.22: Transitions between states in the two treatment groups, allowing transitions to take place in both directions, but only in one direction between visits. The number in the center of the boxes is the person-years, the numbers at the bottom is the number of patients starting, resp. ending their follow-up in each box, and the numbers on the arrows are the number of transitions and the overall transition rates per 100 PY.

	allocation Intensive		Conventional	
lex.Tr	lex.Fail	FALSE TRUE	FALSE TRUE	
Normo->Micro		103 27	72 9	
Normo->D(Norm)		121 9	69 12	
Micro->Normo		227 47	219 28	
Micro->Macro		248 26	203 44	
Micro->D(Mic)		255 19	226 21	
Macro->Micro		51 8	84 15	
Macro->D(Mac)		49 10	77 22	

We then fit the Cox model, using a separate baseline as well as a separate intervention effect for each of the transitions. Note that we are using the time since entry as the timescale, and not correcting for age or sex.

```
> m1 <- coxph( Surv(tfe,tfe+lex.dur,lex.Fail) ~ strata(lex.Tr) + allocation:lex.Tr,
+ data = stalb )
> HR <- cbind( ci.exp( m1, pval=TRUE ),
+ 1/ci.exp( m1 ) ) [c(6,3,1,4,2,5,7), c(1:3,5,7,6,4)]
> colnames(HR)[c(1,4)] <- c("Int vs Conv", "Conv vs Int")
> rownames(HR) <- gsub("allocationIntensive:lex.Tr", "", rownames(HR))
> round( HR, 3 )
```

	Int vs Conv	2.5%	97.5%	Conv vs Int	97.5%	2.5%	P
Macro->Micro	0.967	0.407	2.297	1.034	0.435	2.457	0.939
Micro->Normo	1.612	1.007	2.579	0.620	0.388	0.993	0.047
Normo->Micro	1.806	0.848	3.843	0.554	0.260	1.179	0.125
Micro->Macro	0.514	0.315	0.840	1.944	1.190	3.175	0.008
Normo->D(Norm)	0.455	0.192	1.082	2.197	0.925	5.219	0.075
Micro->D(Mic)	0.640	0.341	1.202	1.562	0.832	2.933	0.166
Macro->D(Mac)	0.670	0.316	1.422	1.493	0.703	3.167	0.297

Here the first 3 columns refer to the intensive vs conventional, and the the last three to the opposite comparison. We see that the only formally significant effect of intervention is the halving of the transition from micro- to macro-albuminuria, HR 0.49 (0.30;0.79), $p=0.004$ — the other three transition rates do not differ significantly between groups, although there is a tendency that the intervention group also have higher transition rates between micro- and normo- albuminuria. The mortality rates are all roughly halved in the intensive group.

```
> Cests <- ci.exp( m1 )[c(6,3,1,4,2,5,7),c(1,3,2)]
> rownames( Cests ) <- gsub("allocationIntensive:lex.Tr","",rownames(Cests))
> plotEst( Cests, y=c(8:7+0.5,6:5,3:1), xlog=T, vref=1, grid=c(1:9/10,1:5),
+         xlab="Intensive vs. Conventional HR" )
```

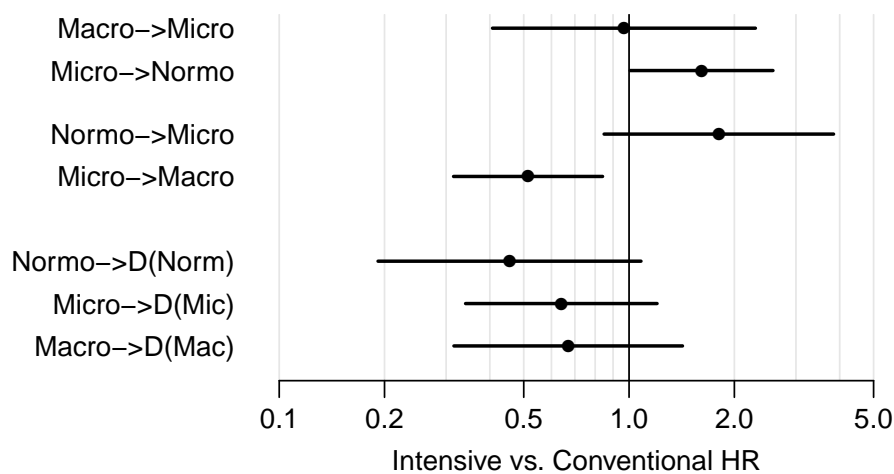


Figure 5.23: Hazard ratios between Intensive and Conventional for the 7 transitions in figure 5.22, based on separate Cox models with time since baseline as time-scale and intervention as the only covariate. The models thus assume proportional hazards between the two intervention groups for each transition.

However these numbers are just from a single imputation of the transition times, so should be taken with some caution.

5.4.3 Sensitivity analysis

In order to get a handle on the uncertainty introduced by the imputation of transition times we wrap up the entire imputation and analysis in a function that produces the estimates and their variance-covariance as result. Collecting these we can produce a summary estimate with corrected standard errors and moreover give an estimate of how large the contribution of the imputation variance is relative to the model.

Note that we only take the diagonal of the variance matrix, since it by the structure of the model *is* diagonal — the model fitting is tantamount to fitting each of the 7 transitions separately.

```
> alb1 <-
+ function()
+ {
+   alb <- make.dlex( rt, rd, incr=FALSE )
```

```

+ levels( alb$lex.Cst ) <-
+ levels( alb$lex.Xst ) <- c("Normo", "Micro", "Macro", "D(Norm)", "D(Mic)", "D(Mac)")
+ stalb <- stack( alb )
+ m1 <- coxph( Surv(tfe,tfe+lex.dur,lex.Fail) ~ strata(lex.Tr) + allocation:lex.Tr,
+           data = stalb )
+ zz <- ci.lin( m1 )[1:2]
+ zz
+   }
> system.time( res <- albi() )
  user system elapsed
 0.439  0.004  0.444

```

With this in place we set up an array to hold the results of analyses of data from multiple different imputations:

```

> nsim <- 400
> impalb <- NArray( c( list(imp=1:nsim), dimnames(res) ) )
> dimnames(impalb)[[2]] <- gsub("allocationConventional:lex.Tr", "", dimnames(impalb)[[2]])
> str( impalb )
logi [1:400, 1:7, 1:2] NA NA NA NA NA NA ...
- attr(*, "dimnames")=List of 3
..$ imp: chr [1:400] "1" "2" "3" "4" ...
..$   : chr [1:7] "allocationIntensive:lex.TrNormo->Micro" "allocationIntensive:lex.TrNormo->D(Norm)"
..$   : chr [1:2] "Estimate" "StdErr"
> system.time( for(i in 1:nsim) impalb[i,,] <- albi() )
  user system elapsed
193.046  0.059 193.480
> vv <- cbind( apply( impalb[, ,2] , 2, sd ),
+           apply( impalb[, ,1] , 2, var ),
+           apply( impalb[, ,2]^2, 2, mean ) )
> colnames( vv ) <- c("SD(sd)", "VAR(est)", "MEAN(var)")
> round( cbind(vv, "VAR(est)/MEAN(var)"=vv[,1]/vv[,3]), 4 )

```

	SD(sd)	VAR(est)	MEAN(var)	VAR(est)/MEAN(var)
allocationIntensive:lex.TrNormo->Micro	0.0003	0.0026	0.1487	0.0022
allocationIntensive:lex.TrNormo->D(Norm)	0.0004	0.0027	0.1953	0.0022
allocationIntensive:lex.TrMicro->Normo	0.0002	0.0004	0.0574	0.0034
allocationIntensive:lex.TrMicro->Macro	0.0005	0.0009	0.0624	0.0075
allocationIntensive:lex.TrMicro->D(Mic)	0.0009	0.0013	0.1031	0.0090
allocationIntensive:lex.TrMacro->Micro	0.0016	0.0061	0.1948	0.0083
allocationIntensive:lex.TrMacro->D(Mac)	0.0012	0.0076	0.1477	0.0083

Thus for all practical purposes it does not matter whether we use the average over multiple imputations or just a single imputation; the variance will be inflated by about 3% at most, and thus the s.e.s with $\sqrt{1.03} - 1 = 1.015 - 1 = 1.5\%$ at most. Moreover, the estimate based on a single imputation of time will with 99% probability be off by less than 5% of the standard error.

Thus we can safely rely on the results derived from the initial imputation, and we shall therefore just use this initially imputed transition times in further analyses.

5.4.4 Prediction of state occupancy

For the sake of illustration we would like to show the state occupancy, and in particular we would like to show how many die from different states — basically we want to see how many persons are in each of the states illustrated in figure 5.22 as a function of time since baseline.

If we want to illustrate how patients fare between the states we will need models for each of the 7 transitions; here we shall fit Poisson-models smoothing the trends and taking age and sex into account. The total number of events for the transitions vary from 20 to 70.

First we split the follow-up in 1-month intervals:

```
> summary( alb )
Transitions:
  To
From  Normo Micro Macro D(Norm) D(Mic) D(Mac)  Records:  Events: Risk time:  Persons:
  Normo  154   36    0      21     0     0      211      57    572.90     69
  Micro   75  336   70      0    40     0      521     185   1419.29    160
  Macro    0   23  103      0     0    32     158     55    425.66     64
  Sum    229  395  173     21    40    32     890    297   2417.85    160
> timeScales( alb )
[1] "per" "age" "tfe"
> spa <- splitLexis( alb, seq(0,24,1/12), "tfe" )
> summary( spa )
Transitions:
  To
From  Normo Micro Macro D(Norm) D(Mic) D(Mac)  Records:  Events: Risk time:  Persons:
  Normo 7024   36    0      21     0     0      7081     57    572.90     69
  Micro  75 17289   70      0    40     0     17474    185   1419.29    160
  Macro   0   23  5211      0     0    32     5266     55    425.66     64
  Sum   7099 17348  5281     21    40    32    29821    297   2417.85    160
```

Then we can fit models for each of the transitions, first we determine the position of the knots for the underlying splines:

```
> ( d.kn <- with( subset( spa, substr(lex.Xst,1,1)=="D" ),
+               c(0,quantile( tfe+lex.dur, probs=0:2/3)) ) )
      0% 33.33333% 66.66667%
0.000000 1.637235 8.276523 13.013005
> tkn <- addmargins( as.table( rbind(
+ ( mino.kn <- with( subset( spa, lex.Cst=="Micro" & lex.Xst=="Normo" ),
+                   c(0,quantile( tfe+lex.dur, probs=0:2/3)) ) ),
+ ( mima.kn <- with( subset( spa, lex.Cst=="Micro" & lex.Xst=="Macro" ),
+                   c(0,quantile( tfe+lex.dur, probs=0:2/3)) ) ),
+ ( nomi.kn <- with( subset( spa, lex.Cst=="Normo" & lex.Xst=="Micro" ),
+                   c(0,quantile( tfe+lex.dur, probs=0:2/3)) ) ),
+ ( mami.kn <- with( subset( spa, lex.Cst=="Macro" & lex.Xst=="Micro" ),
+                   c(0,quantile( tfe+lex.dur, probs=0:2/3)) ) ) ) ),
+               margin=1, FUN=mean )
> round( tkn, 2 )
      0% 33.33333% 66.66667%
A      0.00 0.01      1.70      4.60
B      0.00 0.18      2.79      6.82
C      0.00 2.18      6.39     11.28
D      0.00 1.68      4.55      8.65
mean   0.00 1.01      3.86      7.84
```

There is some variation of the quantiles of the events between the different transitions, but we use the same set of knots anyway — the mean:

```
> ( m.kn <- tkn["mean",] )
      0% 33.33333% 66.66667%
0.000000 1.011906 3.858181 7.839780
```

We can now fit the 5 different models; 4 for the transitions and one for the mortality rates. For the mortality rates we first check if the rates are actually proportional.

```
> ( dth <- levels(spa)[4:6] )
[1] "D(Norm)" "D(Mic)" "D(Mac)"
> m0 <- glm( ( lex.Xst %in% dth ) ~
+           Ns( tfe, knots=d.kn ) + allocation + lex.Cst + age + sex,
+           offset = log(lex.dur),
+           family=poisson,
+           data = spa )
> mi <- update( m0, . ~ . + Ns( tfe, knots=d.kn):lex.Cst )
> mI <- update( m0, . ~ . + Ns( tfe, knots=d.kn):allocation )
> miI <- update( mi, . ~ . + Ns( tfe, knots=d.kn):allocation )
> round( ci.exp( m0 ), 3 )
```

```

exp(Est.) 2.5% 97.5%
(Intercept) 0.000 0.000 0.000
Ns(tfe, knots = d.kn)1 6.719 1.361 33.173
Ns(tfe, knots = d.kn)2 43.131 0.408 4561.508
Ns(tfe, knots = d.kn)3 2.132 0.854 5.323
allocationConventional 1.800 1.181 2.744
lex.CstMicro 0.808 0.475 1.377
lex.CstMacro 1.341 0.763 2.355
age 1.107 1.070 1.145
sexMale 1.392 0.840 2.308
> round( ci.exp( mI ), 3 )

```

```

exp(Est.) 2.5% 97.5%
(Intercept) 0.000 0.000 0.000
Ns(tfe, knots = d.kn)1 2.271 0.309 16.668
Ns(tfe, knots = d.kn)2 13.786 0.040 4792.915
Ns(tfe, knots = d.kn)3 0.945 0.286 3.122
allocationConventional 0.309 0.002 38.358
lex.CstMicro 0.807 0.472 1.381
lex.CstMacro 1.343 0.762 2.367
age 1.108 1.071 1.146
sexMale 1.398 0.844 2.316
Ns(tfe, knots = d.kn)1:allocationConventional 8.366 0.296 236.626
Ns(tfe, knots = d.kn)2:allocationConventional 16.327 0.001 278783.818
Ns(tfe, knots = d.kn)3:allocationConventional 4.484 0.734 27.410
> AoV <- anova( miI, mi, m0, mI, miI, test="Chisq" )[-1,-(1:2)]
> rownames( AoV ) <- c("t*allo/t*st", "t*st", "t*allo", "t*st/t*allo")
> round( AoV, 3 )

```

```

Df Deviance Pr(>Chi)
t*allo|t*st -3 -2.551 0.466
t*st -6 -14.169 0.028
t*allo 3 3.119 0.374
t*st|t*allo 6 13.601 0.034

```

Thus it is pretty clear that we cannot assume proportional hazards of death between different levels of albuminuria, but equally clear that mortality is proportional between treatment groups, thus the model *mi* is the model to use. We will incorporate this in the mortality model, so we now only need the 4 models linking the different levels of albuminuria;

```

> mino <- glm( ( lex.Xst %in% "Normo" ) ~
+           Ns( tfe, knots=m.kn ) + allocation + age + sex,
+           offset = log(lex.dur),
+           family=poisson,
+           data = subset(spa,lex.Cst=="Micro" ) )
> nomi <- update( mino, ( lex.Xst %in% "Micro" ) ~ . , data = subset(spa,lex.Cst=="Normo" ) )
> mima <- update( mino, ( lex.Xst %in% "Macro" ) ~ . , data = subset(spa,lex.Cst=="Micro" ) )
> mami <- update( mino, ( lex.Xst %in% "Micro" ) ~ . , data = subset(spa,lex.Cst=="Macro" ) )

```

The models we fitted are proportional hazards models, so we briefly make formal tests for proportional hazards along the timescale:

```

> ph3 <- rbind(
+ anova( mino, update(mino, .~.+Ns(tfe,knots=m.kn):allocation), test="Chisq" )[2,-(1:2)],
+ anova( nomi, update(nomi, .~.+Ns(tfe,knots=m.kn):allocation,maxit=100), test="Chisq" )[2,-(1:2)],
+ anova( mima, update(mima, .~.+Ns(tfe,knots=m.kn):allocation), test="Chisq" )[2,-(1:2)],
+ anova( mami, update(mami, .~.+Ns(tfe,knots=m.kn):allocation), test="Chisq" )[2,-(1:2)])
> ph1 <- rbind(
+ anova( mino, update(mino, .~.+tfe:allocation), test="Chisq" )[2,-(1:2)],
+ anova( nomi, update(nomi, .~.+tfe:allocation,maxit=100), test="Chisq" )[2,-(1:2)],
+ anova( mima, update(mima, .~.+tfe:allocation), test="Chisq" )[2,-(1:2)],
+ anova( mami, update(mami, .~.+tfe:allocation), test="Chisq" )[2,-(1:2)])
> rownames( ph3 ) <- rownames( ph1 ) <- c("mino","nomi","mima","mami")
> round( cbind(ph3,ph1), 3 )

```

	Df	Deviance	Pr(>Chi)	Df	Deviance	Pr(>Chi)
mino	3	0.748	0.862	1	0.661	0.416
nomi	3	4.273	0.233	1	0.000	0.986
mima	3	5.758	0.124	1	0.864	0.353
mami	3	8.269	0.041	1	1.630	0.202

Thus we see that there is no compelling indication of non-proportional hazards for any of the transitions, neither with a 3 d.f. (except possibly for the transition macro to micro) nor a 1 d.f. alternative.

We then update the HR collector for peripheral neuropathy

```
> mima0 <- update( mima, . ~ . - age - sex )
> zz <- rbind( ci.exp( mima0, subset="allo", pval=TRUE ),
+             ci.exp( mima, subset="allo", pval=TRUE ) )
> zz[,1:3] <- 1/zz[,c(1,3,2)]
> load( file="../data/mainCI.Rda" )
> dimnames( mainCI )[[1]][10]
[1] "Macroalbuminuria"
> mainCI[10,,] <- zz
> round( ftable(mainCI,row.vars=1), 3 )
```

outcome	model				age/sex				P
	what	Estimate	2.5%	97.5%	P	Estimate	2.5%	97.5%	
All cause mortality		0.551	0.364	0.835	0.005	0.526	0.346	0.800	0.003
CVD mortality		0.379	0.191	0.754	0.006	0.353	0.177	0.705	0.003
non-CVD mortality		0.703	0.413	1.197	0.195	0.686	0.401	1.172	0.168
Death or 1st CVD		NA	NA	NA	NA	NA	NA	NA	NA
Death CVD state		0.835	0.536	1.301	0.425	0.874	0.564	1.355	0.548
CVD event CVD state		0.552	0.394	0.772	0.001	0.575	0.412	0.803	0.001
Retinopathy progression		0.668	0.507	0.881	0.004	0.673	0.511	0.887	0.005
Autonomic neuropathy		0.594	0.397	0.890	0.011	0.573	0.382	0.859	0.007
Peripheral neuropathy		1.120	0.707	1.774	0.630	1.101	0.694	1.747	0.683
Macroalbuminuria		0.516	0.316	0.842	0.008	0.495	0.302	0.811	0.005

```
> save( mainCI, file="../data/mainCI.Rda" )
```

We can then derive the HRs associated with treatment in the different models:

```
> Pests <- Cests*0
> Pests["Macro->Micro" ,] <- 1/ci.exp(mami,subset="allo")
> Pests["Micro->Normo" ,] <- 1/ci.exp(mino,subset="allo")
> Pests["Normo->Micro" ,] <- 1/ci.exp(nomi,subset="allo")
> Pests["Micro->Macro" ,] <- 1/ci.exp(mima,subset="allo")
> Pests["Normo->D(Norm)",] <- 1/ci.exp(mi,subset="allo")
> Pests["Micro->D(Mic)" ,] <- 1/ci.exp(mi,subset="allo")
> Pests["Macro->D(Mac)" ,] <- 1/ci.exp(mi,subset="allo")
```

and we can then plot these comparing with the raw ones from the Cox model

```
> par( mfrow=c(1,1), mar=c(3,3,1,1), mgp=c(3,1,0)/1.6 )
> plotEst( Pests, y=c(8:7+0.5,6:5,3:1)+0.1, xlog=T, vref=1, grid=c(1:9/10,1:5),
+         xlab="Intensive vs. Conventional HR" )
> linesEst( Cests, y=c(8:7+0.5,6:5,3:1)-0.1, xlog=T, col=gray(0.6) )
```

5.4.4.1 State occupancy

With models for all transitions in place we can now set up a transition object to be used in simulating from an initial data frame:

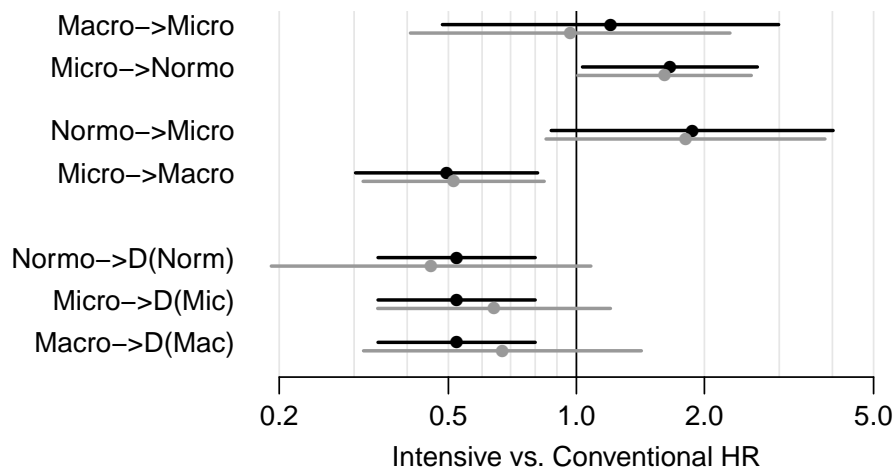


Figure 5.24: Hazard ratios between *Intensive* and *Conventional* for the 7 transitions in figure 5.22, based on Poisson models for transitions (black) based on separate models for transition between albuminuria states and a single model for the mortalities. Models use time since baseline as time-scale effects of intervention and intervention, age and sex as covariates. The gray bars are the estimates from the simple Cox-models.

```
> TR <- list( Normo = list( Micro = nomi,
+                          "D(Norm)" = mi),
+            Micro = list( Normo = mino,
+                          Macro = mima,
+                          "D(Mic)" = mi),
+            Macro = list( Micro = mami,
+                          "D(Mac)" = mi ) )
> lapply( TR, names )
$Normo
[1] "Micro" "D(Norm)"

$Micro
[1] "Normo" "Macro" "D(Mic)"

$Macro
[1] "Micro" "D(Mac)"
> ini <- subset( alb, tfe==0,
+               select=c(timeScales(alb),"lex.id","lex.Cst","allocation","sex") )
> ini <- rbind( cbind( ini, allocation="Intensive" ),
+               cbind( ini, allocation="Conventional" ) )
> ini$lex.Cst <- factor( ini$lex.Cst, levels=levels( ini$lex.Cst )[1:3] )
> levels( ini )
[1] "Normo" "Micro" "Macro"
> str( ini )
Classes 'Lexis' and 'data.frame': 320 obs. of 7 variables:
 $ per      : num 1993 1993 1993 1993 1993 ...
 $ age      : num 61.1 46.5 50 48.6 57.2 ...
 $ tfe      : num 0 0 0 0 0 0 0 0 0 0 ...
 $ lex.id   : num 1 2 3 4 5 6 7 8 9 10 ...
 $ lex.Cst  : Factor w/ 3 levels "Normo","Micro",...: 2 2 2 2 2 2 2 2 2 2 ...
 $ allocation: Factor w/ 2 levels "Intensive","Conventional": 1 1 2 2 2 2 2 1 1 1 ...
 $ sex      : Factor w/ 2 levels "Female","Male": 2 2 2 2 2 2 1 2 2 2 ...
 - attr(*, "breaks")=List of 3
 ..$ per: NULL
 ..$ age: NULL
 ..$ tfe: NULL
 - attr(*, "time.scales")= chr "per" "age" "tfe"
 - attr(*, "time.since")= chr "" "" ""
```

We can now simulate the follow-up through the states for a cohort 100 times bigger than the Steno2 study population:

```
> system.time( Salb <- simLexis( TR, ini, N=100, time.pts=0:100/4 ) )
  user system elapsed
130.483 11.415 142.130
> summary( Salb, by="allocation" )
$Intensive

Transitions:
  To
From  Normo Micro Macro D(Mic) D(Mac) D(Norm) Records: Events: Risk time: Persons:
  Normo 1774 5916 0 0 0 2034 9724 7950 79368.76 8555
  Micro 9724 4150 5676 4189 0 0 23739 19589 175520.39 16000
  Macro 0 1823 1224 0 2629 0 5676 4452 39385.69 5240
  Sum 11498 11889 6900 4189 2629 2034 39139 31991 294274.84 16000

$Conventional

Transitions:
  To
From  Normo Micro Macro D(Mic) D(Mac) D(Norm) Records: Events: Risk time: Persons:
  Normo 1252 2050 0 0 0 2357 5659 4407 53769.82 5428
  Micro 5659 1858 8689 4701 0 0 20907 19049 134062.58 16000
  Macro 0 2857 961 0 4871 0 8689 7728 53513.07 7690
  Sum 6911 6765 9650 4701 4871 2357 35255 31184 241345.47 16000
> save( Salb, file="../data/Salb.Rda" )

> load( file="../data/Salb.Rda" )
```

We then compute the number of persons in each state at 3-month intervals

```
> pm <- c(1:3,5,4,6)
> levels(Salb)[pm]
[1] "Normo" "Micro" "Macro" "D(Mac)" "D(Mic)" "D(Norm)"
> Pi <- pState( nState( subset(Salb,allocation=="Intensive"),
+ at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=pm )
> Pc <- pState( nState( subset(Salb,allocation=="Conventional"),
+ at=seq(0,20,1/4), from=0, time.scale="tfe" ), perm=pm )
> head( Pi )

      State
when  Normo  Micro  Macro  D(Mac) D(Mic) D(Norm)
0     0.000000 1.000000 1.000000 1.000000 1 1
0.25 0.0275625 0.9916875 0.9998750 0.9998750 1 1
0.5   0.0577500 0.9842500 0.9992500 0.9992500 1 1
0.75 0.0903125 0.9768750 0.9986250 0.9986250 1 1
1     0.1208125 0.9676250 0.9980000 0.9980000 1 1
1.25 0.1513125 0.9589375 0.9974375 0.9974375 1 1

> albst <- function(rv=FALSE,leg=15){
+ par( mfrow=c(1,2), mar=c(0,3,0,0), oma=c(3,0,3,3-2*rv), las=1 )
+ plot( Pi, col=clr[c(1:3,6:4)], xlim=c(0,20), xlab="", yaxt="n",
+ col.lab="transparent" )
+ lines( as.numeric(rownames(Pi)), Pi[, "Macro"], lwd=4 )
+ if(is.numeric(leg)) text( rep(leg,6),
+ Pi[paste(leg),]-diff(c(0,Pi[paste(leg),]))/2,
+ colnames(Pi),
+ col=rep(gray(1:0),each=3) )
+ axis( side=4, at=0:5/5 )
+ axis( side=4, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4, at=0:100 /100, tcl=-0.2, labels=NA )
```

```

+ mtext( "Probability", side=2, line=1, las=0 )
+ mtext( "Intensive", side=3, line=1 )
+
+ plot( Pc, col=clr[c(1:3,6:4)], xlim=if(rv) c(20,0) else c(0,20),
+       xlab="", yaxt="n", col.lab="transparent" )
+ lines( as.numeric(rownames(Pc)), Pc[, "Macro"], lwd=4 )
+ axis( side=4-2*rv, at=1:5/5, if(rv) labels=NA )
+ axis( side=4-2*rv, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4-2*rv, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4-2*rv, at=0:100 /100, tcl=-0.2, labels=NA )
+ mtext( "Conventional", side=3, line=1 )
+ mtext( "Time since baseline (years)", side=1, line=2, outer=TRUE )
+ }
> albst(rv=FALSE,leg=18)

> albst(rv=TRUE,leg=18)

```

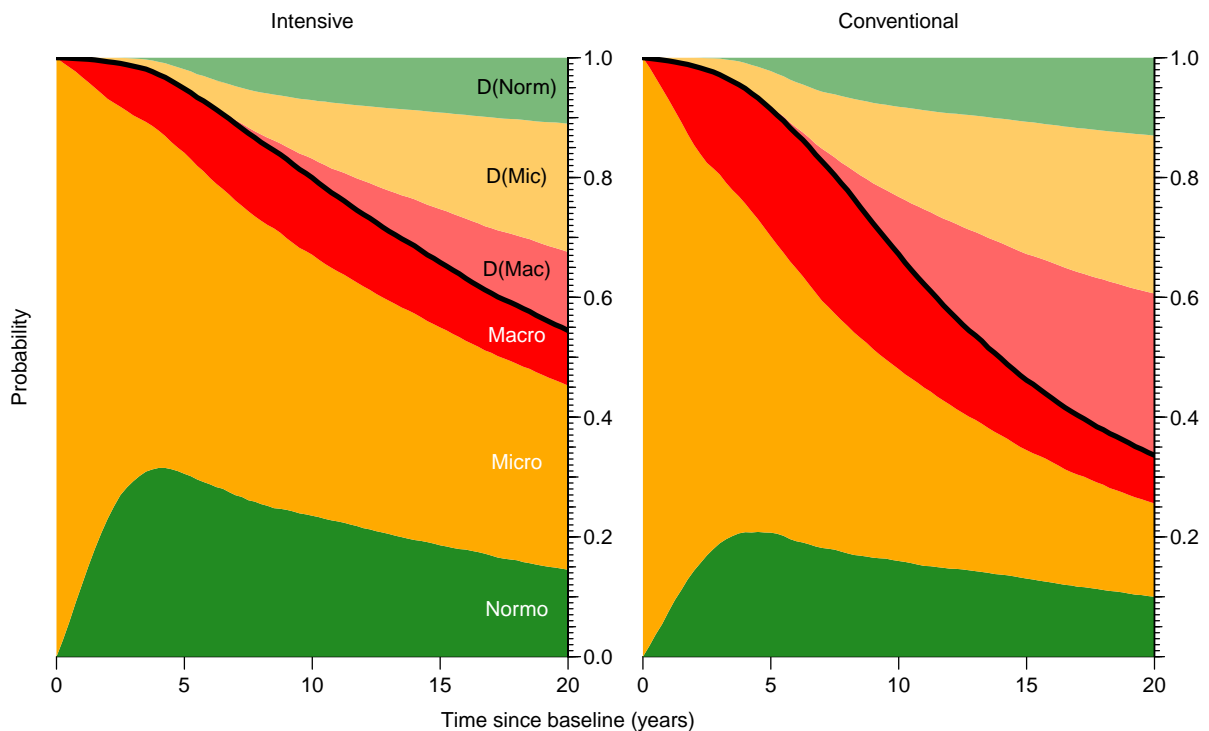


Figure 5.25: Probability of being in each state (from bottom: Normo-, Micro- and Macro-albuminuria) as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

The figures 5.25 and 5.26 show the predicted distribution of patients in Normo-, Micro- and Macro-albuminuria using assumptions that transitions occur at a simulated time between visits where status were assessed. Since we are modeling mortality conditional on albuminuria state, we see survival curves that are slightly different from the ones seen in the overall mortality analysis.

Finally we save the state probability results:

```
> save( Pi, Pc, file="../data/pAlb.Rda" )
```

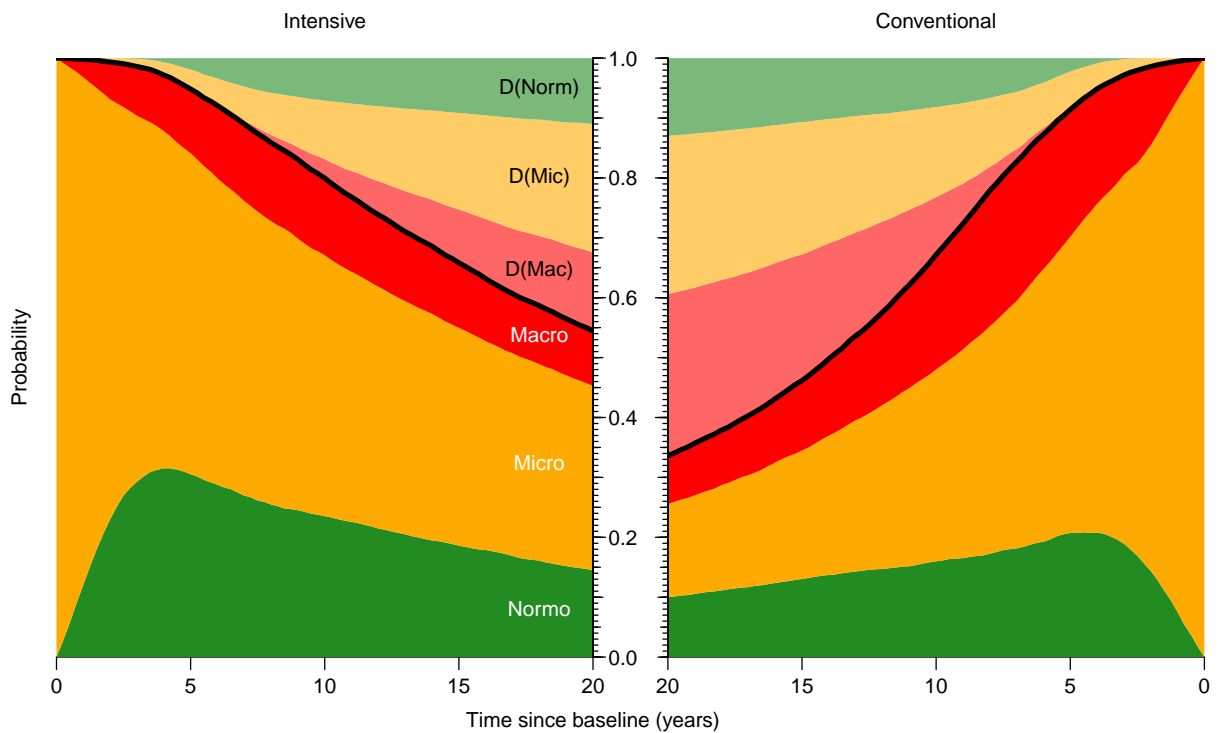



Figure 5.26: Probability of being in each state (from bottom: Normo-, Micro- and Macro-albuminuria) as a function of time since baseline. The black curve is the estimated survival curve, and the pale colors in the dead states represent the states from which person have died.

This is the same figure as 5.25, except that the x-axis of the right-hand panel is reversed.

5.5 Comparative plotting of microvascular complications

The relevant `pState` objects are:

- Autonomic neuropathy: `pAi`, `pAc`
- Peripheral neuropathy: `pPi`, `pPc`
- Retinopathy: `Ei`, `Ec` — the objects from models that only allows progression of retinopathy, using the empirical baseline distribution as starting point.

The objects `rEi`, `rEc` are from the models that allow improvement of retinopathy, but they are likely to be shaky because of the small number of improvement transitions.

- Albuminuria: `Pi`, `Pc`

First we devise a function that plots state distribution from the two groups together

```
> mk.clr <-
+ function( clr )
+ {
+   clr <- c("forestgreen",clr)
+   c(clr,rgb(t(col2rgb(rev(clr))*0.6+255*0.4),max=255))
+ }
> micst <-
+ function( Pi, Pc, rv=TRUE, xl=c(0,20), clr="orange", xaxt="n" )
+ {
+   nc <- ncol(Pi)/2
+   clr <- mk.clr( clr )
+   plot( Pi, col=clr, xlim=xl, xlab="", yaxt="n",
+         col.lab="transparent", xaxt=xaxt )
+   lines( as.numeric(rownames(Pi)), Pi[,nc], lwd=4 )
+   Pi <- cbind(0,Pi)
+   axis( side=2, lend=1, lwd=0, lwd.ticks=1, at=1:10/10, labels=NA )
+   axis( side=2, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:10*10/100, tcl=-0.4, labels=NA )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:20*5 /100, tcl=-0.3, labels=NA )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:100 /100, tcl=-0.2, labels=NA )
+
+   plot( Pc, col=clr, xlim=if(rv) xl[2:1] else xl,
+         xlab="", yaxt="n", col.lab="transparent", xaxt=xaxt )
+   lines( as.numeric(rownames(Pc)), Pc[,nc], lwd=4 )
+   axis( side=2+2*rv, lend=1, lwd=0, lwd.ticks=1, at=1:10/10, labels=NA )
+   axis( side=2+2*rv, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+   axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10, if(rv) labels=NA )
+   axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:10*10/100, tcl=-0.4, labels=NA )
+   axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:20*5 /100, tcl=-0.3, labels=NA )
+   axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:100 /100, tcl=-0.2, labels=NA )
+   clr <-< clr
+ }
> legpl <-
+ function( pP, tit, lnam=colnames(pP) )
+ {
+   plot( NA, ylim=c(0,100), xlim=c(0,100), axes=FALSE, ylab="" )
+   nst <- ncol(pP)/2
+   stn <- lnam[1:nst]
+   text( 5, 90, tit, adj=0, font=2, cex=1.5 )
+   text( rep(5,nst), 85-(nst:1)*10, stn, font=2, cex=1.5, adj=0, col=clr[1:nst] )
+ }
```

With these functions in place we can plot cumulative risks in the two groups:

```
> layout( matrix(1:12,4,3,byrow=T), widths=c(2,2,1.3) )
> par( # mfrow=c(4,3),
+      mar=c(1,2.5,0,0), oma=c(3,2,2.5,1), las=1 )
> micst( pAi, pAc, xl=c(0,20), clr="orange" )
> legpl( pAi, "Autonomic neuropathy", lnam=c("Stable","Progression") )
> micst( pPi, pPc, xl=c(0,20), clr="orange" )
> legpl( pPi, "Peripheral neuropathy", lnam=c("Stable","Progression") )
> micst( rEi, rEc, xl=c(0,20), clr=heat.colors(7)[5:1] )
> legpl( rEi, "Retinopathy", lnam=c("None","Minimal","Moderate","pre-Proliferative","Proliferative",
+      micst( Pi, Pc, xl=c(0,20), clr=heat.colors(5)[c(3,1)], xaxt="s" )
> legpl( Pi, "Albuminuria" )
> mtext( c("Intensive","Conventional"), at=c(1,4)/8, side=3, line=1, las=0, outer=TRUE )
> mtext( "Probability", side=2, line=0, las=0, outer=TRUE )
> mtext( "Time since baseline (years)", side=1, line=1.5, outer=TRUE, at=0.42 )
```

5.6 Comparative plotting of microvascular complications

The relevant `pState` objects are:

- Autonomic neuropathy: `pAi`, `pAc`
- Peripheral neuropathy: `pPi`, `pPc`
- Retinopathy: `Ei`, `Ec` — the objects from models that only allows progression of retinopathy, using the empirical baseline distribution as starting point.

The objects `rEi`, `rEc` are from the models that allow improvement of retinopathy, but they are likely to be shaky because of the small number of improvement transitions.

- Albuminuria: `Pi`, `Pc`

First we devise a function that plots state distribution from the two groups together

```
> mk.clr <-
+ function( clr )
+ {
+   clr <- c("forestgreen",clr)
+   c(clr,rgb(t(col2rgb(rev(clr))*0.6+255*0.4),max=255))
+ }
> micst <-
+ function( Pi, Pc, rv=TRUE, xl=c(0,20), clr="orange", xaxt="n" )
+ {
+   nc <- ncol(Pi)/2
+   clr <- mk.clr( clr )
+   plot( Pi, col=clr, xlim=xl, xlab="", yaxt="n",
+         col.lab="transparent", xaxt=xaxt )
+   lines( as.numeric(rownames(Pi)), Pi[,nc], lwd=4 )
+   Pi <- cbind(0,Pi)
+   axis( side=2, lend=1, lwd=0, lwd.ticks=1, at=1:10/10, labels=NA )
+   axis( side=2, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:10*10/100, tcl=-0.4, labels=NA )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:20*5 /100, tcl=-0.3, labels=NA )
+   axis( side=4, lend=1, lwd=0, lwd.ticks=1, at=0:100 /100, tcl=-0.2, labels=NA )
+
+   plot( Pc, col=clr, xlim=if(rv) xl[2:1] else xl,
+         xlab="", yaxt="n", col.lab="transparent", xaxt=xaxt )
```

```

+ lines( as.numeric(rownames(Pc)), Pc[,nc], lwd=4 )
+ axis( side=2+2*rv, lend=1, lwd=0, lwd.ticks=1, at=1:10/10, labels=NA )
+ axis( side=2+2*rv, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10 )
+ axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=((1:5)*2-1)/10, if(rv) labels=NA )
+ axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:10*10/100, tcl=-0.4, labels=NA )
+ axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:20*5 /100, tcl=-0.3, labels=NA )
+ axis( side=4-2*rv, lend=1, lwd=0, lwd.ticks=1, at=0:100 /100, tcl=-0.2, labels=NA )
+ clr <- clr
+ }
> legpl <-
+ function( pP, tit, lnam=colnames(pP) )
+ {
+ plot( NA, ylim=c(0,100), xlim=c(0,100), axes=FALSE, ylab="" )
+ nst <- ncol(pP)/2
+ stn <- lnam[1:nst]
+ text( 5, 90, tit, adj=0, font=2, cex=1.5 )
+ text( rep(5,nst), 85-(nst:1)*10, stn, font=2, cex=1.5, adj=0, col=clr[1:nst] )
+ }

```

With these functions in place we can plot cumulative risks in the two groups:

```

> layout( matrix(1:12,4,3,byrow=T), widths=c(2,2,1.3) )
> par( # mfrow=c(4,3),
+      mar=c(1,2.5,0,0), oma=c(3,2,2.5,1), las=1 )
> micst( pAi, pAc, xl=c(0,20), clr="orange" )
> legpl( pAi, "Autonomic neuropathy", lnam=c("Stable","Progression") )
> micst( pPi, pPc, xl=c(0,20), clr="orange" )
> legpl( pPi, "Peripheral neuropathy", lnam=c("Stable","Progression") )
> micst( rEi, rEc, xl=c(0,20), clr=heat.colors(7)[5:1] )
> legpl( rEi, "Retinopathy", lnam=c("None","Minimal","Moderate","pre-Proliferative","Proliferative",
+ "Abuminuria" )
> micst( Pi, Pc, xl=c(0,20), clr=heat.colors(5)[c(3,1)], xaxt="s" )
> legpl( Pi, "Abuminuria" )
> mtext( c("Intensive","Conventional"), at=c(1,4)/8, side=3, line=1, las=0, outer=TRUE )
> mtext( "Probability", side=2, line=0, las=0, outer=TRUE )
> mtext( "Time since baseline (years)", side=1, line=1.5, outer=TRUE, at=0.42 )

```

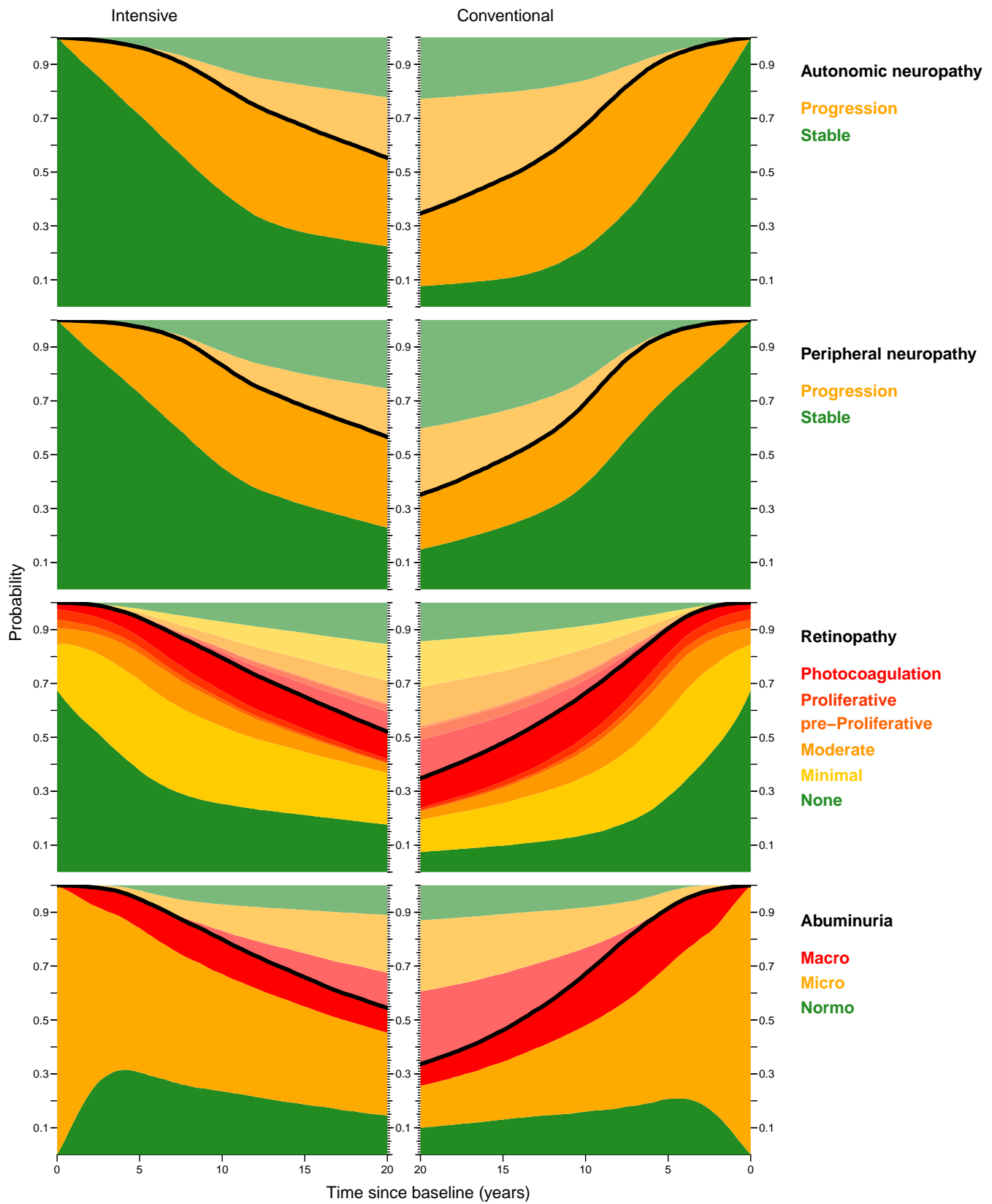


Figure 5.27: Cumulative risks of microvascular complications, controlled for mortality. There is something fishy about the retinopathy — the overall mortality is too high.

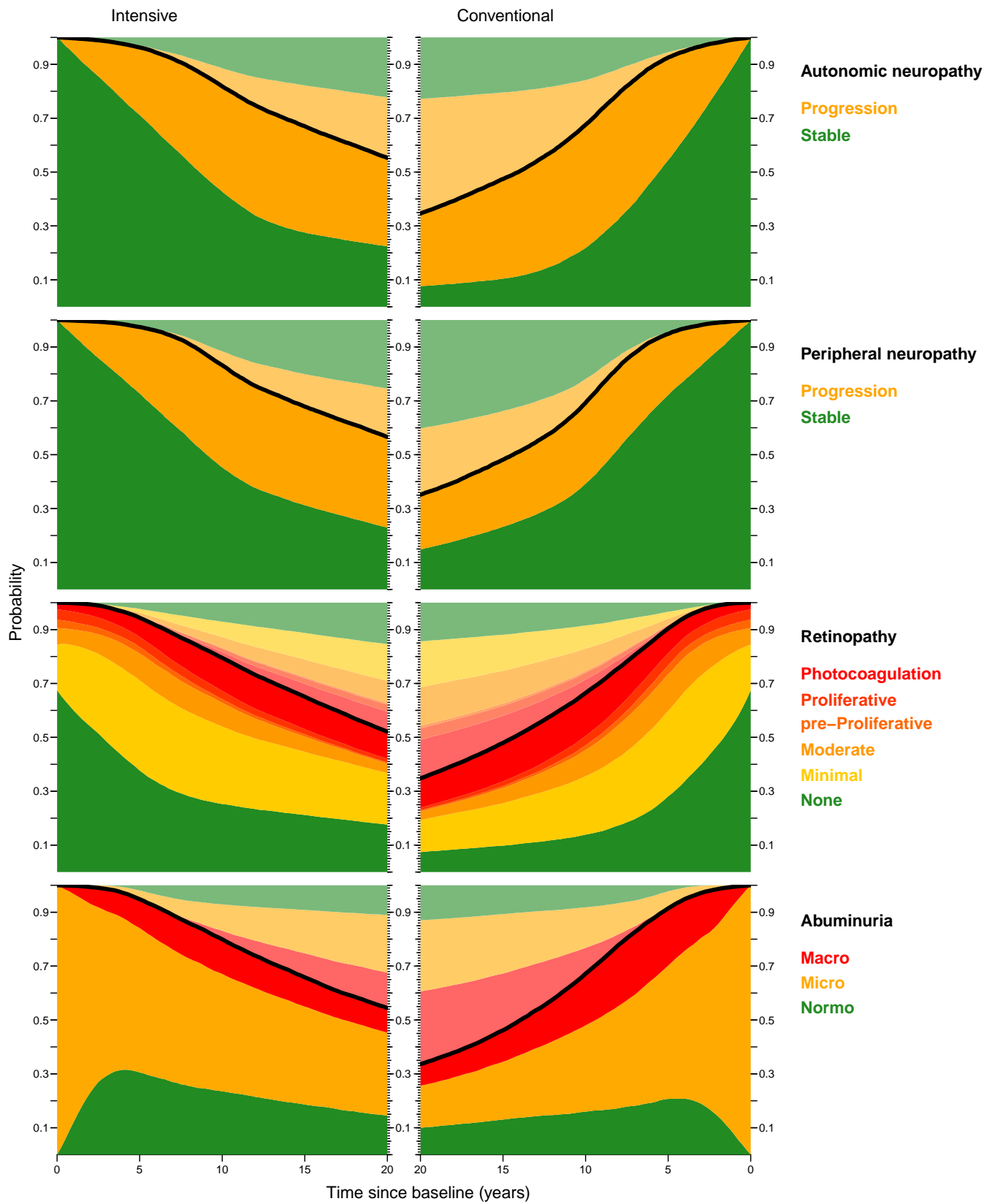


Figure 5.28: Cumulative risks of microvascular complications, controlled for mortality. There is something fishy about the retinopathy — the overall mortality is too high.

5.7 Plotting of summary HRs

```
> dimnames( mainCI )

$outcome
 [1] "All cause mortality"      "CVD mortality"          "non-CVD mortality"
 [4] "Death or 1st CVD"        "Death | CVD state"     "CVD event | CVD state"
 [7] "Retinopathy progression" "Autonomic neuropathy"  "Peripheral neuropathy"
[10] "Macroalbuminuria"

$model
 [1] "raw"      "age/sex"

$what
 [1] "Estimate" "2.5%"      "97.5%"      "P"

> yy <- 13-c(1,3,4,6,7,9:12)
> plotEst( mainCI[-4,"raw",-4], xlog=TRUE, y=yy+0.1, txtpos=yy,
+         vref=1, grid=c(2:10,15,20)/10, xlim=c(0.2,2), lwd=3, cex=1.5,
+         xlab="Hazard ratio - Intensive versus Conventional")
```

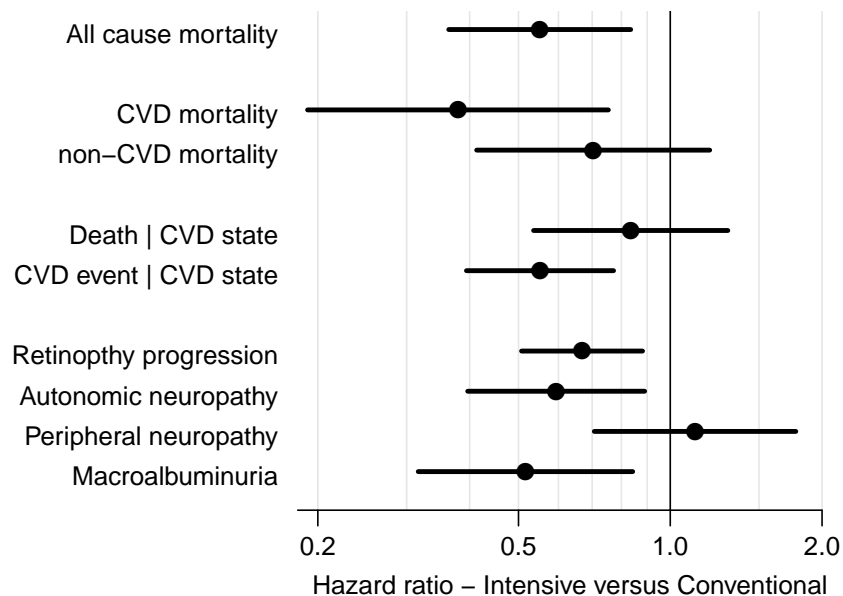


Figure 5.29: *HRs for different outcomes*

```
> fc <- function( x, d=2 ) formatC(x,digits=d,format="f")
> dimnames( mainCI )

$outcome
 [1] "All cause mortality"      "CVD mortality"          "non-CVD mortality"
 [4] "Death or 1st CVD"        "Death | CVD state"     "CVD event | CVD state"
 [7] "Retinopathy progression" "Autonomic neuropathy"  "Peripheral neuropathy"
[10] "Macroalbuminuria"

$model
 [1] "raw"      "age/sex"

$what
 [1] "Estimate" "2.5%"      "97.5%"      "P"
```

```

> yy <- 13-c(1,3,4,6,7,9:12)
> plotEst( mainCI[-4,"raw",-4], xlog=TRUE, y=yy+0.1, txtpos=yy,
+         vref=1, grid=c(2:10,15,20)/10, xlim=c(0.2,5), lwd=3, cex=1.5,
+         xlab="Hazard ratio - Intensive versus Conventional")
> for( i in 1:length(yy) )
+ text( 2.0, yy[i],
+       paste( fc( mainCI[-4,,][i,"raw",1]), " (",
+             fc( mainCI[-4,,][i,"raw",2]), "-",
+             fc( mainCI[-4,,][i,"raw",3]), ")", P=",
+             fc( mainCI[-4,,][i,"raw",4]), 3),
+         sep = "" ),
+       adj = 0, cex=0.8 )

```

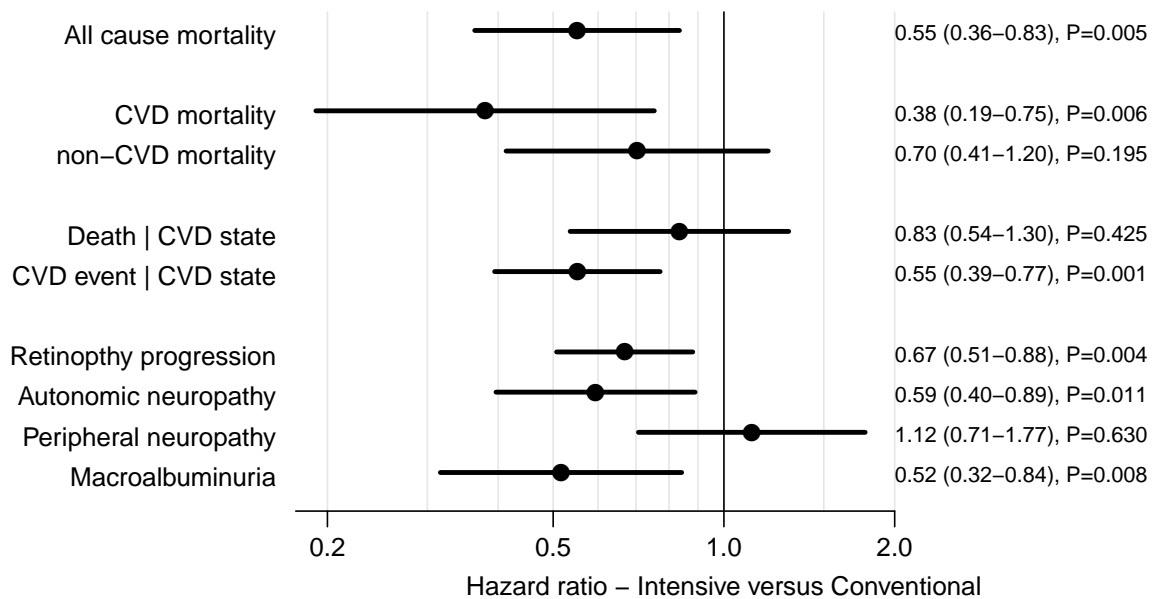


Figure 5.30: *HRs for different outcomes*

References

- [1] P. Gaede, H. Lund-Andersen, H. H. Parving, and O. Pedersen. Effect of a multifactorial intervention on mortality in type 2 diabetes. *N. Engl. J. Med.*, 358(6):580–591, Feb 2008.